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Prevalence and determinants of atrial fibrillation progression in paroxysmal atrial fibrillation

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ABSTRACT

Objective Atrial fibrillation (AF) often progresses from paroxysmal AF (PAF) to more permanent forms. To improve personalised medicine, we aim to develop a new AF progression risk prediction model in patients with PAF.

Methods In this interim-analysis of the Reappraisal of AF: Interaction Between HyperCoagulability, Electrical Remodelling, and Vascular Destabilisation in the Progression of AF study, patients with PAF undergoing extensive phenotyping at baseline and continuous rhythm monitoring during follow-up of ≥ 1 year were analysed. AF progression was defined as (1) progression to persistent or permanent AF or (2) progression of PAF with $>3\%$ burden increase. Multivariable analysis was done to identify predictors of AF progression.

Results Mean age was 65 (58–71) years, 179 (43%) were female. Follow-up was 2.2 (1.6–2.8) years, 51 of 417 patients (5.5%/year) showed AF progression. Multivariable analysis identified, PR interval, impaired left atrial function, mitral valve regurgitation and waist circumference to be associated with AF progression. Adding blood biomarkers improved the model (C-statistic from 0.709 to 0.830) and showed male sex, lower levels of factor XIIa:C1-esterase inhibitor and tissue factor pathway inhibitor, and higher levels of N-terminal pro-brain natriuretic peptide, proprotein convertase subtilisin/kexin type 9 and peptidoglycan recognition protein 1 were associated with AF progression.

Conclusion In patients with PAF, AF progression occurred in 5.5%/year. Predictors for progression included markers for atrial remodelling, sex, mitral valve regurgitation, waist circumference and biomarkers associated with coagulation, inflammation, cardiomyocyte stretch and atherosclerosis. These prediction models may help to determine risk of AF progression and treatment targets, but validation is needed.

Trial registration number NCT02726698.

INTRODUCTION

Atrial fibrillation (AF) is a progressive disease, usually starting with self-terminating short-lasting

WHAT IS ALREADY KNOWN ON THIS TOPIC

- ⇒ Atrial fibrillation progression is associated with adverse cardiovascular outcome.
- ⇒ The rate of atrial fibrillation progression varies and depends among others on type of rhythm monitoring.
- ⇒ Predictors of atrial fibrillation progression have not been well established with long-term continuous rhythm monitoring.

WHAT THIS STUDY ADDS

- ⇒ This study develops an atrial fibrillation progression risk prediction model and elucidates underlying pathophysiological mechanisms in comprehensively phenotyped patients with paroxysmal atrial fibrillation using long-term continuous rhythm monitoring.
- ⇒ Our clinical multivariate model had a C-statistic of 0.709.
- ⇒ The addition of the blood biomarkers improved the initial model to a C-statistic of 0.830.
- ⇒ We found that predictors for progression were multifactorial including atrial remodelling, sex, mitral valve regurgitation, waist circumference and blood biomarkers associated with coagulation, cardiac stretch, cholesterol metabolism, inflammation and the immune system.
- ⇒ Validation is needed before implementation into clinical practice.

paroxysmal episodes that often progresses to more frequent episodes, eventually leading to long-lasting non-self-terminating persistent and permanent AF.¹ Progression of AF has been associated with an increased risk of cardiovascular morbidity and mortality and reduces the efficacy of pharmacological and interventional rhythm control strategies.^{2,3} AF progression rates vary between studies because of differences in duration of follow-up, in comprehensive phenotyping of patients and strategies of rhythm monitoring.^{3–5}

HOW THIS STUDY MIGHT AFFECT RESEARCH, PRACTICE OR POLICY

- ⇒ This model could be clinically useful, and serves to enhance knowledge on underlying mechanisms causing progression of atrial fibrillation.
- ⇒ In combination with extensive phenotyping, our prediction model gives a more in-depth view into predicting risk factors for atrial fibrillation progression.
- ⇒ Continuous rhythm monitoring provides a more detailed and accurate view into atrial fibrillation progression.

Appropriate treatment of risk factors can improve sinus rhythm maintenance, cardiovascular outcome and reverse AF progression.^{6,7} The most established risk factors for AF progression are age, hypertension, obesity, heart failure and diabetes.^{5,8} Interestingly, hypercoagulability may be involved in increasing the risk of stroke and in AF progression.⁹ A detailed and multimodal phenotyping at baseline and continuous rhythm monitoring has potential to increase our knowledge of AF progression and in turn contribute to personalised medicine.^{2,10}

Therefore, the aim of the Reappraisal of AF: Interaction Between HyperCoagulability, Electrical Remodelling, and Vascular Destabilisation in the Progression of AF (RACE V) study is to develop a clinical AF progression risk prediction model using extensive phenotyping and continuous rhythm monitoring in patients with paroxysmal AF (PAF). In addition, to improve the clinical model and elucidate underlying pathophysiological mechanisms of AF progression, we included blood biomarkers in the progression risk prediction model.

METHODS**Study design**

The RACE V study has previously been described.¹¹ In brief, the RACE V study is a prospective, investigator-initiated, Dutch multicentre observational study (Clinicaltrials.gov identifier NCT02726698).

A detailed overview of inclusion and exclusion criteria is provided in online supplemental table S1. Briefly, the aim is to include 750 patients with a history of PAF <10 years. Eligible patients had ≥ 2 documented episodes of PAF or one documented episode combined with ≥ 2 symptomatic episodes suspected of being AF, were willing to undergo implantation of a Medtronic (Minneapolis, USA) Reveal LINQ[®] implantable loop recorder, and did not have a history of persistent AF (intention to undergo), pulmonary vein isolation (PVI) or current amiodarone treatment. Patients with Medtronic pacemakers were also eligible if atrial high rate episodes >190 beats per min lasting >6 min, qualified as AF episodes, were detected. For the current analysis, we included patients that had ≥ 1 year of continuous rhythm monitoring as of 1 May 2020.

Patient and public involvement

Patients and the public were not involved in the design or implementation of the study.

Clinical assessment

At baseline, clinical history, symptomatology, current medication, physical examination and a 12-lead ECG were assessed. Additionally, echocardiography, vascular assessment and cardiac CT was done, processed and analysed in a central core lab (online supplemental figure S1, online supplemental data). In brief, in

addition to the standard echocardiography measurements, strain measurements were performed in sinus rhythm using a point-and-click method to trace endocardial borders with a vendor-independent software (TOMTEC-ARENA, Imaging Systems, Germany). The cardiac CT was performed as a non-contrast ECG-gated scan to assess coronary calcium scores, epicardial and pericardial fat. Vascular assessment of the carotid arteries included measurements of intima-media thickness, pulse wave velocity and plaques.

