Angiotensin converting enzyme inhibitor prevents left ventricular remodelling after myocardial infarction in angiotensin II type 1 receptor knockout mice

M Yoshiyama, Y Nakamura, T Omura, Y Izumi, R Matsumoto, S Oda, K Takeuchi, S Kim, H Iwao, J Yoshikawa

Background: It is well known that angiotensin converting enzyme (ACE) inhibitors and angiotensin II type 1 (AT1) receptor blockers (ARBs) prevent left ventricular (LV) remodelling after myocardial infarction (MI). However, it is still not clear whether inhibition of the AT1 receptor is enough to prevent LV remodelling after MI.

Objective: To elucidate the effects of ACE inhibitors that are not mediated by the AT1 receptor on LV remodelling, MI was experimentally induced in wild-type (WT-MI) mice and AT1 receptor knockout (KO-MI) mice.

Methods: Mice were divided into six groups: WT-control, KO-control, WT-MI, KO-MI, WT-MI treated with an ACE inhibitor, and KO-MI treated with an ACE inhibitor. Four weeks after MI, cardiac function was assessed by Doppler echocardiography and non-infarcted myocardial mRNA expression by northern blot analysis.

Results: Cardiac function decreased significantly in the MI groups compared with the sham operated groups. Additionally, in the MI groups end diastolic dimension, E wave velocity, the ratio of peak velocity of E wave to A wave, deceleration rate of E wave, and mRNA expression of atrial natriuretic peptide, brain natriuretic peptide, and collagens I and III increased significantly compared with the sham groups.

LV remodelling after MI was prevented in KO-MI mice compared with WT-MI mice. ACE inhibitor administration significantly attenuated progressive LV remodelling in both WT and KO-MI groups.

Conclusion: ACE inhibitors can prevent the LV remodelling process that accompanies cardiac dysfunction after MI, even in AT1 KO mice. These findings suggest that ACE inhibitors prevent LV remodelling after MI by mechanisms other than inhibition of angiotensin AT1 receptor mediated effects.

A ccumulating evidence supports the proposal that angiotensin II type 1 (AT1) receptor blockers (ARBs) have nearly the same beneficial effects as angiotensin converting enzyme (ACE) inhibitors on cardiac hypertrophy, remodelling, and heart failure.1–3 However, the pharmacological profiles of ARBs and ACE inhibitors differ substantially. ARBs can inhibit the action of angiotensin II generated through not only ACE but also alternative pathways,4 whereas ACE inhibitors block the breakdown of bradykinin.5–6 Therefore, combination treatment with an ACE inhibitor and ARB may offer benefits greater than the benefits of either agent used alone. In the Val-HeFT (valsartan heart failure trial)7 and CHARM (candesartan in heart failure–assessment of reduction in mortality and morbidity) studies,8 combination treatment with an ACE inhibitor and ARB provided enhanced benefits in heart failure.

Recently, we showed that combined administration of an ACE inhibitor and an ARB prevents left ventricular (LV) remodelling after myocardial infarction (MI) in rats more effectively than either drug administered on its own.9 Valsartan is as effective as captopril in patients at high risk for cardiovascular events after MI; however, combining valsartan with captopril does not improve survival.10

We hypothesised that ACE inhibitors may prevent LV remodelling after MI despite the lack of expression of the AT1 receptor. In the present study, we experimentally induced MI in wild-type (WT-MI) mice and AT1 receptor knockout (KO-MI) mice that were either treated or not treated with an ACE inhibitor. At four weeks after MI, we assessed cardiac function by Doppler echocardiography and cardiac gene expression to ascertain whether ACE inhibitors can prevent LV remodelling after MI without AT1 receptor inhibition.

METHODS

Induction of MI

KO mice and WT mice (all 16 week old C57BL/6 of the same genetic background, weighing 29–31 g) were used in the present study.11 An MI was induced according to a previously described method.12 13 Mice were anaesthetised with pentobarbital sodium (35 mg/kg intraperitoneally) and lidocaine hydrochloride (10 mg/kg intraperitoneally). After tracheal intubation, the mice were artificially ventilated with a small animal respirator. MI was induced by permanently ligating the left coronary artery with a 9-0 nylon surgical suture under a dissection microscope. The same surgical procedures were also performed in sham operated mice, except for the coronary ligation.

