Monocyte proinflammatory cytokine release is higher and glucocorticoid sensitivity is lower in middle aged men than in women independent of cardiovascular risk factors

P H Wirtz, R von Känel, N Rohleder, J E Fischer

Objective: To investigate whether stimulated monocyte cytokine release and its inhibition by glucocorticoids differs between men and women.

Design: In vitro monocyte interleukin 6 (IL-6) and tumour necrosis factor α (TNFα) release after lipopolysaccharide stimulation were assessed with and without co-incubation with increasing doses of dexamethasone and hydrocortisone separately. Glucocorticoid sensitivity was defined as the amount of a particular glucocorticoid required to inhibit lipopolysaccharide stimulated monocyte cytokine release by 50%. The established cardiovascular risk factors of age, body mass index, number of cigarettes smoked daily, low density cholesterol to high density cholesterol ratio, systolic and diastolic blood pressure, and haemoglobin A1c were used as covariates.

Setting: Aircraft manufacturing plant in southern Germany.

Patients: 269 middle aged male and 36 middle aged female employees.

Results: Release of monocyte IL-6 and TNFα (each p = 0.001) was higher in samples from men than in those from women. Inhibition of lipopolysaccharide stimulated IL-6 and TNFα release by either glucocorticoid was less pronounced in samples from men than in those from women (IL-6: dexamethasone p = 0.033, hydrocortisone p = 0.029; TNFα: dexamethasone p < 0.001, hydrocortisone p = 0.089).

Conclusions: The finding suggests that proinflammatory activity of circulating monocytes is higher in men than in women independent of cardiovascular risk factors, thereby providing one explanation for the relatively greater coronary risk in men.

O

n average, women develop heart disease some 10–15 years later than men.1 In terms of this sex dimorphism, most attention has been paid to the hypothesis that endogenous oestrogens may be cardioprotective in premenopausal women.2–4 Further explanations, either additional or associated, relate to lower blood pressure, lower concentrations of low density lipoprotein (LDL) cholesterol, or higher concentrations of high density lipoprotein (HDL) cholesterol in women than in men.5–7

Monocytes have a key role in the pathogenesis of atherosclerosis.8,9 They secrete inflammatory cytokines that further promote inflammatory activity in atherosclerotic lesions.10–13 Increased concentrations of proinflammatory cytokines such as tumour necrosis factor α (TNFα) and interleukin 6 (IL-6) have been positively associated with increased cardiovascular risk.14–16 Therefore, sex differences in monocyte cytokine release may account for distinct coronary risk between the sexes.

Consistent with this reasoning, oestrogens may affect monocyte proinflammatory cytokine production.17 In addition, a recent study suggests that men require lower concentrations of glucocorticoids than women to inhibit stress induced production of proinflammatory cytokines.18 Even though such altered glucocorticoid sensitivity may be associated with chronic inflammatory diseases19–23 such as atherosclerosis, that study examined a relatively small sample size of healthy young people not prone to coronary disease. Therefore, we investigated sex differences in monocyte glucocorticoid sensitivity in a larger population with relatively higher coronary risk due to older age and we controlled for established cardiovascular risk factors.

Regulatory processes in monocyte activation entail synthesis and release of proinflammatory cytokines as well as downregulation of cytokine release. While lipopolysaccharide is one of the most potent activators of monocytes, endogenous glucocorticoids effectively downregulate monocyte cytokine production.24–25 Following previous methods we assessed monocyte reactivity by measuring IL-6 and TNFα in whole blood after lipopolysaccharide stimulation.26–28 Monocyte glucocorticoid sensitivity was calculated as the amount of dexamethasone required to suppress lipopolysaccharide stimulated TNFα and IL-6 release by 50%.14,25