Blood biomarkers

At baseline, peripheral blood samples were collected (only during sinus rhythm with interrupted anticoagulation). With multiplex immunoassays, 92 cardiovascular biomarkers from the Olink Cardiovascular III panel were assessed by Olink Bioscience (Uppsala, Sweden) in EDTA plasma baseline samples (online supplemental table S2). Complexes of activated coagulation enzymes (FXIIa, FXIa, FIXa, FXa and thrombin) with their corresponding natural inhibitors (antithrombin, alpha1-antitrypsin or C1-esterase inhibitor) ELISA assays were performed to assess the degree coagulation activity in EDTA plasma and citrated plasma samples at baseline.¹²

Follow-up

All patients were treated according to the European Society of Cardiology AF guidelines.¹³ Follow-up visits were performed at 1 and 2.5 years (online supplemental figure S1). Patients could consent for 2.5 years continuous rhythm monitoring, until end of battery of Reveal LINQ, or for 4 years in case patients had a pacemaker.

In order to collect continuous data on arrhythmias, all patients received a home monitoring device (Medtronic Carelink). Both Reveal LINQ and pacemaker were set to AT/AF detection settings (online supplemental data).

Definition and outcome

The primary outcome was AF progression. Before assessing AF progression, all collected episodes were independently adjudicated and corrected by five physicians. Two methods were used to assess AF progression and compared. For the first method, all AF episodes were put into a custom-made software using Microsoft Visual Basic to visualise in a graphical overview all AF episodes per patient (figure 1), which was done by six physicians. Four groups were discerned: (1) no AF recurrences during follow-up; (2) recurrences of PAF without apparent increase in number and/or duration of AF episodes based on visual inspection; (3) recurrence of PAF with increase in number and/or duration of AF based on visual inspection, but without persistent or permanent AF; (4) development of persistent or permanent AF (figure 1).

For the second method, a mathematical formula (online supplemental data) was created using a weighed AF burden with AF episodes early during follow-up weighing less than AF episodes at the end of follow-up. AF burden was defined as the cumulative duration of all AF episodes from baseline onwards, divided by total duration of monitoring. For patients without successful PVI, a 90-day blanking period after PVI was applied.

The primary outcome was AF progression, defined as (1) development of persistent or permanent AF during follow-up or (2) an increase of >3% AF burden over the first 6 months or total follow-up. Duration of monitoring for current analysis lasted until 1 May 2020, until last available rhythm monitoring

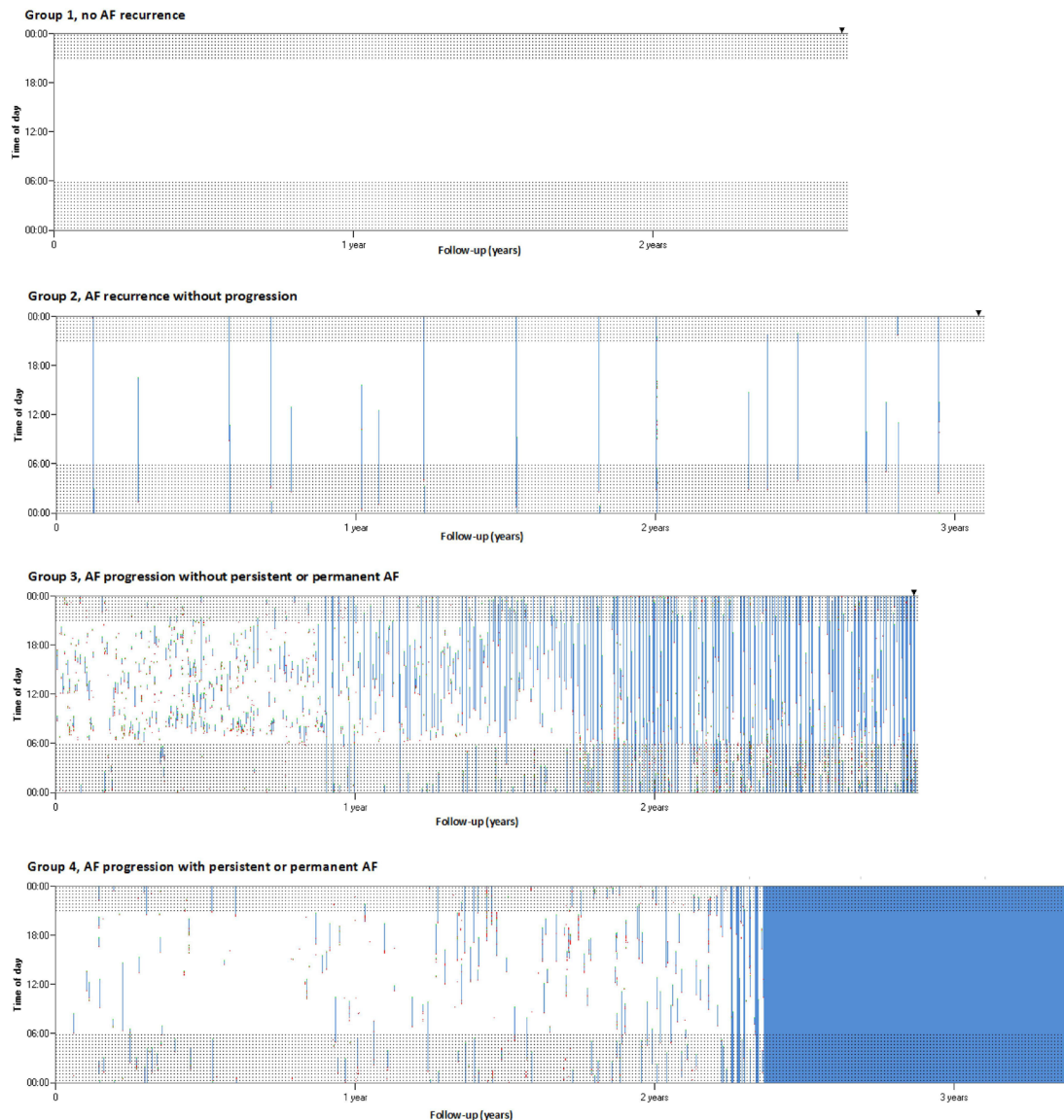


Figure 1 Examples of continuous rhythm monitoring. Examples of individual patients without AF progression (group 1 and group 2) and with AF progression (group 3 and 4) during follow-up. The X-axis presents follow-up in years, the Y-axis is the time of the day. Shaded areas indicate nightly hours. Black triangle presents day of end of analysis. White means no AF is present, and blue represents ongoing episodes of AF. AF initiations are shown in red and AF terminations are shown in green. AF, atrial fibrillation.

for patients that died after >1 year of continuous rhythm monitoring, until date of PVI, or in case of a successful PVI.

