Fatty-acid-binding protein as a plasma marker for the estimation of myocardial infarct size in humans

Jan F C Glatz, Appie H Kleine, Frans A van Nieuwenhoven, Wim T Hermens, Marja P van Dieijen-Visser, Ger J van der Vusse

Abstract

Background—There are substantial amounts of cytoplasmic heart-type fatty-acid-binding protein (FABP) (15 kDa) in myocardial tissue. The rapid release of FABP into plasma during ischaemia indicates the possibility of using this protein as a biochemical marker for ischaemic myocardial injury.

Objective—To study the completeness of the release of FABP from damaged tissue in patients with acute myocardial infarction (AMI) and the suitability of serial plasma FABP concentrations for estimation of myocardial infarct size.

Methods—Immunologically assayed FABP and enzymatically assayed creatine kinase isoenzyme MB (CK-MB) and α-hydroxybutyrate dehydrogenase (HBDH) were determined serially in plasma samples from 49 patients with AMI who had been treated with thrombolytic agents within six hours after the onset of AMI. Previously validated circulatory models and a value of 2-6 h⁻¹ for the fractional clearance rate of FABP from plasma were used to calculate cumulative protein release into plasma.

Results—Release of FABP was completed earlier (24-36 h) after AMI than that of CK-MB (50-70 h) and that of HBDH (>70 h). However, infarct size estimated from the cumulative release of the proteins and expressed as gram equivalents of healthy myocardium per litre of plasma yielded a comparable value of 4-6 for both FABP and the two enzymes.

Conclusion—The data indicate that FABP released from the heart after AMI is quantitatively recovered in plasma and that FABP is a useful biochemical plasma marker for the estimation of myocardial infarct size in humans.

Patients and methods

PATIENTS AND BLOOD SAMPLING
We studied 50 patients (six women, 44 men; age 29-71) with chest pain and ST segment elevation typical of AMI. Patients were eligible for this study if they were admitted to the coronary care unit of the hospital within six hours after the first onset of infarct-related symptoms. Patients were participating in a pilot study of treatment with a new inhibitor of platelet function by selective blockade of thromboxane A₂ synthetase (Ridogrel, Janssen Pharmaceutica, Beerse, Belgium) and were also treated with alteplase (Boehringer Ingelheim, Germany) and heparin. Coronary angiography, performed at 90 minutes after start of treatment, showed successful reperfusion in 40 patients. Subsequent percutaneous transluminal coronary angioplasty (PTCA), in the remaining cases also showed reperfusion. Ischaemia was located anteriorly in 13 patients and inferioposteriorly in 37 patients.
None of the patients had cardioversion. Plasma samples were incomplete for one patient who died: the results of the remaining 49 patients are shown. Further details on patient selection and clinical treatment are described elsewhere.\(^{20}\)

Blood samples were taken upon admission to the hospital (2.8–1 h, range 1–6 h) after the onset of symptoms mean (SD) for \(n = 49\) and 3, 6, 9, 12, 24, 36, 48, 72, and (for 35 patients only) 96 h thereafter, resulting in nine or 10 samples per patient. Samples were collected in glass tubes containing dry heparin. After routine centrifugation plasma was stored in several aliquots at \(-20^\circ\text{C}\) until analysis.

**ANALYTICAL TECHNIQUES**

FABP was measured in plasma and tissue samples by a sensitive non-competitive enzyme-linked immunosorbent assay of the antigen capture type (sandwich ELISA) as described elsewhere.\(^{18}\) Samples were diluted with phosphate buffered saline (pH 7.4) containing 0.1% bovine serum albumin and 0.05% Tween-20. The detection limit of the assay was 0.5 \(\mu\text{g}\)/l. The recovery (mean (SD)) of purified human heart FABP added in various quantities to control human plasma was 94 (12)% (\(n = 11\)) and the interassay coefficient of variation was 6.5%.

The activities of creatine kinase isoenzyme MB (CK-MB) and HBDH were measured spectrophotometrically at 25\(^\circ\text{C}\) in a centrifugal analyser (Cobas Bio System, Hoffmann La Roche, Basel, Switzerland) with commercially available test kits. For CK-MB we used an enzyme assay kit that is based on immunoinhibition of the predominant M unit in creatine kinase (Boehringer Mannheim, Germany). For HBDH, which reflects mainly the activity of lactate dehydrogenase isoenzyme-1, we used an assay kit with 2-oxobutyrate as the substrate (Boehringer Mannheim, Germany). Activities are expressed in \(\mu\text{mol}\) substrate converted per minute (units) per litre of plasma.

