How often are angiotensin II and aldosterone concentrations raised during chronic ACE inhibitor treatment in cardiac failure?

R J MacFadyen, A F C Lee, J J Morton, S D Pringle, A D Struthers

Abstract

Objective—Angiotensin II (AII) and aldosterone are not always fully suppressed during chronic angiotensin converting enzyme (ACE) inhibitor treatment. In congestive heart failure (CHF) such failure of hormonal suppression is associated with increased mortality. This study examined how common AII and aldosterone increases are observed during routine clinical practice.

Patients and methods—91 patients with symptomatic (mean New York Heart Association class 2.7) CHF (mean (SD) left ventricular ejection fraction 29.9 (8)%), range 9–46% were studied 4–6 hours after ACE inhibitor dosing. A representative range of ACE inhibitors (enalapril, lisinopril, captopril, perindopril, and fosinopril) was examined.

Results—Supine measurements showed a wide range of AII (10.5 (25.5) pg/ml), aldosterone (150.8 (136) pg/ml), and serum ACE (12.1 (13.3) EU/l; excludes captopril data) concentrations on diuretics. AII concentrations > 10 pg/ml were seen in 15% of patients, and aldosterone concentrations > 144 pg/ml were seen in 38% of patients. All concentrations were significantly correlated (p < 0.001) with ACE but not with aldosterone concentrations. Aldosterone concentrations were not significantly correlated with ACE concentrations.

Conclusions—AII “reactivation” occurred in 15% and failure of aldosterone suppression in 38% of routine CHF patients taking ACE inhibitor treatment. AII “reactivation” was associated with both low and high levels of ACE activity, which suggests that multiple different mechanisms are at play. In patients with high plasma ACE concentrations, non-compliance should be considered along with inadequate dose titration. In patients with low plasma ACE and high AII concentrations, non-ACE mediated production of AII may be operative. Raised aldosterone concentrations appear to be more common than AII “reactivation”. It is important to establish the cause of detectable or increased AII concentrations in a heart failure patient treated with an ACE inhibitor. The measurement of serum ACE may help to identify the likely cause as poor compliance or inadequate dose.

Keywords: heart failure; hormone suppression; angiotensin II; aldosterone; angiotensin converting enzyme inhibitors; compliance.

The effectiveness of angiotensin converting enzyme (ACE) inhibitors in reducing heart failure mortality may be largely attributable to hormone suppression. However, the reductions in angiotensin II (AII) and aldosterone may not be maintained with chronic ACE inhibitor treatment. While acute treatment with an ACE inhibitor can virtually eliminate AII from the plasma, chronic treatment has been associated with the reappearance of measurable AII in some patients. This has been called AII “reactivation” and is associated with a poor prognosis. It is also known that aldosterone does not remain suppressed during chronic ACE inhibitor treatment and this phenomenon has been called aldosterone escape.

While AII “reactivation” and aldosterone escape have been identified in the setting of clinical trials, very little is known about their incidence in routine clinical practice. The main aim of this study was to examine the prevalence of AII and aldosterone escape in an unselected group of patients with congestive heart failure (CHF) treated with ACE inhibition where compliance and dose titration are less rigorous than in clinical trials. We assessed the degree of neurohormonal suppression of aldosterone and AII in unselected patients with stable heart failure during routine treatment and examined the relation between them in order to examine possible mechanisms of AII “reactivation”.

Patients and methods

Patients were recruited from an outpatient heart failure clinic or during hospitalisation for coincidental illness (n = 25) unrelated to chronic cardiac failure (for example, minor surgery, investigational or diagnostic procedures, intermittent reversible myocardial ischaemia). All patients were being treated for chronic heart failure with stable doses of diuretic and ACE inhibitor. Diagnosis had been made on the basis of medical history, ongoing symptoms, and physical examination and had been confirmed by a one or other of radionuclide scintigraphy (n = 63; MUGA ejection fraction 34 (8)%, range 9%–46%), contrast ventriculography (n = 22; qualitative comments made on the ventriculogram report), or echocardiography (n = 75; mean fractional shortening 22 (5)%, range 8–35%). Some patients had multiple investigations. Details of patient demography, aetiology of...
heart failure, and cardiac treatments are listed in table 1.