We considered the mathematical formula as leading of both methods, because it is easier to apply in other independent cohorts. Results from both methods were compared and showed that no patients classified as ‘without AF progression’ by physicians were ‘with AF progression’ according to the mathematical formula. Fourteen (3%) patients who were classified as AF progressors (from group 3) by physicians did not have AF progression according to the mathematical formula. These patients were eventually categorised as no AF progression.

Statistical analysis

Baseline characteristics are presented as mean±SD for normally distributed data, and median and iQRs for non-normally distributed continuous data. Categorical data are presented as numbers with percentages, biomarker multiplex immunoassay data as arbitrary units on a log2 scale. Fisher’s exact test was used for

binary variables, and T-test or Wilcoxon rank-sum test was used for continuous variables. Collected baseline variables including core lab data, with $p < 0.10$ in the age-adjusted and sex-adjusted logistic regression, with exception of European Heart Rhythm Association (EHRA) class, number of comorbidities, CHA₂DS₂-VASC score and medications, were included in a bidirectional stepwise variable selection leading to a final multivariable logistic regression model. Bidirectional stepping was done for model building and reduction, with a p value ≥ 0.05 as a criterion for removing a variable from the model (online supplemental data). Imputation was implemented for missing values using the R package mice. For each logistic regression, ‘massive imputation’ was performed, which means that all variables in a model were at the same time also used for the imputation needed for the fit of that model. For the second model, the Olink Cardiovascular III panel biomarkers (online supplemental table S6) and coagulation markers were added to the stepwise variable selection process if they reached $p < 0.10$ in initial age-adjusted and sex-adjusted

logistic regression to assess if the model would improve. Age and sex were forced into both multivariate models. Interactions between variables was tested, no significant interactions were found. The Harrell's binary C-index was used for goodness-of-fit measure. P value <0.05 was considered statistically significant. Internal validation was done using bootstrapping. Analyses were conducted with R V.3.3.3 (www.r-project.org).

RESULTS

For the present analysis, we included 417 patients (table 1, online supplemental table S3). Median age was 65 (58–71) years, and 179 (43%) patients were women. Median follow-up of continuous rhythm monitoring was 2.2 (1.6–2.8) years. A total of 162 215 episodes were classified as AF by the automated algorithm, 53 397 (32.9%) were adjusted after adjudication, resulting in 119 120 remaining AF episodes (reasons for adjustments are presented in online supplemental data).

During follow-up, 48 (11.5%) patients showed no AF recurrences, and 318 (76.3%) had AF recurrences without AF progression. AF progression was seen in 51 (12.2%, 5.5% per year) patients: with an increase of >3% of AF burden but without deterioration into persistent or permanent AF in 16 (3.8%) patients, and with development of persistent or permanent AF in 35 (8.4%) patients (online supplemental table S4, figure 1).

Patients with AF progression were more often men, had more often coronary artery disease, larger waist circumference, longer PR interval, larger left atrial (LA) volume and reduced atrial contractile function (table 1).

During follow-up up, one patient died of an unknown cause. Eighteen patients received a pacemaker, 1 due to AV block and 17 patients due to sick sinus syndrome. Figure 2 presents rhythm control therapy during follow-up. No differences were seen in rhythm control therapy at baseline, follow-up and end of analysis between the AF progression and no AF progression group (online supplemental figure S2).

Blood biomarkers

The baseline levels of the 92 biomarkers are presented in online supplemental table S5). At baseline, a significant difference between the groups was observed for 14 biomarkers.

Baseline coagulation markers are presented in online supplemental table S6). At baseline, the levels of factor XIIa:C1-esterase inhibitor complex and factor XIIa:antithrombin were significantly lower in the AF progression group compared with those without AF progression.

Prediction models

The logistical analysis adjusted for age and sex with clinical variables showed that 14 variables were associated with AF progression (online supplemental table S7). The clinical multivariable model showed that a longer PR interval, an impaired LA contractile function, moderate mitral valve regurgitation and a higher waist circumference were associated with higher risk of AF progression (table 2A), C-statistic is 0.709 (95% CI 0.614 to 0.799). The optimism caused by overfitting in the C-statistic was 3.03%.

To improve the prediction model and to assess underlying pathophysiological mechanisms, an additional analysis including blood biomarkers was performed. The logistical analysis adjusted for age and sex showed 25 variables associated with AF progression (online supplemental table S8). Table 2B and figure 3 show the multivariable predictors of AF progression including blood

biomarkers. The addition of the blood biomarkers improved the initial model (C-statistic 0.830 (95% CI 0.750 to 0.898)).

Based on the clinical multivariable model, a point risk score was developed for estimating an individual's risk of AF progression at 2 years (table 3).

DISCUSSION

In the RACE V study, we assessed AF progression in comprehensively phenotyped patients with self-terminating PAF using long-term continuous rhythm monitoring. We showed that AF progression occurred in 5.5% of patients per year. Furthermore, using the clinical model markers of atrial remodelling, mitral valve regurgitation and waist circumference were associated with AF progression. The addition of blood biomarkers improved the C-statistic of the model and showed male sex, lower levels of coagulation markers and markers involved in cardiac stretch, cholesterol metabolism, inflammation and the immune system to be associated with AF progression.

Determining AF progression importantly depends on the type and amount of rhythm monitoring. Previous studies used limited rhythm monitoring and typically focused on progression from PAF to persistent or permanent AF.^{3,4} Yet, more studies suggest that increase of AF burden in PAF is also of importance.¹⁴ Therefore, we included increase of AF burden in our AF progression definition to avoid excluding patients with low burden that progressed to a significantly higher PAF burden. Although the majority of patients who showed AF progression deteriorated into persistent or permanent AF, 30% in the AF progression group were classified as progressors because of a likely clinically relevant increase of PAF burden.

In line with previous studies, we found multiple factors involved in AF progression associated with different underlying pathophysiological mechanisms.^{3,8} A longer PR interval and an impaired LA contractile function were associated with AF progression. Both can be seen as signs of more severe atrial structural remodelling (atrial cardiomyopathy) promoting AF progression.¹⁵ Previous studies showed that the PR interval was associated with incident AF but not with AF progression.¹⁶ Mitral valve regurgitation, well known to induce volume overload and LA enlargement and thus atrial remodelling, was also associated with AF progression.¹⁷ Atrial enlargement has been associated with incident and recurrent AF.¹⁸ Atrial contractility dysfunction has been related to duration of AF and may increase compliance of the atria, causing atrial cardiomyopathy, which is in turn associated with AF progression.¹⁹ A higher waist circumference was also associated with AF progression. A high body mass index (BMI) and obesity are well known risk factors for incident AF and AF progression.⁵ However, BMI does not take visceral fat distribution into account, which has been shown to be an independent marker for cardiovascular morbidities associated with AF and AF progression.⁵ Excess of visceral fat induces inflammation, which can promote atrial remodelling.²⁰ Waist circumference could therefore be seen as a marker of visceral adipose tissue and thus being associated with AF progression.