MATERIALS AND METHODS

Experimental patients

The study was part of a project conducted in an aircraft manufacturing plant in southern Germany26 and was formally approved by the institutional review board. From a total of 1760 employees, participation was offered to a representative sample of 532 men and women, who voluntarily completed a questionnaire based assessment of working conditions. Funding allowed medical examination and blood sampling of the first 325 participants (280 men, 45 women) who presented consecutively to the laboratory within the next 14 days, leaving 207 employees of the total representative sample whose biochemical measures were not assessed. Although the proportions of sex and duration of employment in years were the same in the two groups, those

Abbreviations: ELISA, enzyme linked immunosorbent assay; HDL, high density lipoprotein; IC50, concentration of dexamethasone or hydrocortisone that achieves a 50% inhibition of lipopolysaccharide stimulated cytokine release in the absence of any glucocorticoid; IL, interleukin; LDL, low density lipoprotein; MONICA, monitoring trends and determinants in cardiovascular disease; TNFα, tumour necrosis factor α.
who were not assessed were slightly younger (mean (SD) age 38.9 (12.6) v 41.6 (9.6) years). Exclusion criteria for the present study were self reported previous cardiac surgery, coronary angiography, and current medication with glucocorticoids, oral contraceptives, or antidiabetic drugs. This procedure left a study sample of 269 men and 36 women. Table 1 provides health factors including established cardiovascular risk factors of subjects.

**Experimental protocol**

Self reported medical history and health behaviour (smoking) were assessed by a 96 item questionnaire derived from the nurses’ health study and the MONICA (monitoring trends and determinants in cardiovascular disease) study.27 28 After completing the questionnaire, participants rested for 15 minutes while sitting. Thereafter, blood pressure was determined by sphygmomanometry twice within five minutes. Within two weeks, all participants were rescheduled on a work day and two hours after awakening to have a fasting blood sample collected. Assays were started within five minutes of blood collection in a cell culture facility adjacent to the blood collection room. Blood was processed by standard techniques in cooled (4 °C) citrate tubes for the TNFα assay. Overnight urine collection for cortisol measurement started at 9 pm the night before blood sampling and included the first void after awakening.29

**Glucocorticoid sensitivity and assay**

Monocytes are the main cytokine producing cells in lipopolysaccharide stimulated whole blood.30 31 The whole blood assay minimises postcollection artefacts arising from cell preparation and maintains the person’s natural environment (that is, plasma compounds and hormone concentrations) as much as possible during in vitro assays.27 29

To assess monocyte glucocorticoid sensitivity, whole blood samples from each participant were stimulated with lipopolysaccharide and then co-incubated with dexamethasone and hydrocortisone separately in incremental doses.16 22 23

The lipopolysaccharide stock solution (Escherichia coli O55:B5, no L2880, Sigma-Aldrich Chemie GmbH, Steinheim, Germany) was prepared by dissolving lipopolysaccharide in pyrogen-free sterile saline (NaCl 0.9%, Fresenius Kabi, Stans, Switzerland) to achieve a final concentration of 15 ng/ml pyrogen-free sterile saline (NaCl 0.9%, Fresenius Kabi, Stans, Germany) was prepared by dissolving lipopolysaccharide in O55:B5, no L2880, Sigma-Aldrich Chemie GmbH, Steinheim, Germany; detection limit 0.1 mg/l). LDL and HDL cholesterol, haemoglobin A1c, and urinary cortisol were determined by a commercial laboratory. To obtain a complete blood count including a differential count, blood was collected into 2.7 ml EDTA tubes (Sarstedt, Rommelshof, Germany) and processed on a Sysmex SE-9000 cell counter (Sysmex, Norderstedt, Germany) within three hours after collection.

**Biochemical analyses**

Stimulated plasma concentrations of TNFα and IL-6 were determined by commercially available enzyme linked immunosorbent assay (ELISA) kits (BD Pharmingen, San Diego, California, USA). High sensitivity ELISAs were chosen to measure plasma concentrations of TNFα (Quantikine HS, R&D Systems Europe, Abingdon, UK) and of C reactive protein (Immunolite, DPC Biermann GmbH, Germany; detection limit 0.1 mg/l). LDL and HDL cholesterol, haemoglobin A1c, and urinary cortisol were determined by a commercial laboratory. To obtain a complete blood count including a differential count, blood was collected into 2.7 ml EDTA tubes (Sarstedt, Rommelshof, Germany) and processed on a Sysmex SE-9000 cell counter (Sysmex, Norderstedt, Germany) within three hours after collection.