**PLASMA REFERENCE VALUES**

Reference values (upper normal concentration or activity) for FABP, CK-MB, and HBDH in plasma were estimated in non-haemolytic blood samples obtained from 72 healthy blood donors. Mean (SD) plasma FABP concentration was 9 (5) \(\mu\text{g}\)/l, CK-MB activity 4 (3)U/l, and HBDH activity 90 (35)U/l. The reference values (mean plasma concentration or activity plus twice the standard deviation) were 19 \(\mu\text{g}\)/l for FABP, 10 U/l for CK-MB, and 160 U/l for HBDH.

**CALCULATION OF CUMULATIVE PROTEIN RELEASE**

Cumulative release of cardiac proteins from the onset of AMI (\(t = 0\)) up to time \(t\), \(Q(t)\), was calculated for a two-compartment model\(^{21}\) as follows:

\[
Q(t) = C(0) + \int_0^t \text{TER} \cdot \exp\left[\text{ERR}(t-0)\right] \text{C}(t) \, dt + \text{FCR} \cdot \text{C}(t) \, dt
\]

where the three terms are the actual protein concentration (or enzyme activity) in plasma, the extravascular concentration, and the amount of protein eliminated from plasma, respectively, each expressed per litre of plasma. TER, ERR, and FCR are the fractional rate constants for transcapillary escape, extravascular return, and catabolism (elimination) of protein, respectively.

For CK-MB the values used were: \(\text{TER} = 0.014 \text{ h}^{-1}\), \(\text{ERR} = 0.018 \text{ h}^{-1}\), and \(\text{FCR} = 0.34 \text{ h}^{-1}\).\(^{21}\) For HBDH the values for TER and ERR were equal to those of CK-MB, whereas FCR was 0.015 \(\text{h}^{-1}\).\(^{21}\)

Calculation of the cumulative release of FABP was hampered by the fact that its fractional clearance rate is not known. However, turnover studies in humans of other small proteins such as myoglobin (17–2 kDa), lysozyme (14 kDa)\(^{24}\) and retinol binding protein (21 kDa)\(^{25}\) all showed elimination from plasma predominantly by rapid renal clearance with a half life of 10–20 min. Hence the cumulative release of myoglobin into plasma after AMI was calculated in a one-compartment (that is plasma volume) model\(^{22,26}\) where the second term in the above formula was left out, so that the cumulative protein release equals the integrated plasma curve, multiplied by FCR. Because the mean half life of myoglobin in AMI patients is 16 min\(^{21}\) the FCR amounts to \((\ln 2)/t_1/2\) = 2.6 \(\text{h}^{-1}\). This approach was validated in a separate set of 10 patients with clinically confirmed AMI in whom plasma samples were taken every hour (fig 1). The ratio of the plasma concentrations of myoglobin and FABP remained constant throughout the period that plasma values were raised. The mean plasma myoglobin: FABP ratio differed less than 16% from the plasma reference ratio of 5:1. In addition a one-compartment model for circulating FABP was validated in dogs.\(^{27}\)

For each patient measured plasma FABP concentrations and enzyme activities were expressed by subtraction of the normal steady-state values. For this we used the respective plasma reference values (give above) or the concentration or activity measured in the first plasma sample taken from the patient when this value was lower than the reference value.

---

**Figure 1** Plasma myoglobin: FABP ratio as a function of time after AMI in 10 patients from whom hourly plasma samples were obtained (mean (SEM)). AMI, acute myocardial infarction; FABP, fatty-acid-binding protein.
Inclusion of patients who presented up to six hours after onset of pain introduced an error into the calculation of the cumulative release of FABP because it ignored prior release. However, only one patient entered the study more than 4-8 h after the onset of AMI, and in the absence of thrombolysis-induced reperfusion the release of cardiac proteins up to 4-8 h is relatively small.