**DRUG TREATMENT**

All subjects had received a constant dosage of ACE inhibitor for not less than four weeks before study and had no changes to diuretic regimen. A variety of ACE inhibitors was being used. ACE inhibitor treatment was normalised to an equivalent dose in mg of enalapril per day using accepted pharmacological bioequivalence relations.

**PROCEDURE**

The protocol for the study was passed by the local research and ethics committee before recruitment. All patients received an information leaflet before giving written and informed consent to participate in the study.

Patients were studied between 11:00 and 13:00 after routine self administration of drug treatment (approximately 4–6 hours after ACE inhibitor dosing). Intentionally no inquiries were made as to compliance with any treatment.

On reporting for the study, patients were weighed, underwent symptomatic inquiry and physical examination, and had a resting ECG documented. Patients were then rested supine (minimum 30 minutes) after insertion of a heparinised venous cannula in an antecubital vein to facilitate blood sampling.

Blood samples (40 ml) for hormone and electrolyte studies were drawn rapidly into chilled syringes and distributed into appropriate aliquots with inhibitors (see below) on ice. Samples were immediately centrifuged at 2–3°C at approximately 3500 rpm, and serum or plasma separated and stored at −70°C for later batched hormone analyses (within six months). Samples for serum electrolytes were processed on the day of collection.

**LABORATORY METHODS OF ANALYSIS**

**Serum ACE activity**

ACE was determined using direct radioimmunoassay operating within 2% intra- and 4% interassay coefficient of variation and with a lower limit of detection of 0.5 EU/l.

**Plasma angiotensin peptides**

Samples were collected into a chilled inhibitor cocktail containing renin inhibitor (enalikiren), ACE inhibitor (enalaprilat), and phenyl methy sulpho ethionamide before the separation of plasma to prevent further in vitro generation or interconversion of angiotensins. Preparative column chromatography was conducted according to the methodology of Nussberger and colleagues before the determination of angiotensin I (AI) and AI by conventional radioimmunoassay (Cardiovascular Assays, Glasgow, UK) using antiserum with minimal AI/AII cross-reactivity.

**Serum and urinary electrolytes**

Samples and aliquots were assayed on the day of collection using standard autoanalyser technology in our routine hospital clinical chemistry service operating with quality control standards (± 2.5% interassay coefficient of variation).

**Plasma aldosterone**

Samples of plasma were analysed from frozen plasma using a commercial radioimmunoassay (Peninsula Laboratories, California, USA).

**Statistical methods**

Statistical analyses were conducted using the SPSS program. Linear correlation (Pearson) was assessed, and analysis of variance (ANOVA) and covariance (ANCOVA) with Bonferroni correction were applied to investigate differences between samples where relevant. All data are given as mean (SD).

**Results**

**Serum hormones**

In the whole sample there was a large variation in measured hormone values for each of the parameters studied (table 2).

**Serum ACE**

Patients taking captopril (n = 10) have been excluded from analyses involving ACE. Captopril’s poor affinity for the enzyme in vitro makes serum ACE concentrations an inaccurate representation of the in vivo situation. During treatment with ACE inhibition in the doses outlined there was an appreciable failure of suppression of serum ACE activity during the period at which peak inhibitory effect would be anticipated—that is, 4–6 hours after dosing (fig 1). The distribution of measured serum ACE activity was wide. During chronic diuretic treatment one third of patients (27 of 81) had undetectable ACE (< 2 EU/l), one third (26 of 81) measurable ACE activity (2–10 EU/l), and one third (28 of 81) un-suppressed (> 10 EU/l) ACE.