The mice were separated into six groups (7–10 mice in each group): sham operated WT mice (WT-controls); sham operated KO mice (KO-controls); WT-MI mice; KO-MI mice; WT-MI mice treated with an ACE inhibitor (imidapril

Abbreviations: ACE, angiotensin converting enzyme; ANP, atrial natriuretic peptide; ARB, angiotensin II type 1 receptor blocker; AT1, angiotensin II type 1; BNP, brain natriuretic peptide; CHARM, candesartan in heart failure–assessment of reduction in mortality and morbidity; %FS, percentage of fractional shortening; KO, knockout; LV, left ventricular; MI, myocardial infarction; RV, right ventricular; Val-HeFT, valsartan heart failure trial; WT, wild type
10 mg/kg/day); and KO-MI mice treated with an ACE inhibitor (imidapril 10 mg/kg/day). Immediately after coronary ligation, imidapril was administered in drinking water based on the observed average water consumption of mice. Four weeks after MI, the heart rate and systolic blood pressure of conscious mice were measured by the tail cuff method, echocardiography was performed, and after the animals were killed the ventricles were excised. Infarct size was calculated and expressed as a percentage of LV surface area as previously described. Mice with an infarct size < 20% were excluded from analysis.

**Echocardiographic studies**

Mice were lightly anaesthetised with ketamine hydrochloride (25 mg/kg intraperitoneally) and xylazine (10 mg/kg intraperitoneally). Echocardiography was performed with a commercially available echocardiographic system equipped with a 12 MHz phased array transducer (SONOS 5500, Phillips, Andover, Massachusetts, USA). A two dimensional short axis view of the LV was obtained at the level of the papillary muscles. Pulsed wave Doppler spectra of mitral inflow velocities were recorded from the apical four chamber view, with the sample volume placed near the tips of the mitral leaflets and adjusted to the position at which velocity was maximum and the flow pattern was laminar. The sample volume was set at the smallest size available. All Doppler spectra were recorded at a paper speed of 200 mm/s and analysed off line.

**RNA preparation and northern blot hybridisation**

All procedures were performed as previously described. In brief, total RNA was isolated from the individual non-infarcted LV and right ventricle by the guanidium thiocyanate-phenol-chloroform method and 20 mg of total RNA was subjected to 1% agarose gel electrophoresis and transferred to a nylon membrane. Hybridisation was carried out with a (12P)-dCTP labelled cDNA probe for atrial natriuretic peptide (ANP), brain natriuretic peptide (BNP), collagen type I, collagen type III, or glyceraldehyde-3-phosphate dehydrogenase. The density of an individual mRNA band was measured with a bioimaging analyser (BAS-2000, Fuji Photo Film Co, Tokyo, Japan).

**Statistical analysis**

All results are expressed as mean (SEM). Significance was determined by analysis of variance. Differences were considered significant at p < 0.05.

**RESULTS**

**Anatomical pathology**

Figure 1 shows the heart shapes from the six groups. WT-MI and KO-MI (MI groups) hearts were enlarged compared with WT-controls or KO-controls; the enlargement in KO-MI mice was less than in WT-MI mice. ACE inhibitor treatment significantly attenuated the enlargement in MI even in the KO mice model. Figure 2 shows transverse LV sections stained by azan from the six groups taken from the apical view. The LV cavities in WT-MI and KO-MI mice were dilated. LV cavity dilatation in KO-MI mice was less than in WT-MI mice. ACE inhibitor treatment significantly attenuated dilatation of the LV cavity in MI, even in the KO model.

**Haemodynamic data and ventricular weights**

Table 1 shows the haemodynamic data and ventricular weights in the six groups. Blood pressure was significantly lower in KO-controls than in WT-controls (p < 0.01). In KO-MI mice, blood pressure was significantly lower than in WT-MI mice (p < 0.01). Although ACE inhibitor treatment significantly decreased blood pressure in WT-MI mice (p < 0.05), a trend towards decreased blood pressure in KO-MI mice was also observed. LV and right ventricular (RV) weights, corrected for body weight, in the MI groups were significantly greater than in the corresponding control groups. ACE inhibitor treatment significantly reduced the LV and RV weights in WT-MI mice and tended to reduce the LV and RV weights in KO-MI mice. Infarct size was similar in all groups.

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**Figure 1** Anatomical pathology from a wild-type (WT) control mouse, a knockout (KO) control mouse, a WT mouse with experimentally induced myocardial infarction (MI), and a KO-MI mouse. Angiotensin converting enzyme inhibitor (ACEI) was administered to WT-MI and KO-MI mice. WT-MI and KO-MI mice had enlarged hearts compared with WT-control mouse or KO-control mouse. ACEI prevented this enlargement.
Doppler echocardiographic assessments of LV geometry and function

Figure 3 shows representative echocardiographic short axis images taken during the internal diastolic phase. Table 2 shows the echocardiographic assessments. The LV diastolic dimension in the MI groups was significantly larger than in the control groups. Moreover, ACE inhibitor significantly reduced diastolic dimension in the corresponding MI groups, even in the KO model. The MI groups had significant systolic dysfunction, as shown by the percentage of fractional shortening (%FS). The %FS in KO-MI mice was significantly higher than in WT-MI mice. Moreover, ACE inhibitor treatment significantly improved the %FS in the corresponding MI groups, even in the KO model. The ratio of E wave peak velocity to A wave peak velocity in the MI groups was much higher than in the control groups. The ratio in KO-MI mice was significantly lower than in WT-MI mice (p < 0.01). However, ACE inhibitors did not significantly further lower the ratio in the corresponding MI groups, even in the KO model.