**Statistical analyses**

As an index for monocyte glucocorticoid sensitivity, we calculated the IC50 of the dose–response curve for dexamethasone or hydrocortisone inhibition of lipopolysaccharide induced cytokine release. The IC50 reflects the particular dexamethasone or hydrocortisone concentration that achieves a 50% inhibition of lipopolysaccharide stimulated cytokine release in the absence of any glucocorticoid. The IC50 is inversely related to glucocorticoid sensitivity—that is, a higher IC50 indicates lower sensitivity and a lower IC50 indicates higher sensitivity. The IC50 is calculated intrindividually by a logistic curve fit function (mean determination coefficient of $r^2 = 0.99$). The resulting measure is independent of the absolute number of circulating monocytes.

All calculations were done with SPSS version 10.0 (SPSS Inc, Chicago, Illinois, USA) and CurveExpert 1.3 (Daniel G Hyams, Hixson, Texas, USA) statistical software packages. All data are presented as mean (SEM). Results were considered significant at the p ≤ 0.05 level; all tests were two tailed. To approximate a normal distribution, IC50 values were log transformed. In case of missing data, cases were excluded listwise.

Across the two subject groups, univariate analyses of variance were calculated for lipopolysaccharide stimulated cytokine production and IC50 values. Repeated measure analyses of variance were calculated for dose-response curves

### Table 1 Health factors of 305 men and women studied

<table>
<thead>
<tr>
<th></th>
<th>Women (n = 36)</th>
<th>Men (n = 269)</th>
<th>p Value</th>
</tr>
</thead>
<tbody>
<tr>
<td>Age (years)</td>
<td>42.0 (1.8)</td>
<td>41.4 (0.6)</td>
<td>0.71</td>
</tr>
<tr>
<td>Body mass index (kg/m²)</td>
<td>24.77 (0.69)</td>
<td>26.76 (0.23)</td>
<td>0.004</td>
</tr>
<tr>
<td>LDLC:HDL ratio</td>
<td>2.30 (0.14)</td>
<td>3.00 (0.06)</td>
<td>0.000</td>
</tr>
<tr>
<td>Tumour necrosis factor (pg/ml)</td>
<td>1.89 (0.14)</td>
<td>1.98 (0.10)</td>
<td>0.74</td>
</tr>
<tr>
<td>C reactive protein (mg/l)</td>
<td>1.82 (0.32)</td>
<td>2.08 (0.24)</td>
<td>0.70</td>
</tr>
<tr>
<td>Systolic blood pressure (mm Hg)</td>
<td>125 (2.7)</td>
<td>133 (1.0)</td>
<td>0.005</td>
</tr>
<tr>
<td>Diastolic blood pressure (mm Hg)</td>
<td>78.2 (1.6)</td>
<td>82.8 (0.6)</td>
<td>0.013</td>
</tr>
<tr>
<td>Cigarettes/day</td>
<td>4.4 (1.3)</td>
<td>5.9 (0.6)</td>
<td>0.40</td>
</tr>
<tr>
<td>Haemoglobin A1c (%)</td>
<td>5.17 (0.11)</td>
<td>5.17 (0.03)</td>
<td>0.99</td>
</tr>
<tr>
<td>Urinary cortisol (µg/l)</td>
<td>38.8 (4.6)</td>
<td>43.5 (2.0)</td>
<td>0.41</td>
</tr>
<tr>
<td>Monocytes (×10⁹/ml)</td>
<td>5.72 (0.32)</td>
<td>5.92 (0.01)</td>
<td>0.50</td>
</tr>
</tbody>
</table>

Values are mean (SEM).