In addition to a clinical prediction model, we sought to explore the underlying pathophysiological mechanisms for AF progression adding 101 blood biomarkers including coagulation markers to our analysis. The latter improved the prediction model significantly. Furthermore, it revealed that N-terminal pro-brain natriuretic peptide (NTproBNP) was a marker for risk of AF progression. NTproBNP is secreted by myocytes in response to multiple factors, including wall stress and is increased during AF, even without overt heart failure. Our results are comparable

Arrhythmias and sudden death

Table 1 Baseline characteristics

Characteristic	AF progression (n=51)	No AF progression (n=366)	Total population (n=417)	P value
Age (years)	64 (60–73)	65 (58–71)	65 (58–71)	0.278
Female sex	15 (29%)	164 (45%)	179 (43%)	0.049
Total history AF (years)	2.8 (0.9–4.9)	2.6 (0.7–5.2)	2.6 (0.7–5.1)	0.803
Heart failure	20 (39%)	104 (28%)	124 (29%)	0.274
HFrEF	4 (8%)	6 (2%)	10 (2%)	0.025
HFpEF	16 (31%)	98 (27%)	114 (27%)	1
Hypertension	46 (90%)	292 (80%)	338 (81%)	0.086
Diabetes mellitus	5 (10%)	29 (8%)	34 (8%)	0.59
Coronary artery disease	11 (22%)	37 (10%)	48 (12%)	0.031
Atherosclerosis*	26 (51%)	178 (49%)	204 (49%)	0.767
Peripheral artery disease	2 (4%)	1 (0%)	3 (1%)	0.041
Ischaemic stroke	1 (2%)	18 (5%)	19 (5%)	0.491
Pacemaker	9 (18%)	16 (4%)	25 (6%)	0.001
Number of comorbidities†	3 (2–4)	2 (2–3)	2 (2–3)	0.05
CHA ₂ DS ₂ -VASC score‡				0.016
<2	6 (12%)	101 (28%)	107 (26%)	
≥2	45 (88%)	265 (72%)	310 (74%)	
Physical examination				
Height (cm)	178 (170–185)	177 (169–184)	178 (169–184)	0.492
Weight (kg)	88 (73–102)	84 (74–96)	85 (74–97)	0.268
Body mass index (kg/m ²)	27 (25–32)	27 (24–30)	27 (24–30)	0.708
Waist circumference (cm)	105 (99–113)	100 (92–108)	100 (93–108)	0.004
Laboratory results				
eGFR (mL/min/1.73 m ²)	74 (67–86)	81 (70–90)	81 (69–90)	0.016
ECG				
PR interval	178 (160–199)	164 (149–184)	166 (150–186)	0.003
QRS interval	96 (90–106)	94 (86–103)	94 (88–104)	0.191
Medications				
β-Blocker	32 (63%)	181 (49%)	213 (51%)	0.099
Verapamil/Diltiazem	7 (14%)	66 (18%)	73 (18%)	0.557
Digoxin	2 (4%)	4 (1%)	6 (1%)	0.16
Class I antiarrhythmic drugs	5 (10%)	89 (24%)	94 (23%)	0.02
Class III antiarrhythmic drugs	3 (6%)	15 (4%)	18 (4%)	0.473
ACE inhibitor	11 (22%)	71 (19%)	82 (20%)	0.709
Angiotensin receptor blocker	14 (27%)	66 (18%)	80 (19%)	0.129
Statin	26 (51%)	119 (33%)	145 (35%)	0.012
Anticoagulant	45 (88%)	244 (67%)	289 (69%)	0.002
Vitamin K antagonist	10 (20%)	45 (12%)	55 (13%)	0.182
NOAC	35 (69%)	199 (54%)	234 (56%)	0.07
Echocardiographic variables				
Left atrial volume index (mL/m ²)	34 (25–39)	29 (23–36)	29 (23–36)	0.038
Left atrial reservoir function (%)	31 (26–39)	37 (30–43)	36 (29–43)	0.045
Left atrial contractile function (%)	13 (11–17)	17 (13–22)	16 (13–21)	0.003
Left atrial conduction function (%)	18 (14–25)	19 (14–24)	19 (14–24)	0.965
Left ventricular ejection fraction (%)	50±8	51±8	51±8	0.893
Left ventricle strain	−14.2±2.5	−14.0±2.3	−14.0±2.4	0.76
Moderate aortic valve stenosis	0 (%)	3 (1%)	3 (1%)	1
Moderate aortic valve regurgitation	0 (%)	0 (%)	1 (0%)	1
Moderate mitral valve regurgitation	3 (6%)	4 (1%)	7 (2%)	0.045
CT				
Calcium score (Agatston)	131 (5–492)	25 (0–228)	29 (0–275)	0.004
Pericardial fat	186 (148–235)	166 (121–231)	168 (124–233)	0.205
Epicardial fat	105 (77–130)	98 (71–128)	98 (72–128)	0.349
Vascular assessment				
IMT max-CCA >1 mm	19 (46%)	109 (34%)	128 (35%)	0.122
IMT max-all segments >1 mm	20 (49%)	154 (48%)	174 (48%)	1
Plaques	15 (29%)	125 (34%)	140 (34%)	0.407

Data are presented as mean±SD, number of patients (%) or median (IQR).

*Atherosclerosis is presence of history of myocardial infarction, percutaneous coronary intervention, coronary artery bypass graft, ischaemic cerebral infarction, peripheral vascular disease, Agatston score >400 or plaque.

†The number of comorbidities was calculated by awarding points for hypertension, heart failure, age >65 years, diabetes mellitus; coronary artery disease, BMI >25 kg/m², moderate or severe mitral valve regurgitation and kidney dysfunction (eGFR <60).

‡The CHA₂DS₂-VASC score assesses thromboembolic risk. C=congestive heart failure/LV dysfunction, H=hypertension; A₂=age ≥75 years; D=diabetes mellitus; S₂=stroke/transient ischaemic attack/systemic embolism; V=vascular disease; A=age 65–74 years; Sc=sex category (female sex).

