No evidence to implicate *Borrelia burgdorferi* in the pathogenesis of dilated cardiomyopathy in the United Kingdom

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Abstract

**Objective**—To determine whether *Borrelia burgdorferi* is implicated in the pathogenesis of dilated cardiomyopathy in the United Kingdom.

**Design**—A controlled prospective study. Patients’ notes were reviewed for evidence of Lyme disease and serum samples were tested by enzyme linked immunoabsorbent assay (ELISA) for antibodies to *B burgdorferi*. Samples with raised antibody concentrations were subsequently analysed by immunoblotting to determine their antibody binding specificity.

**Setting**—Tertiary referral centre.

**Patients**—97 consecutive patients with dilated cardiomyopathy diagnosed according to World Health Organisation criteria were studied. Serum samples were taken from two matched control groups. The first group (n = 38) was age, sex, and geographically matched. The second control group (n = 39) was environmentally matched and consisted of members of the patients’ own households.

**Main outcome measures**—Clinical evidence of Lyme disease. Presence of raised antibody concentrations to *B burgdorferi*.

**Results**—No patients had a previous illness compatible with Lyme disease. Analysis of the ELISA data showed eight of 97 patients with dilated cardiomyopathy (8.2%) and two of 77 controls (3.9%) had raised antibody concentrations. Immunoblot analysis, however, did not show binding patterns consistent with the presence of IgG specific for *B burgdorferi* in any of these samples.

**Conclusions**—There was no clinical or serological evidence to implicate *B burgdorferi* in the pathogenesis of idiopathic dilated cardiomyopathy in the United Kingdom. In the absence of specific symptoms or likely exposure to *B burgdorferi* routine serological testing for Lyme disease in this group of patients is not recommended. Furthermore, raised antibodies to *B burgdorferi* are not diagnostic of active infection and ELISA results should be interpreted with caution unless specific *B burgdorferi* antibody bands have been found by immunoblot analysis.

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of infection and reported improvement after antibiotic treatment in eight out of nine of these patients;15 however they failed to mention the duration of dilated cardiomyopathy in their study and the findings should be interpreted with caution as raised antibody titres are not diagnostic of active infection.15

As *B burgdorferi* has been implicated in the pathogenesis of dilated cardiomyopathy and cardiac manifestations may be the sole presentation of Lyme disease we considered it important to determine whether there was any clinical or serological evidence for infection with *B burgdorferi* in patients from the United Kingdom with idiopathic dilated cardiomyopathy.

**Patients and methods**

**PATIENTS WITH DILATED CARDIOMYOPATHY**

The study group consisted of 97 consecutive patients with dilated cardiomyopathy (mean range) age 43 (12–74), 71 male) referred to our centre from several locations: Greater London (n = 38), south east counties (n = 31), Hampshire (n = 5), Wales (n = 7), west country (n = 5), Italy and Greece (n = 5), Yorkshire (n = 4), and Jersey (n = 2). None of the patients were at occupational risk of exposure to tick bites—for example, farmers, forestry workers, and park workers. The clinical diagnosis of dilated cardiomyopathy was made according to World Health Organisation criteria,14 and all patients underwent cardiac catheterisation and right ventricular endomyocardial biopsy. Patients had been symptomatic for a mean (SD) duration of 34 (48) months, 30 (31%) described an acute viral illness at disease onset, and 13 (14%) had either clinical (n = 7) or histological (n = 6) evidence of myocarditis.15

**CONTROLS**

Serum samples were obtained from two matched control groups. The first control group (n = 38; mean range) age 45 (14–72), 34 male) was age, sex, and geographically matched by selecting the next person on the patient’s own general practitioner’s list with the same sex and within five years of the patient’s age. Subjects of the second control group (n = 39; mean range) age 42 (7–68), 12 male) were environmentally matched and each consisted of a member of the patient’s own household, who was usually the spouse (n = 32), but occasionally a parent (n = 5), or child (n = 2).

**NORMAL POPULATION**

Serum samples from healthy blood donors (n = 50; mean range age 45, (20–70), 25 male) was used to determine the normal range for IgG *B burgdorferi* antibodies in a non-susceptible population.

**Methods**

The patients’ notes were reviewed for clinical evidence of Lyme disease. Antibodies to *B burgdorferi* were detected by ELISA with sonicated whole cell spirochaetes (strain B31) as antigen, and samples with raised antibody concentrations were analysed by immunoblot to further define antibody specificity.

**Enzyme linked immunosorbent assay**

Half of 96-well Immulon 1 immunoassay plates (Dynatech) were coated with 50 µl of a 5 µg/ml suspension of sonicated spirochaetes in carbonate buffer (pH 9.6) by incubation overnight at 4°C. The plates were subsequently blocked with 3% bovine serum albumin in phosphate buffered saline for one hour at 37°C. Serum samples were diluted one in 800 in phosphate buffered saline containing 0.05% Tween-80 (BDH), 3% goat serum, and 1% bovine serum albumin. Each sample (100 µl) was added in triplicate to both the antigen coated wells and uncoated control wells and incubated for one hour at 37°C. The same sample from a patient with raised antibody concentrations (positive control) was also added in triplicate to each plate. After washing, the wells were further incubated for one hour at 37°C with goat antihuman IgG-alkaline phosphatase conjugate (Sigma Chemicals). Antibody binding was detected with p-nitrophenyl phosphate substrate (Sigma Chemicals) and the optical density was read at 405 nm. Background binding to the uncoated control wells was subtracted from the readings and results were expressed as a ratio of the mean optical density of the test sample to the positive control.