HDL, high density lipoprotein cholesterol; LDL, low density lipoprotein cholesterol.
of dexamethasone or hydrocortisone inhibition of lipopolysaccharide induced cytokine production. We applied Huynh-Feldt corrections for repeated measures, which allow comparisons of cytokine release per cell. All analyses of variance were computed and are presented for the established cardiovascular risk factors of age, body mass index, number of cigarettes smoked daily, LDL:HDL ratio, systolic and diastolic blood pressure, and haemoglobin A1c as covariates throughout.

RESULTS
Cardiovascular risk factors and health factors
Men had higher body mass index, LDL:HDL ratio, and systolic and diastolic blood pressures than women (table 1). Of note, lack of a difference in overnight urinary cortisol excretion indicates that men and women had equal hypothalamic–pituitary–adrenal axis activity.

Stimulation of cytokine production by lipopolysaccharide
There were no sex differences in basal plasma C reactive protein and TNFα concentrations (table 1). After controlling for the absolute monocyte count and all established cardiovascular risk factors mentioned above, the release of lipopolysaccharide stimulated TNFα (F1,249 = 12.2, p = 0.001, n = 258) and IL-6 release (F1,248 = 12.0, p = 0.001, n = 267) was significantly higher in samples from men than in those from women (figs 1 and 2).

Inhibition of stimulated cytokine production
Figures 1 and 2 show inhibition curves for TNFα and IL-6 releases following lipopolysaccharide stimulation plotted against increasing concentrations of the two glucocorticoids dexamethasone and hydrocortisone. The curves show reduced capacities of both glucocorticoids to inhibit cytokine release in men as compared with women. When controlling for all established cardiovascular risk factors, TNFα inhibition was less pronounced in samples from men than in those from women with both dexamethasone (F2,6,614.9 = 6.7, p < 0.001, n = 250) and hydrocortisone (F3.1,726.5 = 4.1, p = 0.006, n = 246). Likewise, in cells obtained from men IL-6 release was inhibited by increasing concentrations of either dexamethasone or hydrocortisone to inhibit lipopolysaccharide stimulated release of IL-6 and TNFα than did samples from women, while the cytokine release without glucocorticoid co-incubation was higher in samples from men than in those from women. The findings suggest higher proinflammatory activity in blood monocytes of men than in monocytes harvested from women. The notion becomes even stronger given that we controlled for established cardiovascular risk factors. Previous studies have shown that, on average, men have higher body mass index, higher LDL:HDL ratio, and higher systolic and diastolic blood pressures than women.14,24-28 Therefore, to prevent confounding, cardiovascular risk factors need to be controlled for when testing for a unique sex difference in monocyte glucocorticoid sensitivity.

What may the clinical implications of our findings be and how do they fit with the literature? Our findings indicate that the proinflammatory response of monocytes to an equal

---

**Figure 1** Curves showing the concentration of dexamethasone or hydrocortisone that achieved a 50% inhibition of lipopolysaccharide stimulated cytokine release in the absence of any glucocorticoid (IC50). Lipopolysaccharide stimulated release of tumour necrosis factor α (TNFα) and inhibition of TNFα release by glucocorticoids in blood samples from men and women (mean (SEM)). Note that the starting points of the individual IC50 curves refer to the uninhibited release of TNFα in response to lipopolysaccharide. Samples from men produced more TNFα than did those from women when stimulated with lipopolysaccharide (F1,249 = 12.2, p = 0.001) and when inhibited by increasing concentrations of dexamethasone (F3.1,726.5 = 4.1, p = 0.006).
amount of lipopolysaccharide, such as that produced by bacteria present within an atherosclerotic lesion, is higher in men than in women. Corroborating these findings, some investigators found greater TNFα release with lipopolysaccharide stimulation in men than in women, while others did not detect such differences. Further driving such controversy, Rohleder and colleagues found lower sensitivity of monocyte IL-6 release with dexamethasone inhibition in men than in women, while Daun and colleagues did not find such a sex difference in hydrocortisone regulation of lipopolysaccharide stimulated production of IL-1β from peripheral blood mononuclear cells. In contrast to our study, however, those studies had relatively smaller population samples that consisted mainly of young and healthy people who conceivably have a lower cardiovascular risk than our middle aged participants.