AF, atrial fibrillation; BMI, body mass index; CCA, common carotid artery; eGFR, estimated glomerular filtration rate; HFpEF, heart failure with preserved ejection fraction; HFrEF, heart failure with reduced ejection fraction; IMT, intima-media thickness; LV, left ventricular; NOAC, novel oral anticoagulation.

to previous studies showing that elevated NTproBNP levels are associated with incident AF and AF progression.^{5 10 21} Proprotein convertase subtilisin/kexin type 9 (PCSK9), also associated

with AF progression in our model, is an enzyme involved in the homeostasis of cholesterol. Higher levels of PCSK9 are associated with cardiovascular events in patients with AF, possibly



Figure 2 Flow chart of all patients. Four-hundred seventeen patients were included in current analysis. One patient died during follow-up and was included in the analysis until last rhythm monitoring date. AAD, anti-arrhythmic drugs; AF, atrial fibrillation; ECV, electrical cardioversion; PVI, pulmonary vein isolation.

through atherosclerosis and inflammation.²² Peptidoglycan recognition protein 1 (PGLYRP1), a protein important in the innate immune response, was also associated with AF progression. PGLYRP1 is also involved in inflammation and associated with atherosclerosis. Elevated levels of PGLYRP1 have been associated with aortic wall thickness, aortic plaques and elevated Agatston scores.²³ The fact that PCSK9 and PGLYRP1 were markers for AF progression suggests that vascular processes are of importance in AF progression.^{24 25}

Lastly, lower levels of TFPI were associated with progression. This indicates that there is less inhibition of the extrinsic coagulation cascade in patients with AF progression due to reduced TFPI, resulting in increased activity of tissue factor and factor VIIa, and thus increased activation of the extrinsic coagulation pathway. Also, lower levels of factor XIIa:C1-esterase inhibitor, an enzyme inhibitor complex of the intrinsic coagulation cascade, were associated with AF progression. The origin of both and the role in AF progression remains unknown, but the postulated enhanced potential of tissue factor stimulated coagulation, due to lower TFPI activity, by itself would be in accordance with

a role of hypercoagulability in driving AF as previously shown in preclinical studies.⁹ Recently, it was shown that duration of PAF was associated with higher levels of von Willebrand factor and factor VIII.²⁶ Clearly, more research is warranted on the role of hypercoagulability in AF progression.

The model with additional biomarkers also revealed, unexpectedly, male sex as a clinical marker associated with AF progression. None of the previous studies showed sex differences involved in AF progression but data are still scarce.^{3 5 10} In our study, the percentage of females was 43%, higher than in most studies. Interestingly, women with AF are usually older, having more comorbidities.^{27 28}

In summary, our models, including the point risk score, may help to identify patients at risk for AF progression. It again emphasises that AF progression is a multifactorial disease and also suggests differences between sexes. The RACE V clinical risk score may contribute to determine individuals' risks of AF progression and treatment targets. However, before introduction into clinical practice it first warrants validation. As a result, such a model may increase the complexity and burden for the

Table 2 (A) Multivariable clinical predictors for AF progression

	OR	95% CI	P value
Male sex	1.8	0.87 to 3.51	0.116
PR interval (per SD)	1.5	1.14 to 2.06	0.004
Impaired left atrial contractile function (per SD)	1.8	1.16 to 2.69	0.008
Moderate mitral valve regurgitation	5.9	1.02 to 33.97	0.048
Waist circumference (per SD)	1.5	1.06 to 2.03	0.023

(B) Multivariable predictors of AF progression including blood biomarkers

	OR	95% CI	P value
Male sex	3.5	1.65 to 7.41	0.001
PR interval (per SD)	1.6	1.21 to 2.21	0.002
Impaired left atrial contractile function (per SD)	1.7	1.05 to 2.70	0.031
Factor XIIa:C1-esterase inhibitor (below median)	2.7	1.26 to 5.56	0.01
TFPI decrease (per SD)	1.8	1.23 to 2.53	0.002
NTproBNP (per SD)	1.9	1.28 to 2.81	0.002
PCSK9 (per SD)	1.6	1.09 to 2.21	0.015
PGLYRP1 (per SD)	1.5	1.11 to 2.11	0.009

AF, atrial fibrillation; NTproBNP, N-terminal pro-brain natriuretic peptide; PCSK9, proprotein convertase subtilisin/kexin type 9; PGLYRP1, peptidoglycan recognition protein 1; TFPI, tissue factor pathway inhibitor.

physician. The Horizon 2020 EHRA-PATHs project aims to develop a software tool that may contribute to improve the feasibility of such a personalised therapeutic strategy.²⁹

LIMITATIONS

Our study has several limitations. First, in the RACE V study treatment was at the discretion of the treating physician, which

may have influenced AF progression. However, although low numbers, we did not find significant differences in rhythm control therapy during follow-up between the groups. The clinical risk model obviously depends on the population included in the trial. Second, the existence of missing values in the data, which might have impacted the model, although we used multiple imputation to use the non-missing part of the data as much as possible as opposed to the removal of information from the analysis when doing regression analyses only with patients with complete information only. Third, follow-up was a median of 2.2 years, and did not met the calculated sample size of 750 patients and expected 187 AF progression events, due to a slow inclusion rate as result of the COVID-19 pandemic. To further assess AF progression, more patients and longer follow-up is needed. Fourth, for validation of the models and risk score an impact study is needed before use in clinical practice. However, according to our knowledge such a cohort with continuous rhythm monitoring and thorough phenotyping is not yet available. Fifth, not all factors contributing to AF progression may have been systematically, and repeatedly, assessed in RACE V. Finally, we did not implement temporal dynamic risk profiling into our study.³⁰

CONCLUSION

The RACE V study shows that AF progression in patients with PAF occurs in 5.5% per year as assessed with continuous rhythm recording. Clinical predictors for AF progression included markers of an atrial cardiomyopathy, higher waist circumference and male sex. The addition of cardiovascular biomarkers improved the risk prediction model and showed that increased levels of markers for atrial remodelling,