**Immunoblot analysis**

*Borrelia burgdorferi* proteins (strain B31) were separated on a 10% polyacrylamide gel as described elsewhere,16 and were transferred on to a nitrocellulose membrane by semi-dry blotting. Excess binding sites were blocked by incubation in 5% dried skimmed milk in phosphate buffered saline. Strips cut from the nitrocellulose were incubated for one hour at 37°C in serum diluted one in 500 in phosphate buffered saline containing 5% dried skimmed milk and 0.2% Tween-80. After washing, the strips were incubated for one hour at 37°C with biotinylated goat antihuman IgG (Amersham) and for a further hour in streptavidin horseradish peroxidase (Amersham), both were diluted one in 1000 with phosphate buffered saline-Tween, and antibody binding was subsequently detected by the enhanced chemiluminescence system (Amersham). Binding of antibody to any of 14 protein bands with molecular weights of 17, 22, 29, 31, 34, 39, 41, 46, 55, 60, 66, 75, 83, 94 kilodaltons (kDa) previously reported to be associated with *B burgdorferi*,17 was assessed blindly by an experienced and independent observer.

**Detection of antibodies to Treponema pallidum**

Samples with raised *B burgdorferi* antibody concentrations by ELISA (n = 11) were screened for binding to *T pallidum* by the *T pallidum* haemagglutination (TPHA) test and the venereal disease reference laboratory (VDRL) test.

**Statistical analysis**

A serum sample was defined as positive for antibodies to *B burgdorferi* by ELISA if the
optical density ratio was greater than the mean optical density ratio + 2 SDs of the normal population. A χ² probability test with Yate's correction was used to compare differences in the number of positive samples in the diluted cardiomyopathy and control groups.

**Results**

**CLINICAL**

On reviewing the notes, none of the patients had either a documented rash consistent with erythema migrans or a previous illness compatible with Lyme disease.

**DETECTION OF ANTIBODIES**

Analysis of the ELISA data showed the presence of raised antibodies to *B burgdorferi* in eight of 97 (8.2%) patients with dilated cardiomyopathy, two of 38 (5.3%) geographically matched controls, and one of 39 (2.6%) environmentally matched controls. There was no significant difference between the dilated cardiomyopathy group and either the geographically matched controls (χ² = 0·03, p = 0·8), environmentally matched controls (χ² = 0·6, p = 0·4), or the control group overall (χ² = 0·736; p = 0·4). Immunoblot analysis showed only very weak antibody binding to the 60 kDa protein in one patient with dilated cardiomyopathy, to the 17kDa protein in a further patient and 17 and 20 kDa in one other. One of the controls had very weak binding to 30 and 41 kDa proteins and another to the 41 kDa protein. These patterns of antibody binding with weak reactivity do not indicate active infection with *B burgdorferi* and most likely result from the presence of cross reacting antibodies.

**Antibodies to Treponema pallidum**

One sample was positive in the TPHA test and no samples tested were positive in the VDRL test.

**Discussion**

It is likely from the evidence reviewed that *B burgdorferi* is the cause of some cases of dilated cardiomyopathy in western Europe. In this study we have investigated, for the first time in the United Kingdom, patients with idiopathic dilated cardiomyopathy for clinical and serological evidence of infection with *B burgdorferi*. The group studied were referred to a tertiary referral centre in southern England, including 17 patients (18%) from regions (Hampshire, Wales, Norfolk, and the west country) in which ticks infected with *B burgdorferi* have been identified or cases of Lyme disease reported. Furthermore, patients not resident in these areas, may have visited endemic areas for Lyme disease within this country, Europe, or North America. Of note, none of the patients with dilated cardiomyopathy were at occupational risk of exposure to tick bites.

In this study group, we found no clinical evidence of Lyme disease and although 8-2% of the patients with dilated cardiomyopathy tested had raised serum *B burgdorferi* antibody concentrations on ELISA, this was not significantly more than in the control groups and subsequent immunoblot analysis did not show antibody binding patterns consistent with *B burgdorferi* infection in any of the samples. This lack of specificity with ELISA is consistent with other published data. False positive antibody concentrations to *T pallidum*, a potentially cross reactive infection only occurred in one patient.

In conclusion, from this group of patients (some of whom live in or near endemic areas for Lyme disease), we have found no evidence to implicate *B burgdorferi* in the pathogenesis of idiopathic dilated cardiomyopathy in the United Kingdom. In the absence of specific symptoms or likely exposure to *B burgdorferi* we would not recommend routine serological testing for Lyme disease in these patients. Our data also confirm that ELISA results should be interpreted with caution unless specific *B burgdorferi* antibody bands have been found on immunoblot analysis. Furthermore, raised antibodies to *B burgdorferi* are not necessarily indicative of active cardiac infection, which can only be diagnosed with certainty by the presence of the spirochaetes in endomyocardial biopsies by culture or detection of *B burgdorferi* DNA by the polymerase chain reaction.

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