Epidemiological data support the notion that increased plasma concentrations of TNFα and IL-6 increase the risk of coronary artery disease. Thus, differential regulation of monocyte TNFα and IL-6 production may provide a link between sex and the progression of atherosclerotic lesions. Our data suggest that monocytes from men not only are predisposed towards more pronounced release of TNFα and IL-6 but also are less sensitive to endogenous signals (glucocorticoids) responsible for curtailing inflammatory responses. This may lead to a more rapid development of atherosclerosis in men than in women.

Although speculative, one possible explanation for observed sex differences in lipopolysaccharide stimulated cytokine release and in glucocorticoid sensitivity may relate to different concentrations of gonadal steroids. The female immune system is influenced by both oestrogens and progesterones. Oestrogens may dose dependently affect lipopolysaccharide associated changes in immune responses of macrophages. Compared with men, women in the luteal phase of the menstrual cycle had diminished monocyte release of TNFα and IL-6 with lipopolysaccharide. In that study, these effects correlated with the concentration of oestradiol in plasma. The male immune system, on the other hand, is under the control of testosterone and other androgens, which apparently do not affect macrophage function.

Despite increasing knowledge of the pathways through which gonadal steroids and glucocorticoids influence immunological target tissues, it remains unknown how these steroids interact at peripheral inflammatory sites. The activated oestrogen receptor has been shown to inhibit the transcription factors nuclear factor κB and activator protein 1. Progesterone, because of its structural similarities with
the glucocorticoids receptor, has been reported to increase
cytosolic mRNA concentrations of the nuclear factor xB
inhibitor.5 By these mechanisms, both female gonadal
steroids seem to exert anti-inflammatory effects.
Da Silva has proposed several mechanisms by which
gonadal steroids may modulate the sensitivity of target
tissues towards glucocorticoids.49 In animal and cell culture
studies, 17β oestradiol has been shown to decrease the
expression of the glucocorticoid receptor in the cytoplasm in
an inactive state (although it can be ligand activated), are
modulated by female gonadal steroids.50
Several limitations of our study require consideration.
Firstly, our investigation focused on a working population
and, thus, may not be generalisable to clinical populations
and to people with overt atherosclerotic disease in particular.
Secondly, we can only speculate as to whether the observed
differences between men and women are clinically relevant.
Thirdly, as we did not assess sex steroids, possible explana-
tions for the observed sex differences in cytokine release
and glucocorticoid sensitivity are not unequivocally attributable
to hormonal issues. While Levene’s test showed that variances
of uninhibited lipopolysaccharide stimulation of TNFα and IL–6 and of the four IC₅₀ estimates were equal, three of the 20 computations
across the four inhibition curves suggested a violation of this
assumption (data not shown). Nonetheless, since we built
our argument on sex differences in uninhibited stimulation
of proinflammatory cytokines and calculated IC₅₀ values, we
feel that this caveat did not invalidate our findings.
In summary, our data show that monocytes from middle-
aged men have reduced responsiveness to glucocorticoids
even when established cardiovascular risk factors are con-
trolled for. This finding of reduced glucocorticoid sensitivity
in men implies relatively sustained cytokine production once
monocytes have encountered stressful stimuli (such as
lipopolysaccharide). Such a mechanism may be one possible
biological pathway linking sex to progression of athero-
sclerotic disease and increased coronary risk in men.

ACKNOWLEDGEMENTS
This work was supported by grants from the EADS GmbH, Werk
Augsburg, Germany, and from the Swiss Federal Institute of
Technology.