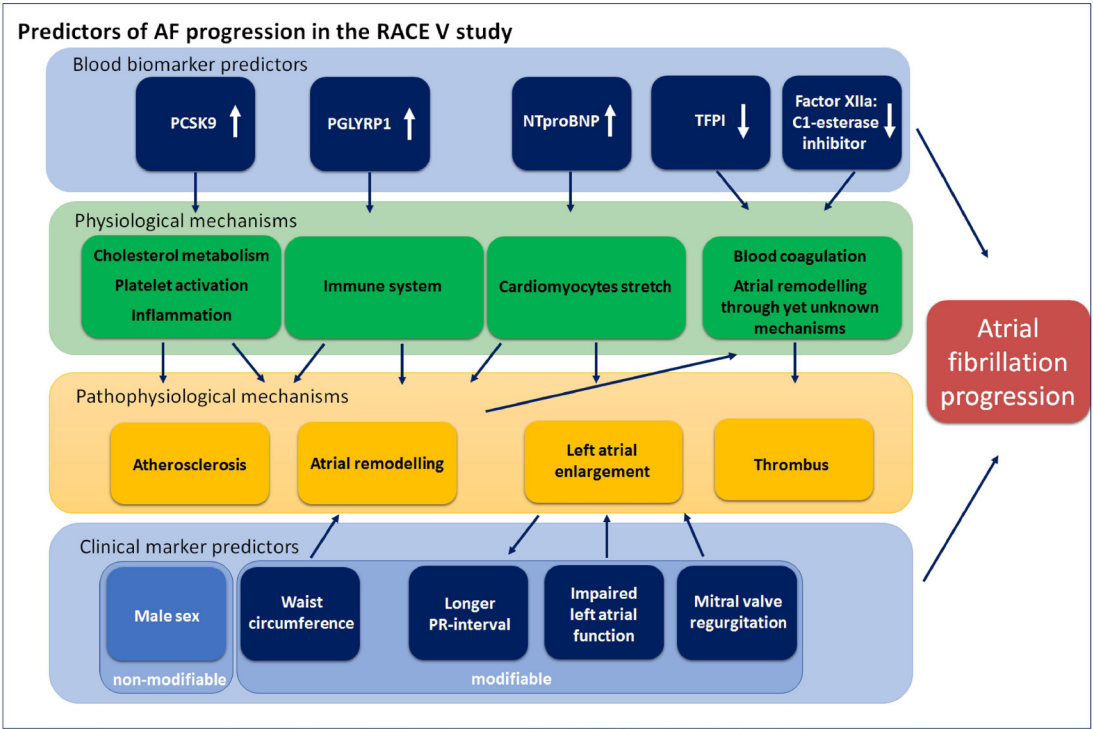


Figure 3 Predictors of atrial fibrillation (AF) progression in the Reappraisal of AF: Interaction Between HyperCoagulability, Electrical Remodelling, and Vascular Destabilisation in the Progression of AF study. Clinical markers and blood biomarkers as predictors for atrial fibrillation progression and their physiological and pathophysiological mechanisms. The blue boxes represent the multivariable predictors of atrial fibrillation progression. The green boxes represent the physiological mechanisms, the yellow boxes represent the pathophysiological mechanisms. NTproBNP, N-terminal pro-brain natriuretic peptide; PCSK9, proprotein convertase subtilisin/kexin type 9; PGLYRP1, peptidoglycan recognition protein 1; TFPI, tissue factor pathway inhibitor.

Table 3 Clinical point risk score

Sex											Points
Female											−2
Male											0
PR interval (ms)											Points
≤122											0
123–148											1
149–174											2
175–200											3
201–226											4
227–252											5
253–278											6
279–304											7
305–330											8
331–356											9
>356											10
Left atrial contractile function (%)											Points
≤12											3
13–17											2
18–22											1
23–27											0
28–32											-1
>32											-3
Waist circumference											Points
≤84											0
85–96											1
97–108											2
109–120											3
121–132											4
>132											5
Mitral valve regurgitation											Points
Yes											5
No											0
2-year risk estimation atrial fibrillation progression based on total points											
Total points	0	1	2	3	4	5	6	7	8	9	10
Risk %	2.3	3.1	4.2	5.6	7.5	9.9	13.0	16.9	21.6	27.2	33.7
AF, atrial fibrillation.											

AF, atrial fibrillation.

inflammation, atherosclerosis and coagulation were predictive for AF progression.

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Supplementary Data

Prevalence and determinants of AF progression in self-terminating atrial fibrillation - data from RACE V.

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Core lab measurement methods

Reveal LINQ and pacemaker arrhythmia episode adjustments

All collected episodes continuous data on arrhythmias of the included patients up until 1 May 2020 saved by the Reveal LINQ and pacemaker were independently adjudicated and corrected by 5 physicians. Any episode of AF ≥ 2 minutes was automatically detected. Arrhythmias with ≥ 182 beats per minute (cycle length ≤ 330 ms) with a duration of ≥ 24 beats were automatically classified as tachycardia. Arrhythmias with ≤ 30 beats per minute (cycle length ≥ 2000 ms), lasting 12 beats were automatically classified as bradycardia. An asystole ≥ 4.5 seconds was classified as a pause. The most common reasons for adjustments were false positive AF episodes (premature atrial or ventricular complexes or artefacts), on-going episodes, and episodes classified as atrial tachycardia instead of AF.

Echocardiography

Echocardiography recordings were anonymized and transferred to a core-lab facility for further analysis. Strain analysis was conducted offline, during one cardiac cycle, in sinus rhythm by one experienced observer blinded to clinical data and outcomes. Analysis was performed using vendor-independent software (TOMTEC-ARENA, Imaging Systems, Germany). LV global longitudinal strain (GLS) was analysed in the apical two-, three- and four-chamber views. Left atrial, right atrial and right ventricular strain were assessed in apical four-chamber view only. The region of interest was determined by a manual point-and-click method to trace endocardial borders during LV end-systolic frame. End-systole was automatically defined by the software and was manually adjusted for accuracy when needed. The software automatically produced myocardial speckle tracking in each frame during one cardiac cycle (RR-interval). Atrial contractile function measurements were performed by setting the base or zero strain reference at left ventricular end-diastole. Therefore, left atrial (LA) reservoir strain was measured as difference of the strain value at mitral valve opening minus the zero strain reference. LA contractile function was measured as the difference of the peak strain value at the onset of atrial contraction minus the zero strain reference.

LA conduit was measured as the difference of the strain value at mitral valve opening (LA reservoir) minus the peak strain value at the onset of atrial contraction.

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Vascular assessment

Vascular assessment of the carotid arteries included measurements of intima media thickness (IMT), pulse wave velocity (PWV) and plaques. PWV was assessed by Complior (Alam Medical, France) or SphygmoCor (Atcor Medical Blood Pressure Analysis System, Australia) at the carotid and femoral arteries. The aortic PWV was determined by using ≥ 20 consecutive pressure waveforms at the carotid and femoral artery. The wave transit time was calculated by the system software using the R-wave from the simultaneous ECG recording. of the simultaneously recorded ECG. Distance between both measure points was determined and corrected by multiplying the distance by 0.8. The PWV was calculated by dividing the distance between the femoral and carotid artery by the wave transit time. IMT and presence of plaques was assessed by ultrasound (Siemens Acuson S2000) with the Syncho US Workplace 3.5, Arterial Health Package for automated IMT measurement. Assessment of the IMT was done bilaterally in the common carotid artery, the carotid bifurcation, and internal carotid artery.

Cardiac computed tomography (CT)

Epicardial fat was measured on ECG-triggered, native CT heart scans according to the methodology introduced by Fox et al.(1). Tube voltage of scan protocols varied between 80–120kV. The region of interest (ROI) was defined as described by Versteyslen(2): The cranial slice limit was set at the level of the

carina of the pulmonary artery, and the caudal slice limit was the last slice containing any portion of the heart. The anterior border of the ROI was defined by the sternum, the posterior border by the ribs and vertebral column. Images were reconstructed using a soft-tissue algorithm. The pericardium was traced by a blinded reader placing 5-7 control points per slice using axial views as described earlier. Afterwards Catmull-Rom cubic spline functions are then automatically generated to obtain a smooth closed pericardial contour. Ultimately fat was automatically summed with a dedicated volumetric software (syngo.via Frontier, Cardiac risk assessment package, Siemens Healthineers, Forchheim, Germany). Epicardial and pericardial fat were defined as previously described by Iacobellis: Epicardial fat is located between the outer wall of the pericardium and the visceral layer of the pericardium. Pericardial fat is localized between visceral and pericardial myocardium(3).