**MYOCARDIAL INFARCT SIZE**

The cumulative release of each cardiac protein per litre of plasma was divided by the myocardial content of the specific protein per gram of wet weight tissue, so that myocardial injury could be expressed as gram equivalents of healthy heart muscle per litre of plasma. The myocardial content of FABP was measured in parts of intact human heart (left ventricular tissue) obtained from either the De Weyer Hospital in Heerlen or the Academic Hospital Maastricht after necropsy (performed within 12 hours after death) on patients who died from non-cardiac causes. FABP content, assayed by the same method as used for plasma, was 0.56 (±0.07) mg/g wet weight (mean (SD) for 17 individuals). Regional and transmural differences in the left ventricle were not significant (data not shown). Myocardial enzyme content, measured under the same assay conditions as plasma in the present study, were 132 U/g for CK-MB and 123 U/g for HBDH.

**VALIDATION OF SAMPLING PROTOCOL**

We validated the present sampling protocol for calculation of the cumulative release of FABP in the 10 patients (one woman, nine men; age 45–75) referred to earlier, in whom hourly plasma samples were obtained up to 12 hours after admission to hospital. (About 80% of total FABP release is completed at this time). The cumulative protein release calculated on the basis of samples obtained at zero, three, six, nine and 12 hours after admission to hospital correlated significantly with the release calculated on the basis of all 13 samples (fig 2) (r = 0.97, p < 0.05), indicating that the less frequent sampling protocol will yield good estimates of cumulative protein release.

**STATISTICAL ANALYSIS**

The release curves for proteins into plasma are presented as means (SEM). Statistical analysis of differences (between groups) was performed with Student’s t-test. The level of significance was set at p < 0.05.

**Results**

Mean plasma concentration or activities of the three proteins examined as a function of time for 49 patients (fig 3) showed a large difference between the plasma kinetics of FABP and those of CK-MB and HBDH. The peak plasma concentration FABP was reached 5–7 (1–4) h after AMI, whereas that of CK-MB was reached 11–7 (4–4) h after AMI and that of HBDH 28–2 (13–5) h after AMI (mean (SD), n = 49). Within 24 hours the plasma concentration of FABP had returned to normal, whereas CK-MB took 50–70 hours and HBDH more than 70 hours (fig 3).

In one patient a recurrent myocardial infarction developed soon (<10 h) after the initial AMI. The appearance of this recurrent infarction is reflected clearly in the plasma curve for FABP but is less apparent from the CK-MB and HBDH plasma curves (fig 4).

The cumulative release patterns of the three proteins, expressed in gram equivalents of tissue per litre of plasma, also show a difference between FABP and CK-MB and HBDH

![Figure 2](https://example.com/figure2.png)

**Figure 2** Relation between the cumulative release over 14 hours (Q14) of FABP into plasma as calculated from plasma concentrations from 5 (3-hourly) or 13 (hourly) blood samples. The line of identity is given (dotted line). The calculated regression line (solid line) is y = 0.96 x + 0.07, FABP, fatty acid-binding protein.

![Figure 3](https://example.com/figure3.png)

**Figure 3** Plasma concentration of FABP (× 5) and plasma activities of CK-MB (× 10) and HBDH as a function of time after AMI in forty-nine patients (mean (SEM)). AMI, acute myocardial infarction; CK-MB, creatine kinase isoenzyme-MB; FABP, fatty acid-binding protein; HBDH, hydroxybutyrate dehydrogenase.

![Figure 4](https://example.com/figure4.png)

**Figure 4** Plasma concentration of FABP (× 5) and plasma activities of CK-MB (× 10) and HBDH as a function of time after initial AMI in a patient in whom AMI recurred. AMI, acute myocardial infarction; CK-MB, creatine kinase isoenzyme-MB; FABP, fatty acid-binding protein; HBDH, hydroxybutyrate dehydrogenase.
The conclusion that FABP, like myoglobin, is eliminated from plasma predominantly by rapid renal clearance is supported by the reported detection of FABP in urine samples collected as early as two hours after onset of chest pain.19

RECOVERY OF FABP IN PLASMA AFTER ISCHAEMIC MYOCARDIAL DAMAGE

The total quantities of CK-MB and HBDH depleted from dog hearts after coronary occlusion equal the calculated release of these proteins into plasma.38 The recovery of FABP in plasma after ischaemic myocardial injury has not yet been measured in experimental studies. In view of the stability of CK-MB and HBDH29 it is generally assumed that in humans too these cardiac proteins are quantitatively recovered in plasma after AMI. The fact that in the present study the estimate of infarct size (expressed in gram equivalents of healthy myocardium) based on cumulative release of FABP resemble estimates based on cumulative release of CK-MB or HBDH suggests that the recovery in plasma of FABP depleted from the heart is also essentially complete.