Authors’ affiliations
P H Wirtz, Institute of Clinical Psychology II, University of Zurich, Zurich,
Switzerland
R von Känel, Division of Psychosomatic Medicine, Department of
Internal Medicine, University Hospital, Barne, Switzerland
N Rohleder, Institute of Biopsychology, Technical University, Dresden,
Germany
J E Fischer, Institute for Behavioural Sciences, Swiss Federal Institute of
Technology, Zurich, Switzerland

REFERENCES
1 Rossow JE. Hormones, genetic factors, and gender differences in car-
2 Schwerz DW. Pencoff S. Sex differences and the effects of sex hormones on
3 Mendolos-NH, Karas RH. The protective effects of estrogen on the
4 Reckelhoff JF. Gender differences in the regulation of blood pressure.
Hypertension 2001;37:119–208
5 Kirkland AE, Moxley JS, Probstfield JL, et al. Decrease in plasma high-density
lipoprotein cholesterol levels at puberty in boys with delayed adolescence:
correlation with plasma testosterone levels. JAMA 1987;257:502–7
1999;340:115–26
7 Plutjus J. Inflammatory pathways in atherosclerosis and acute coronary
syndromes. Am J Cardiol 2001;88:10K–SK
8 Pena LR, Hill DB, McClain CJ. Treatment with glutathione precursor decreases
9 Van der Wal AC, Gasteiger KM, Tappes, et al. Adhesion molecules on the
endothelium and mononuclear cells in human atherosclerotic lesions.
Am J Pathol 1992;141:1427–33
10 Ridker PM, Rifai N, Pfeffer MA. Elevation of tumor necrosis factor-alpha
and increased risk of recurrent coronary events after myocardial infarction.
Circulation 2000;101:2149–53
and the risk of future myocardial infarction among apparently healthy men.
Circulation 2000;101:1767–72
12 Kookunen B, Penttila K, Kemppainen A, et al. C-reactive protein, fibrinogen,
interleukin-6 and tumour necrosis factor-alpha in the prognostic classification
13 Miller L, Hunt JS. Sex steroid hormones and macrophage function. Life Sci
1996;59:1–14
14 Rohleder N, Schommer NC, Hellhammer DH, et al. Sex differences in
glucocorticoid sensitivity of proinflammatory cytokine production after
in quiescent Crohn’s disease: an ex vivo study using whole blood cell cultures.
16 Schlaghecke R, Kernely E, Wollenhaupt J, et al. Glucocorticoid receptors in
17 Diboltis JA, Martel-Pelletier J, Antaky T, et al. Reduced expression of
glucocorticoid receptor levels in human osteoarthritic chondrocytes: role in
the suppression of metalloproteinase synthesis. J Clin Endocrinol Metab
1993;76:1128–34
18 Lane SJ, Lee TH. Mechanisms of corticosteroid resistance in asthmatic patients.
19 Spha J, Landempe L, Nimmagadda S, et al. Effects of glucocorticoids on
lymphocyte activation in patients with steroid-sensitive and steroid-resistant
20 Wick G, Hu Y, Schwarz S, et al. Immunodendocrine communication via the
hypothalamo-pituitary-adrenal axis in autistic diseases. Endocr Rev
1993;14:539–63
21 Chrousos GP. The hypothalamic-pituitary-adrenal axis and immune-mediated
22 Breuning LM, Dempsey W, Uhl J, et al. Hydrocortisone regulation of
interleukin-6 polyalanine production by a purified population of human peripheral
variations in plasma cortisol differentially regulate interleukin-1 beta (IL-1 beta),
IL–6, and tumor necrosis factor-alpha (TNF α) production in humans: high sensitivity of TNF alpha and resistance of IL–6. J Clin Endocrinol Metab
1997;82:2182–91
24 Maier MD, Martens H, Hagleitner M, et al. Tumor necrosis factor alpha decreases,
and interleukin-10 increases, the sensitivity of human monocytes to
dexamethasone: potential regulation of the glucocorticoid receptor. J Clin
Endocrinol Metab 1999;84:2834–9
glucocorticoid sensitivity in healthy adults. J Clin Endocrinol Metab
2000;85:3733–9
26 Schnorpeil P. Allergic load, vital exhaustion and job characteristics in
employees of the manufacturing industry. Zurich, University of Zurich, 2001.
27 Michael YL, Colditz GA, Coakley E, et al. Health behaviors, social networks,
and healthy aging: cross-sectional evidence from the nurses’ health study.
Qual Life Res 1999;8:711–22
28 Jonsson D, Rosengren A, Dotteval A, et al. Job control, job demands and
social support at work in relation to cardiovascular risk factors in MONICA
and its health consequences. MacArthur studies of successful aging. Arch
Intern Med 1997;157:2259–68
30 Wright SD, Ramos RA, Tobias PA, et al. CD14, a receptor for complexes of
lipopolysaccharide (LPS) and LPS binding protein. Science 1990;249:1431–3
1998;840:787–802
32 De Groote D, Zangerle PF, Govaert Y, et al. Direct stimulation of cytokines
(IL–1 beta, TNF alpha, IL–6, IL–2, IFN-gamma and GM-CSF) in whole blood.
33 De Groote D, Zangerle PF, Govaert Y, et al. Novel method for the measurement of
cytokine production by a one-stage procedure. J Immunol Methods
1993;163:259–67
34 Winkelm H, Hoegerm A, Christensen HR, et al. 24-h ambulatory blood
pressure in 352 normal Danish subjects, related to age and gender.
Am J Hypertens 1995;8:978–84
monitoring in a nonacademic setting: effects of age and sex. Am J Hypertens
1992;5:616–23
human peripheral blood mononuclear cells: role of estrogen in modulating
LPS-induced cytokine secretion in an in vivo septic model. Shock
2001;16:240–3
37 Schwartz E, Schafer C, Bode JC, et al. Influence of the menstrual cycle on the