Cardiac gene expression of ANP, BNP, and collagen types I and III

Table 3 and fig 5 show the results of cardiac gene expression determination in the six groups. mRNA expression of ANP, BNP, and collagen types I and III by the non-infarcted myocardium was significantly increased in the MI groups compared with the control groups. mRNA expression in non-infarcted myocardium was significantly decreased in KO-MI mice compared with WT-MI mice. Moreover, ACE inhibitor significantly decreased mRNA expression in the corresponding MI groups, even in the KO model.

DISCUSSION

Chronic heart failure may develop after MI, which causes LV dilatation and adaptive responses in both infarcted and non-infarcted regions of the heart. These changes, called post-MI remodelling, may contribute to LV systolic and diastolic dysfunction after MI. Previous studies have suggested that the cardiac renin-angiotensin system is activated during this remodelling process. In addition, many studies have
shown that inhibition of the cardiac renin–angiotensin system prevents geometric remodelling and LV dysfunction after MI.52 Experimental evidence indicates that the influence of ACE inhibitor treatment on LV remodelling after MI may involve both direct angiotensin II effects acting through a variety of angiotensin II receptor subtypes and indirect effects on the kininogen–kinin system.52 The observations made in this study suggest that ACE inhibitor treatment has a beneficial effect on cardiac remodelling without inhibiting angiotensin II effects through the AT1 receptor.

As indicated by echocardiography, mice with experimentally induced MI had significant systolic and diastolic dysfunction as shown by the major decrease of %FS and the increase in E:A. In this study, it was found that ACE inhibition significantly improved cardiac dysfunction in MI mice. Doppler echocardiography is the primary technique for evaluating LV diastolic function.22 Increased E wave velocity, decreased peak A wave velocity (or absent A wave), and rapid E wave deceleration were observed in the mice, and these flow patterns were similar to transmitral flow profiles obtained at the level of the papillary muscles for consistency. Note the prominent increase in LV diastolic dimension in the mice with the MI. ACE inhibitors attenuated the deterioration of LV function and remodelling in animals with chronic heart failure caused by MI.16 This effect was either blocked by a B2 kinin receptor antagonist17 or blunted in rats with kininogen deficiency due to spontaneous mutation of the kininogen gene,27 indicating that kinins are important in the cardioprotective mechanism of ACE inhibitors. The mechanism explaining the potential antiremodelling action of bradykinin may relate to

![Image of echocardiograms](http://heart.bmj.com/)

**Figure 3** Examples of two dimensional echocardiograms (parasternal short axis view) from a WT-control mouse, a KO-control mouse, a WT-MI mouse, a KO-MI mouse treated with ACEI, and a KO-MI mouse treated with ACEI. Short axis images were obtained at the level of the papillary muscles for consistency. Note the prominent increase in LV diastolic dimension in the mouse with the MI. ACE decreased LV diastolic dimensions in WT-MI and KO-MI mice.