Release of FABP was completed earlier than that of the two enzymatic markers. The relative delay in the appearance of the enzymes in plasma may relate either to a slower release of enzymes from damaged myocardial cells or a slower transport from the interstitial space to the vascular space by lymph drainage and by increased endothelial transport in the diseased tissue. There is evidence of binding of cytoplasmic creatine kinase to structural elements in heart muscle.33 However, a delay in the interstitial or transendothelial transport of enzymes appears most likely, because experimental studies with isolated rat hearts subjected to low-flow ischaemia and reperfusion showed no differences in the release pattern of various cardiac proteins.14 Because CK-MB (80 kDa) and HBDH (130 kDa) are much larger than FABP (15-0 kDa) it is tempting to suggest that molecular size may well be a main determinant of the rate of protein transport from the interstitial space to the plasma space.

ESTIMATION OF INFARCT SIZE

Estimates of infarct size 72 h after the onset of AMI based on the various markers were not significantly different (fig 5). However, infarct size calculated from the cumulative release of FABP tended to be higher. This tendency may have been caused by an underestimation of the myocardial content of FABP, but the presently measured value (0.56 mg/g) was assayed by the same method as used for plasma and is similar to that published by others (0.6 mg/g).27 The difference may also be due to an overestimation of the fractional clearance rate of FABP, which was taken to equal that of myoglobin. The difference in the isoelectric points of FABP (pI 5.1) and myoglobin (pI 7.0) means that at physiological pH FABP is more negatively charged, causing FABP to be cleared less rapidly by

Discussion

The rationale for using fatty-acid-binding protein (FABP) as a plasma marker for myocardial injury is based on this soluble protein being present in the myocardium in large amounts and its virtual confinement to the cytoplasmic space. FABP has also been detected in the matrix of bovine heart mitochondria,30 31 but total mitochondrial FABP represents less than 1% of the cellular content.30 Earlier studies18 19 showed that in humans plasma FABP concentrations increased significantly with ischaemic myocardial injury and, hence, that FABP was a useful qualitative index for the assessment or exclusion of acute myocardial infarction. In the present study we found that the release of FABP from damaged tissue was essentially complete within 36 hours after the onset of AMI and that the cumulative release of FABP into plasma can be used to estimate myocardial infarct size.

PLASMA KINETICS OF FABP

For the calculation of cumulative release we assumed that the plasma kinetics of FABP were similar to those of myoglobin. This implies that the elimination of these proteins from plasma can be described with a one-compartment (that is, a plasma volume) model, because their plasma kinetics are dominated entirely by the high rate of protein elimination from plasma so that extravasation can be neglected.21 Even if there were significant extravasation of FABP, this would affect only the shape of the release curve and, provided that a full curve is recorded, not the cumulative release over time, which is the variable of interest.

Figure 5 Cumulative release of FABP, CK-MB, and HBDH in plasma after AMI in forty-nine patients. Data are expressed in gram equivalents of healthy myocardium per litre of plasma (mean (SEM)). *Cumulative release of FABP significantly higher (p < 0.05) than that of both CK-MB and HBDH. AMI, acute myocardial infarction; CK-MB, creatine kinase isoenzyme-MB; FABP, fatty-acid-binding protein; HBDH, hydroxybutyrate dehydrogenase.
the kidneys and hence to stay longer in the circulation than myoglobin. This would lead to a lower fractional clearance rate and in turn to lower values of calculated infarct size.

Alternatively, infarct sizes calculated from the cumulative release of CK-MB and HBDH in plasma might have been underestimated because these enzymes were measured as activity whereas FABP was measured by an immunochemoassay (that is, as protein mass). Measurement of CK-MB in plasma as protein mass was more reliable than measurement as activity.

Despite these uncertainties we conclude that, provided frequent blood samples are taken, FABP gives a clinically useful estimate of myocardial infarct size, because 72 h after the onset of AMI we found no significant differences between the estimates based on the various markers.