www.heartjnl.com

Downloaded from http://heart.bmj.com/ on April 28, 2017 - Published by group.bmj.com
Unstable angina in a patient with single coronary artery

A 58 year old woman with hypertension, hypercholesterolaemia, and family history of coronary heart disease presented with a five year history of exertional chest discomfort, relieved by rest and nitrate administration.

Coronary angiography demonstrated a single left coronary ostium (aortography confirmed absence of right coronary artery), left circumflex artery was dominant, and the right coronary artery arose from an unusual location, just after the first septal branch of the left anterior descending artery (LAD), passing anteriorly to the pulmonary trunk and crossing to the right side of the heart. According to the Lipton classification, this anomaly could be classified as LII-A (panels A and B).

Interestingly, although the coronary tree was generally free of atheroma, a significant stenosis was demonstrated within the abnormal artery (panels A and B, arrowheads). This was wired and stented directly with a 2.75 x 8 mm stent, but resulted in some impingement in the main LAD, perhaps due to “snow-plough” (panel C, arrowhead). After preinflation of both branches, a “kissing balloon” inflation was performed in the LAD and in the aberrant vessel, with good angiographic final appearance (panel D).

Coronary artery anomalies occurs in 0.64–5.6% of patients undergoing coronary angiography. They have been associated with increased risk of sudden death, mainly when an aberrant branch is compressed between the great vessels. In our case the unusual anatomy may have played a role in determining the location of this single lesion, as an almost right angle takeoff is associated with decreased mechanical shear stress, and could predispose to atheroma formation.

I Porto
A P Banning
i.porto@eudoramail.com
Unstable angina in a patient with single coronary artery

I Porto and A P Banning

Heart 2004 90: 858
doi: 10.1136/hrt.2003.028753

Updated information and services can be found at:
http://heart.bmj.com/content/90/8/858

These include:

Email alerting service
Receive free email alerts when new articles cite this article. Sign up in the box at the top right corner of the online article.

Notes

To request permissions go to:
http://group.bmj.com/group/rights-licensing/permissions

To order reprints go to:
http://journals.bmj.com/cgi/reprintform

To subscribe to BMJ go to:
http://group.bmj.com/subscribe/