| Table 2 Doppler echocardiographic measurements in control mice and MI mice |
|---------------------------------|-----------------|-----------------|-----------------|-----------------|-----------------|-----------------|-----------------|-----------------|-----------------|-----------------|-----------------|-----------------|-----------------|-----------------|-----------------|
|                                | WT-control:    | KO-control:     | WT-MI:           | KO-MI:          |
|                                | ACEI (-)       | ACEI (-)        | ACEI (-)         | ACEI (+)        | ACEI (-)        | ACEI (+)        | ACEI (-)        | ACEI (+)        | ACEI (-)        | ACEI (+)        | ACEI (-)        | ACEI (+)        | ACEI (-)        | ACEI (+)        |
| LVEDD (mm)                     | 3.6 (0.1)      | 3.8 (0.1)       | 5.2 (0.2)**      | 4.5 (0.1)**††   | 4.6 (0.1)††     | 4.0 (0.1)††     | 4.0 (0.1)††     | 4.0 (0.1)††     | 4.0 (0.1)††     | 4.0 (0.1)††     | 4.0 (0.1)††     | 4.0 (0.1)††     | 4.0 (0.1)††     |
| LVEDD (mm)                     | 2.9 (0.20)     | 3.0 (0.2)       | 4.8 (0.2)**      | 3.8 (0.2)**††   | 3.8 (0.2)**††   | 3.4 (0.2)**††   | 3.4 (0.2)**††   | 3.4 (0.2)**††   | 3.4 (0.2)**††   | 3.4 (0.2)**††   | 3.4 (0.2)**††   | 3.4 (0.2)**††   | 3.4 (0.2)**††   |
| %FS                            | 32 (3)         | 28 (3)          | 10 (2)**         | 16 (2)**††      | 15 (1)**††      | 20 (1)**††      | 20 (1)**††      | 20 (1)**††      | 20 (1)**††      | 20 (1)**††      | 20 (1)**††      | 20 (1)**††      | 20 (1)**††      |
| E wave (cm/s)                  | 41 (4)         | 46 (4)          | 84 (3)**         | 58 (3)**††      | 63 (3)**††      | 42 (3)**††      | 42 (3)**††      | 42 (3)**††      | 42 (3)**††      | 42 (3)**††      | 42 (3)**††      | 42 (3)**††      | 42 (3)**††      |
| A wave (cm/s)                  | 34 (3)         | 32 (4)          | 11 (2)**         | 26 (2)**††      | 23 (2)**††      | 24 (2)††        | 24 (2)††        | 24 (2)††        | 24 (2)††        | 24 (2)††        | 24 (2)††        | 24 (2)††        | 24 (2)††        |
| E/A                            | 1.7 (0.2)      | 1.8 (0.2)       | 7.8 (1.2)**      | 3.4 (0.4)**††   | 3.6 (0.3)**††   | 2.5 (0.4)       | 2.5 (0.4)       | 2.5 (0.4)       | 2.5 (0.4)       | 2.5 (0.4)       | 2.5 (0.4)       | 2.5 (0.4)       | 2.5 (0.4)       |
| DR (cm/s²)                     | 877 (77)       | 922 (65)        | 1675 (109)**     | 1356 (53)**††   | 1256 (59)**††   | 989 (44)**†††   | 989 (44)**†††   | 989 (44)**†††   | 989 (44)**†††   | 989 (44)**†††   | 989 (44)**†††   | 989 (44)**†††   | 989 (44)**†††   |

Values are mean (SEM).

DR, deceleration rate; %FS, percentage of fractional shortening; LVEDD, left ventricular diastolic dimension; LVESD, left ventricular end systolic dimension.

*p < 0.05 vs WT-control; **p < 0.01 vs WT-control; †p < 0.05 vs WT-MI ACEI (-); ††p < 0.01 vs WT-MI ACEI (-); ‡p < 0.05 vs KO-control; §p < 0.01 vs KO-control; †‡p < 0.05 vs WT-MI ACEI (-); ††‡p < 0.01 vs WT-MI ACEI (-); †††p < 0.01 vs KO-MI ACEI (-).
increased nitric oxide synthesis or an effect on prostaglandin metabolism. However, further work is needed to elucidate in more detail the mechanisms responsible for the beneficial effects of ACE inhibitors in LV remodelling with AT1 KO mice after MI.

ACE inhibition decreased blood pressure in WT-MI mice and prevented myocardial remodelling. On the other hand, ACE inhibition did not change blood pressure in KO-MI mice. However, ACE inhibition prevented the increases of ANP, BNP, and collagen mRNAs. Myocardial remodelling is regulated by mechanical and neurohumoral factors. It is well known that the effect of ACE inhibition on blood pressure is related to myocardial remodelling. Our data suggest that the direct effects of ACE inhibition on the local myocardial renin–angiotensin system may have an important role in preventing myocardial remodelling.

There is a common flaw in any study that uses the knockout model. The gene being knocked out may regulate a multiplicity of processes outside the targeted gene. In this model, chronic hypotension was observed in the heterozygous and homozygous mutant mice compared with WT littermates, and the amounts of renin mRNA in the kidney and plasma renin activity were greatly increased only in the homozygous mutant mice. The results of this experiment need to be carefully considered in light of non-AT1 receptor or other effects. We cannot distinguish between the direct effects of non-AT1 receptors on myocardial remodelling and the effects of these receptors on multiple processes outside of the heart and their subsequent indirect effects on myocardial remodelling.

In conclusion, we observed the ability of ACE inhibitors to prevent LV remodelling after experimentally induced MI in AT1 KO mice, thereby showing that non-AT1 receptor mediated mechanisms have an important role.
STAMPS IN CARDIOLOGY

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