CLINICAL APPLICATION
As a plasma marker for ischaemic myocardial injury, FABP shares several characteristics with other cardiac proteins (myoglobin, troponin T) and with cardiосpecific enzymes (CK-MB, HBDH), but it also shows some unique differences that will enhance the detection and evaluation of an acute myocardial infarction. Like myoglobin, the rapid appearance of FABP in plasma after tissue damage permits the early assessment of exclusion of AMI as well as the immunohistochemical confirmation of very recent myocardial infarction. This application is also enhanced by the relatively rapid elimination of FABP from plasma because this keeps the steady state plasma concentration of this protein at a low level. A further example of this concept is the fact that the ratio of the cytoplasmic to the vascular concentration of FABP (amounting to about $2 \times 10^7$) is one order of magnitude higher than that of any of the cardiac enzymes (CK-MB, 6 $\times 10^4$; HBDH, 5 $\times 10^4$). The consequence of such a high ratio is that the release of only minute amounts of FABP from myocardial cells with significantly raise its plasma concentration, making FABP a diagnostic marker with high sensitivity. Furthermore, the rapid clearance of FABP also allows recurrent infarctions to be more easily identified.

Despite the occurrence of various types of FABP, with some types found solely in a single tissue—such as intestinal FABP in interstitial epithelial cells—heart-type FABP is found not only in cardiac muscle but, like myoglobin, also in striated skeletal muscle. However, the myoglobin:FABP ratio of human skeletal muscle is 32–70 (depending on the muscle fibre type composition) and that of heart 4–9 (1–2), making FABP more cardiосpecific than myoglobin. Because FABP and myoglobin show similar patterns of release from tissue and of elimination from plasma, the plasma myoglobin:FABP ratio will allow discrimination between cardiac and skeletal muscle damage. Another condition that may cause erroneous values of plasma FABP (and of myoglobin) is a decreased glomerular filtration rate (GFR). In such cases the FABP released from damaged myocardial tissue will accumulate in the plasma, thus leading to a possible overestimation of infarct size.

The rapid release of FABP into plasma makes possible a reliable measure of myocardial infarct size within 24 hours of AMI. However, this application must meet two conditions. First, sufficient plasma samples have to be obtained during the first day of hospital admission. Secondly, the use of the FABP plasma concentration as an early diagnostic tool of AMI requires a fast assay system. This is not yet available (the sandwich ELISA used in the present study takes a few hours to complete), but recent developments with immunodiagnostic tests should lead to a sensitive assay for plasma FABP that will give quantitative data within minutes. In addition, immunochemo detection of biochemical plasma markers is generally considered to have the advantage of being free from problems inherent to the measurement of enzyme activity of proteins.

We thank Prof Dr M L Simoons, Department of Cardiology and Thoraxcenter, University Hospital Dijkzigt, Rotterdam, The Netherlands, for his interest and valuable advice. This study was supported by StiP, Executive Agency for Technology Policy (MTR 88002). J F C G is an Established Investigator of the Netherlands Heart Foundation.