1. Fox CS, Gona P, Hoffmann U, Porter SA, Salton CJ, Massaro JM, et al. Pericardial fat, intrathoracic fat, and measures of left ventricular structure and function: the Framingham Heart Study. *Circulation*. 2009;119(12):1586-91.
2. Versteylen MO, Takx RA, Joosen IA, Nelemans PJ, Das M, Crijs HJ, et al. Epicardial adipose tissue volume as a predictor for coronary artery disease in diabetic, impaired fasting glucose, and non-diabetic patients presenting with chest pain. *Eur Heart J Cardiovasc Imaging*. 2012;13(6):517-23.
3. Iacobellis G. Epicardial and pericardial fat: close, but very different. *Obesity (Silver Spring)*. 2009;17(4):625; author reply 6-7.

Blood biomarkers

At baseline peripheral blood samples were collected. Patients needed to be in sinus rhythm during blood sampling and oral anticoagulation was temporarily interrupted. All blood samples were processed and stored at -80°C.

Clinical definitions used in this study

Heart Failure definition

At baseline presence of:

- 1) history of heart failure admission;
- 2) left ventricular ejection fraction $\leq 45\%$;
- 3) left ventricular ejection fraction $>45\%$,
with either signs of:
 - structural heart disease (left ventricular hypertrophy [left ventricular mass index >95 g/m² in women and >115 g/m² in men] OR posterior wall thickness ≥ 11 mm OR septal wall thickness ≥ 11 mm)
 - and/or signs of diastolic dysfunction (mean E' velocity <8 cm/s & deceleration time >220 ms & E/e' >8).

Heart Failure with Preserved Ejection Fraction (HFpEF) definition

- combination of LVEF $>45\%$ + structural heart disease
- and/or
- combination of LVEF $>45\%$ + diastolic dysfunction

Hypertension definition

At baseline presence of:

- History of hypertension
- Use of a beta blocker, with exception of not daily used.
- Use of any calcium channel blocker
- Use of any ACE-inhibitor
- Use of any angiotensin receptor blocker
- Use of any diuretic, including mineralocorticoid receptor antagonist, excluding furosemide amiloride and bumetanide use.
- Use of an alpha blocker
- Baseline blood pressure $>140/90$ mmHg.

Atherosclerosis definition

At baseline presence of:

- history of myocardial infarction
- history of percutaneous coronary intervention
- history of coronary artery bypass graft
- history of ischemic cerebral infarction
- history of peripheral vascular disease
- coronary Agatston score of >400
- plaque on vascular measurement

Extensive statistical description

Fisher's exact was used for binary variables and the T-test and Wilcoxon test were used depending on normality for continuous variables. For non-binary categorical variables, the Chi-squared test with simulation

Multivariable logistic regression model

Collected baseline variables including core lab data, with $p < 0.10$ in the age and sex adjusted logistic regression, with exception of EHRA class, number of comorbidities, CHA₂DS₂-VASc score and medications, were included in a bidirectional step-wise variable selection, starting with age and sex in the model. Variables were then added to the model in order of (increasing) p-value of age and sex-adjusted analyses, starting with the variable that has the lowest p-value in the age and sex-adjusted logistic regressions.

Before a new variable is added, variables in the model with $p \geq 0.05$ were identified and removed, starting with the one with the biggest p-value. Before each potential next removal, the model was refit and thus the recalculated p-value was used to determine if there was a next variable with $p \geq 0.05$. If no variables were to be removed the next variable was added.

The bi-directional stepping consists of a single forward stepping of all the variables, interspersed with backwards stepping of the variables in the model before each next step in the forward-stepping.

The statistical criterion for removing a variable from the model (during each backwards stepping) was $p \geq 0.05$.

The step-wise variable selection will ensure that no variables with $p \geq 0.05$ will end up in the final multivariable model. However, due to possible negative confounding even a variable with $p \geq 0.05$ in an age and sex-adjusted model, may have $p < 0.05$ in a model with additional covariates. The aim was also not to keep a too big set of variables to be included in the step-wise process. Therefore, a $p < 0.10$ was selected as a trade-off between the most stringent selection (based on $p < 0.05$) and the least-stringent selection (that is without taking into account the p-value of age and sex-adjusted regressions).

In the final multivariate model (obtained at the end of the bidirectional stepping), testing for each possible second-order interaction was done (i.e. an interaction between two variables or an interaction with itself (quadratic term), what happened if this interaction is added to the model). Specifically, the p-value of the interaction was checked. Of all possible interactions, none reached Bonferroni significance (taking into account multiple testing).

Hosmer and Lemeshow test was used for goodness of fit test with 8 degrees of freedom, Chi-squared = 6.68, p-value = 0.572

Discrimination slope of the main model = 0.082

Imputation

Imputation was implemented for missing values using the R package mice. Mice creates multiple imputations for multivariate missing data. For this article 4000 imputations for each model fit that required imputation were performed. Each incomplete variable was imputed by a separate model. The default methods of predictive mean matching for numeric data and logistic regression for binary data.

For each logistic regression "massive imputation" was performed, which means that all variables in a model were at the same time also used for the imputation needed for the fit of that model. Internally, mice performs the logistic regression fit on all 4000 imputations. It pools the results according to Rubin's rules for imputation, with a small sample refinement of the method to compute degrees of freedom according to Barnard and Rubin.

1. van Buuren, Stef, Groothuis-Oudshoorn, Karin. Mice: Multivariate imputation by chained equations in R. *Journal of Statistical Software*. 2011;45(3):1-67.

Internal validation

Internal validation was accessed using bootstrapping.(1) Fifty bootstrap samples were used.(2). The optimism caused by overfitting in the C-statistic of our model without biomarkers to be 3.03%.”

1. Moons KGM, Kengne AP, Woodward M, *et al*

Risk prediction models: I. Development, internal validation, and assessing the incremental value of a new (bio)marker *Heart* 2012;**98**:683-690.

2. Fernandez-Felix BM, García-Esquinas E, Muriel A, Royuela A, Zamora J. Bootstrap internal validation command for predictive logistic regression models. *The Stata Journal*. 2021;**21**(2):498-509.