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<table>
<thead>
<tr>
<th>Table 1: Patient demography and baseline treatment</th>
<th>Mean or n</th>
<th>SD</th>
</tr>
</thead>
<tbody>
<tr>
<td>Total number</td>
<td>91</td>
<td></td>
</tr>
<tr>
<td>Male</td>
<td>78</td>
<td></td>
</tr>
<tr>
<td>Body mass index (kg/m²)</td>
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<td>6</td>
</tr>
<tr>
<td>Age (years)</td>
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</tr>
<tr>
<td>Cause of CHF</td>
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<td></td>
</tr>
<tr>
<td>Previous infarction</td>
<td>84</td>
<td></td>
</tr>
<tr>
<td>Idiopathic dilated cardiomyopathy</td>
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<td></td>
</tr>
<tr>
<td>AF and hypertensive heart disease</td>
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<td></td>
</tr>
<tr>
<td>Left ventricular ejection fraction (%)</td>
<td>29.8</td>
<td>9</td>
</tr>
<tr>
<td>Treatment</td>
<td></td>
<td></td>
</tr>
<tr>
<td>Fruside (mg/day)</td>
<td>67</td>
<td>57</td>
</tr>
<tr>
<td>ACE inhibitor</td>
<td></td>
<td></td>
</tr>
<tr>
<td>Enalapril</td>
<td>43</td>
<td></td>
</tr>
<tr>
<td>Lisinopril</td>
<td>35</td>
<td></td>
</tr>
<tr>
<td>Captopril</td>
<td>10</td>
<td></td>
</tr>
<tr>
<td>Perindopril</td>
<td>2</td>
<td></td>
</tr>
<tr>
<td>Fosinopril</td>
<td>1</td>
<td></td>
</tr>
<tr>
<td>Nitrates (n = 35) (mg/day)</td>
<td>49</td>
<td>13</td>
</tr>
<tr>
<td>Digoxin (n = 31) (mg/day)</td>
<td>0.155</td>
<td>0.06</td>
</tr>
<tr>
<td>Aspirin (n = 39) (mg/day)</td>
<td>132</td>
<td>67</td>
</tr>
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</table>

*Excluding patients receiving oral captopril.

<table>
<thead>
<tr>
<th>Table 2: Total sample measured hormones (mean (SD))</th>
</tr>
</thead>
<tbody>
<tr>
<td><strong>ACE activity</strong> (EU/l)</td>
</tr>
<tr>
<td>-------------------------</td>
</tr>
<tr>
<td>On diuretic</td>
</tr>
</tbody>
</table>

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AF, atrial fibrillation.
Serum AII
Considering the whole sample including captopril treated patients, there was again a notable variation in measured concentrations (fig 2). During chronic treatment full suppression of AII to very low concentrations (< 2 pg/ml) was seen in nearly one third of patients (28 of 91). In these patients it may be said that AII had been eliminated from serum by treatment. There was some suppression below normal (1–10 pg/ml) in 45 of 91 (49%) patients. Normal or modestly raised AII (10–20 pg/ml) concentrations were seen in 3 of 91 (3%) patients, and increased AII (> 20 pg/ml) concentrations were seen in 10 of 91 (11%) of the sample. Again these values are seen despite chronic drug treatment.

Serum aldosterone
Including the patients receiving captopril, supine aldosterone showed a wide spectrum of measured values (fig 3). Some patients had considerable increases of aldosterone regardless of ACE inhibitor or diuretic treatment. In one patient the measured concentration (on diuretic treatment) was in the range associated with primary aldosteronism (> 1000 pg/ml). During chronic treatment suppression of aldosterone (< 144 pg/ml) was noted in 56 of 91 patients (61%), 35 of 91 (38%) had raised aldosterone (> 144 pg/ml), and 13 of 91 (14%) had notably increased aldosterone (> 300 pg/ml).

Relation between ACE activity, AII, aldosterone, and ACE inhibitor treatment
Looking at the associations between the measured parameters, plasma ACE activities were very variable at any given dose of ACE inhibitor (fig 1). The same was true for both aldosterone and AII (figs 2 and 3). Low doses of ACE inhibitor (equivalent of 2.5–5 mg enalapril/day) was seen to produce excellent AII and aldosterone suppression in some individuals.

There was no significant relation between measured AII and aldosterone (fig 4). If AII “reactivation” is assumed to be a measured value of > 10 pg/ml, despite chronic ACE inhibitor treatment, it occurred in the presence of a wide range of measured ACE activities. In those patients with AII concentrations > 10 pg/ml approximately 50% had a plasma ACE activity > 20 EU/l.