14 Knowlton AA, Apstein CS, Saeed R, Brecher P, Laskey
140
of heart fatty acid-binding protein with ischemia and
15 Knowlton AA, Burrier RE, Brecher P. Rabbit heart fatty
acid-binding protein. Isolation, characterization, and
16 Glatz JFC, Van der Vusse GJ. Cellular fatty acid-binding
proteins: Current concepts and future directions. Mol
17 Veerkamp J, Peeters RA, Maatman RGHJ. Structural and
functional features of different types of cytoplasmic
fatty acid-binding proteins. Biochim Biophys Acta 1991;
1081:1–24.
18 Kleine AH, Glatz JFC, Van Nieuwenhoven FA, Van der
Vusse GJ. Release of heart fatty acid-binding protein
into plasma after acute myocardial infarction in man.
19 Tanaka T, Hirota Y, Sohmiya K-I, Nishimura S,
Kawamura K. Serum and urinary heart fatty acid-binding
protein in acute myocardial infarction. Clin Biochem
20 Van der Wurken LR, Nijssen KM, Siooms ML. Ridgedorl
as an adjunct to thrombolysis in acute myocardial infarc-
21 Willems GM, Visser MP, Krill MTA, Hermens WTh.
Quantitative analysis of plasma enzyme levels based on
simultaneous determination of different enzymes.
22 Sylven C. The kinetics of myoglobin in old volunteers
and in patients with acute myocardial infarction. Scand J
23 Grotth S, Sylven C. Myoglobin kinetics in patients suffering
from acute myocardial infarction in its early phase—as
studied by the single injection method. Scand J Clin
Lab Invest 1981;41:79–85.
24 Hansen NE, Karle H, Andersen V, Ologaard K. Lysosome
25 Vahlquist A, Peterson PA, Wibell L. Metabolism of the
vitamin A transporting protein complex. Eur J Clin
26 Ellis AK, Saran BR. Kinetics of myoglobin release and
prediction of myocardial depletion after coronary artery
27 Tsujii R, Tanaka T, Somiya K, Yoshimoto K, Hirota Y,
Kawamura K. Canine heart fatty acid-binding protein
(ch-FABP). Purification and kinetic studies [abstract].
J Mol Cell Cardiol 1992;24:S183.
28 Hermens WTh, Willems GM, Davids HA, Hollaar L, Van
der Laarse A. Enzymatic assessment of myocardial
injury after infarction or cardiac surgery. Is enzyme
29 Van der Laarse A, Dijkstraum NJ, Hollaar L, Caspers T.
The in-vitro enzymatic activities of lactate dehydrogenase,
aldehydehydroxybutyrate dehydrogenase, creatine kinase
and aspartate aminotransferase in human myocardial
biopsies and autopsies. Clin Chim Acta 1980;104:
381–91.
30 Börchers T, Unterberg C, Rödel H, Robenek H, Spener F.
Subcellular distribution of cardiac fatty acid-binding
protein in bovine heart muscle and quantitation with an
enzyme-linked immunosorbent assay. Biochim Biophys
31 Unterberg C, Börchers T, Hojrup P, Roepstorff P,
Knutsen J, Spener F. Cardiac fatty acid-binding
proteins: Isolation and characterization of the mitochondrial
fatty acid-binding protein and its structural relationship
with the cytosolic isoforms. J Biol Chem 1990;265:
16235–61.
32 Hermens WT, Van der Veen FH, Willems GM, Mullers-
Boumans ML, Schijvers-Van Schendel A, Reneman
RS. Complete recovery in plasma of enzymes lost from
the heart after permanent coronary occlusion in the dog.
33 Ortauwyn JH. Evidence for binding of cytoplasmic creatine
kinase to structural elements in heart muscle. Nature
34 Vork MM, Glatz JFC, Surfet DAM, Van der Vusse GJ.
Protein release from isolated rat heart during normoxia,
low-flow ischemia and reperfusion. Can J Physiol
Pharmacol (in press).
35 Paulussen RJA, Van Moerkerk HTB, Veerkamp JH.
Immunochromatographic quantitation of fatty acid-binding
proteins. Tissue distribution of liver and heart FABP types
in human and porcine tissues. Int J Biochem 1990;
22:393–8.
36 Guyton AC. Glomerular filtration and glomerular filtrate.
37 Eisenberg PR, Shaw D, Saba C, Jaffe AS. Concordance of
creatine kinase-MB activity and mass. Clin Chim
38 Kleine AH, Glatz JFC, Havenith MG, Van Nieuwenhoven
FA, Van der Vusse GJ, Bosman FT. Immunohisto-
chromic detection of very recent myocardial infarctions
in humans with antibodies against heart-type fatty
39 Sacchettini JC, Hauff SM, Van Camp SL, Cistola DP,
Gordon JJ. Development and structural studies of an
intracellular lipid binding protein in the ileal epithelium.
40 Kanda T, Nakatomi Y, Ishiiwa H, Hitomi M, Matsubara
Y, Ono T, et al. Intestinal fatty acid-binding protein as a
sensitive marker of intestinal ischemia. Digest Dis Sci
41 Peeters RA, Veerkamp JH, Van Kessel AG, Kanda T, Ono
T. Cloning of the cDNA encoding human skeletal muscle
fatty acid-binding protein, its peptide sequence and
42 Van Nieuwenhoven FA, Kleine AH, Keizer HA, Van
diejen-Visser MP, Van der Vusse GJ, Glatz JFC.
Comparison of myoglobin and fatty acid-binding protein
as plasma markers for muscle damage in man [abstract].
43 Newman JD, Turner APF. Biosensors: principles and
Fatty-acid-binding protein as a plasma marker for the estimation of myocardial infarct size in humans.


Br Heart J 1994 71: 135-140
doi: 10.1136/hrt.71.2.135

Updated information and services can be found at:
http://heart.bmj.com/content/71/2/135

These include:

Email alerting service
Receive free email alerts when new articles cite this article. Sign up in the box at the top right corner of the online article.

Notes

To request permissions go to:
http://group.bmj.com/group/rights-licensing/permissions

To order reprints go to:
http://journals.bmj.com/cgi/reprintform

To subscribe to BMJ go to:
http://group.bmj.com/subscribe/