

195

FUNCTIONAL DEFECTS IDENTIFIED IN OUTGROWTH ENDOTHELIAL CELLS FROM DIABETIC PATIENTS COMPARED TO HEALTHY CONTROLS

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10.1136/heartjnl-2016-309890.195

Background Circulating endothelial progenitor cells (EPCs) are thought to play a pivotal role in endothelial repair. Clinical trials utilising EPCs to promote therapeutic angiogenesis are already underway. Previous reports suggest a reduced EPC number and impaired functional activity in patients with type 2 diabetes mellitus. However, the actual identity of the cell type involved and the functional role played in the repair process needs to be further defined before EPCs can be successfully utilised in the clinic. In the present study, we assessed the functional capacity of circulating late out-growth endothelial progenitor cells (OECs) to further assess the contribution of the diabetic environment to diminished OEC function in the context of wound healing.

Methods and results OECs were isolated from 7 diabetic patients, presenting with ulcers ranging in size from 60 mm³–2500 mm³. OECs were characterised as CD34⁺, CD31⁺, vWF⁺, positive for AcLDL and UEA uptake and negative for the hematopoietic marker CD45 by immunohistochemistry. Migration of CD34⁺ OECs, evaluated by a scratch assay, demonstrated that migration is impaired in diabetic OECs compared to healthy control OECs, with 40–42% closure *vs* 100% over 24 h respectively. In addition, a chemotaxis transwell migration assay showed a decreased response to SDF-1 by diabetic cells *vs* healthy OECs. An angiogenesis tube formation assay also established a reduced capacity of the diabetic OECs to form an endothelial network as compared to healthy OECs ($p < 0.0001$), measured using the closed loop perimeter, and the number of branch points ($p < 0.05$). Nitrite concentrations were measured using a Griess assay, and the results showed that diabetic OECs produced less nitric oxide as compared to healthy OECs ($p < 0.0005$).

Conclusion OECs from diabetic patients show an impaired migration and response to chemotactic agents *in vitro* compared to OECs isolated from healthy controls. In addition, the reduced nitric oxide bioavailability found by diabetic cells may contribute to OEC dysfunction in diabetes. Future work will focus on assessing the secretome of healthy *vs* diabetic OECs.

196

STRATEGIES FOR INHIBITING ADVANCED GLYCATION ENDPRODUCT (AGE) INDUCED VASCULAR CALCIFICATION IN A SMOOTH MUSCLE CELL CULTURE MODEL

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10.1136/heartjnl-2016-309890.196

Vascular calcification is implicated in a range of cardiovascular disease mechanisms, leading to an associated increase in

morbidity and mortality. One such trigger are advanced glycation endproducts (AGEs), the tissue accumulation of which increases with age and is more prevalent in diabetic subjects due to oxidative stress and poor glycaemic control. The aim of this study was to investigate the osteogenic potential of AGEs and elucidate mechanisms of inhibiting these processes in a smooth muscle cell (SMC) culture model.

Osteogenic differentiation of SMCs was induced using Î²-glycerophosphate (Î²-GP), carboxymethyllysine (CML), carboxyethyllysine (CEL), methylglyoxal (MGO) and glycated low density lipoprotein (gly-LDL). The cells were subsequently treated with aminoguanidine (AG), an inhibitor of AGE formation, and novel glycomimetic compounds in order to determine their anti-calcification potential *in vitro* using qPCR, ELISA, Alkaline phosphatase (ALP) activity and Alizarin red staining.

Gly-LDL (10 µg/ml) and CML (2.5nM) increased the level of calcification observed compared to the Î²-GP (5 mM) positive control after 21 days ($p < 0.05$), with gly-LDL induced calcification apparent after 14 days. Both AG (250 µM) and the novel glycomimetic compounds reduced the level of mineralisation observed at 21 days compared with osteogenic treatments ($p < 0.05$). CEL (2.5 nM) and MGO (0.1 mM) both induced calcification, however mineralization was not as extensive as with Î²-GP. When compared to the structure of CML, the side-chain of CEL contains an extra methyl group, suggesting this group impacts RAGE receptor binding. It was also shown that Î²-GP combined with increased glucose concentration induced more extensive calcification unlike low glucose levels and Î²-GP alone. ALP activity, when stimulated with Î²-GP, CML and gly-LDL was greater at day 4 than at day 7, with AG reducing ALP activity measurements at day 4. Gly-LDL increases gene expression of OCN at day 4 compared with Î²-GP and CML, however this was reduced at day 7, corresponding with an increased expression of OPN and OPG. NOTCH-3 gene expression was also reduced at day 7. Gene expression of OPN, OPG and NOTCH-3 were reduced at both day 4 and day 7 compared with osteogenic treatments (Î²-GP, CML and gly-LDL).

In summary, we conclude that gly-LDL and CML are potent inducers of calcification compared with Î²-GP, and that their osteogenic potential can be modulated by both AG and novel glycomimetic compounds.

197

MIRNA-22 REGULATES VASCULAR SMOOTH MUSCLE CELL FUNCTIONS AND PREVENTS NEOINTIMA FORMATION BY TARGETING EVI-1

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10.1136/heartjnl-2016-309890.197

Background/objective We have recently reported that microRNA-22 (miR-22) regulates vascular smooth muscle cell (VSMC) differentiation from stem cells *in vitro* and *in vivo*. However, little is known about the functional involvements of miR-22 in VSMC functions and vessel injury-induced neointima formation. In the current study, we aimed to establish the causal role of miR-22 and its target genes in VSMC proliferation, migration and neointima lesion formation.

Methods and results miR-22 was significantly down-regulated in VSMCs upon various pathological stimuli, and in injured arteries. miR-22 over-expression in serum-starved VSMCs