There was a significant correlation (p < 0.001) between plasma AII and plasma ACE concentrations (fig 5). There was no significant correlation between plasma aldosterone and ACE concentrations (fig 6). Figures 5 and 6 also show that AII “reactivation” and aldosterone escape can both occur at a wide range of different, prevailing concentrations of ACE.
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ACE activity is a very sensitive marker of the presence of an ACE inhibitor drug. Patients with high ACE activity (> 10 EU/l) and AII “reactivation” may have this pattern because of poor compliance or inadequate individual dose titration of the ACE inhibitor. In patients with raised AII concentrations and low or undetectable ACE, non-ACE mediated AII generation or “reactivation” may be responsible.

Our findings have direct relevance to therapeutic strategies designed to maximise AII suppression. In patients with AII “reactivation” with high (> 20 EU/l) or detectable (10–20 EU/l) ACE concentrations, it may be appropriate to either increase the dose of ACE inhibitor or to check compliance by assessing the biochemical response (ACE activity 4–5 hours after dosing) to supervised administration of the current ACE inhibitor dose. Clearly if poor compliance was identified (ACE eliminated below detection limit on single supervised dose) then the correct response would be to enhance compliance rather than increase the ACE inhibitor dose or change to an alternative agent, such as an AII antagonist. The only practical exception to this would be if the non-compliance was specifically related to ACE inhibitor cough. For those with AII “reactivation” but low or undetectable ACE it may be appropriate to change from an ACE inhibitor to an AT₁ receptor antagonist or consider the combination of these treatments. These patients may be generating AII by non-ACE mediated catalysis of the breakdown of AI to AII. While this alternative AII generating system has been well demonstrated at a biochemical and cellular level, its importance in integrated human pathology or physiology is as yet unclear.

The cut off values we selected for both AII and aldosterone are inevitably arbitrary. The reason for this is simply that no quantitative definition of hormone “suppression” exists despite much clinical evidence of the overall importance of failure to suppress either hormone.

Serum ACE activity in the absence of an ACE inhibitor is known to be affected by genotypic variation, but genotype has very little effect on the measured serum ACE activity once the patient is taking an ACE inhibitor. In our previous work, serum ACE fell by only 3 EU/l when lisinopril dose was quadrupled from 5 mg/day to 20 mg/day. We found undetectable ACE and AII concentrations occurred in many patients being treated chronically with what would be considered “low” doses of ACE inhibitor. Between 5–20 mg enalapril equivalents, a low dose (< 10 mg/day) was not invariably associated with raised AII or aldosterone, nor was a high prescribed dose (> 10 mg/day) a guarantee of “elimination” of AII or aldosterone in the presence of an ACE inhibitor drug. An average dose of an ACE inhibitor should virtually always eliminate ACE activity (always < 5 EU/l) on chronic dosing (seven days). Anything less than this indicates poor compliance. There are currently remarkably few data on non-compliance with drug treatment in
Chronic ACE inhibitor treatment in CHF patients, despite the fact that preliminary studies suggest that it may account for a substantial proportion of heart failure readmissions.

In summary, this observational study revealed a wide diversity of AII and aldosterone suppression seen in routinely treated CHF patients. Many patients had normalised AII and aldosterone but few had undetectable concentrations and a substantial proportion had high normal or raised values. AII and aldosterone “reactivation” occurred in the presence of both high and low ACE concentrations, which suggests that different mechanisms contribute to hormonal “reactivation”. In those with high ACE activity, non-compliance is likely to be the explanation. Since different mechanisms appear to produce AII “reactivation”, it may be important to establish the precise mechanisms in each patient before deciding how to overcome hormone “reactivation” in each individual. Serum ACE concentrations may help to distinguish between the various possible causes of AII “reactivation” in individual patients and help the doctor to take the appropriate corrective action.

We are grateful to our clinical colleagues in the departments of medicine and cardiology at Ninewells Hospital, Dundee who allowed us to approach patients under their care to take part in the study. We gratefully acknowledge the nursing support of Jessamine Robson and excellent technical assistance of Mrs WJ Coutie and Mrs LM MacFarlane with hormone and electrolyte analyses. This work was made possible by a grant from the Scottish Office (acute care health research committee).


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