Risk score

The multivariable model was used to calculate the linear predictor for all patients with complete data. This was done in the standard way, namely linear combinations of the variables with the beta coefficients as weights, specifically we obtained the following expression for the linear predictor (centering continuous variables on their respective means):

Linear prediction.= Female sex * (-0.5586208) + (PR-interval - 168.8658) * 0.01309933 + (LA contraction function - 17.30064) * (-0.08169752) + (waist circumference - 101.1294) * 0.02787357 + mitral valve regurgitation * 1.771005

Next, these linear predictors was used to calculate a factor (called F here for simplicity), such that the 95% interval of the linear predictors (from the 2.5% quantile to the 97.5% quantile), when multiplied with this factor, is of length 10. This was done, because the aim was a point-based risk score that can vary from 0 up to and including 10 with only very few occurrences outside of this interval.

F can be interpreted as a conversion factor such that the product of a variable with its beta coefficient and F represents a certain number of points. This factor F was then used to obtain a preliminary scoring scheme in the following way:

For binary variables, the no-level gets zero points and the yes-level gets beta * F points. The number of points was rounded to the nearest integer value. For continuous variables, the step size is defined as the inverse of the absolute value of its beta coefficient and F rounded up to the nearest integer value. Step size can be interpreted as the number of units of the variable per point. The number of levels the variable has in the point-based risk score is then set to the range of the variable (maximum minus minimum) divided by the step size rounded down to the nearest integer value.

The range of the variable is then divided using intervals of length equal to the step size and centered in the range of the variable. The number of points assigned to the interval is the value of the variable in its midpoint times its beta coefficient times F, rounded to the nearest integer value. Finally, the first interval is the extended to include also the smallest values of the variable and the last one to include also the largest values, so that the entire range of the variable is covered. Age was removed from the point based risk score.

Mathematical formula of AF progression

$$B = \sum_i T_i$$

B = AF burden

\sum_i = sum of all AF episodes in the time period of which the AF burden or weighted AF burden is calculated.

T_i = time of AF episode i .

$$B_{weighted} = \sum_i w_i T_i$$

$B_{weighted}$ = weighted AF burden,

\sum_i = sum of all AF episodes in the time period of which the AF burden or weighted AF burden is calculated.

w_i = weight factor for AF episode i .

$$w_i = 2 \times (t_i - t_{start}) / (t_{stop} - t_{start})$$

t_i = time when AF episode took place (time chosen is in the middle of the episode)

t_{start} = start time of the period of which the weighted AF burden is calculated,

t_{stop} = end time of the period of which the weighted AF burden is calculated.

T_i = length of time of AF episode i .

AF progression is calculated with the formulas of B and $B_{weighted}$.

$$P = B_{weighted} - B$$

P = AF progression.

Progression is presented as percentage

$$100 \times (P / (t_{stop} - t_{start})) \%$$

t_{start} = start time of the period of which the weighted AF burden is calculated,

t_{stop} = end time of the period of which the weighted AF burden is calculated.

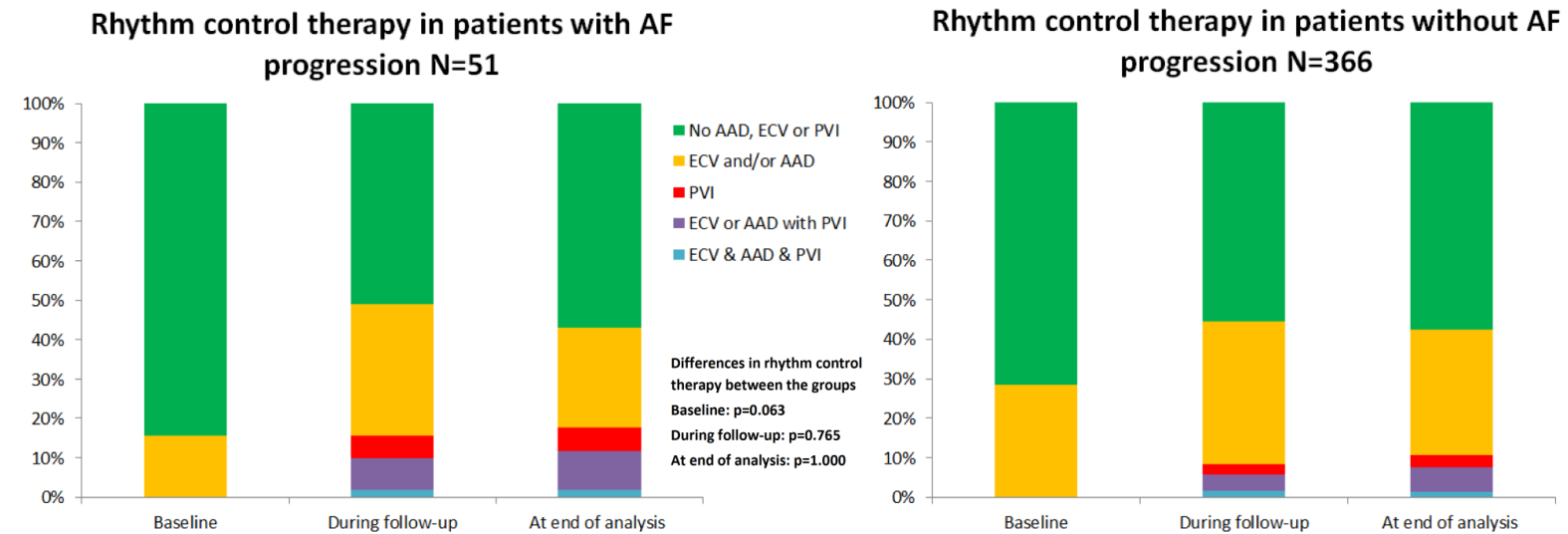
An increase >3% AF burden over the first six months or total follow-up was chosen as definition for atrial fibrillation progression. This cut-off point was chosen because the results were most consistent with the assessment of the physicians.

Supplementary Figure S1. RACE V study design overview

Investigations at baseline and during follow-up	T = 0	T = 1 year	T = 2.5 years	End of device/ End of study*
Clinical history	●	●	●	
Examination	●	●	●	
Blood	●		●	
Electrocardiogram	●	●	●	
Echocardiography	●		●	
Blood biomarkers	●		●	
Questionnaires	●	●	●	
Genotyping	●			
Vascular assessment	●			
CT heart	●			
Adverse events				
Rhythm monitoring				

* End of study was variable and dependent on form of consent for study follow-up with continuous rhythm monitoring until 2,5 years, at end of battery of Reveal LINQ or at 4 years for patients with a pacemaker.

Supplementary Figure S2. RACE V rhythm control therapy overview



Use of rhythm control therapy.

AAD= antiarrhythmic drug; AF= atrial fibrillation; ECV = electro cardioversion; PVI = pulmonary vein isolation

2 patients with AF progression used amiodarone, both started during follow-up, 1 stopped during follow-up, 1 continued until end of analysis.

4 patients without AF progression used amiodarone, all started during follow-up, 1 stopped during follow-up, 3 continued until end of analysis.

5 of 26 patients (19%) undergoing PVI showed AF progression. 9 of 30 patients (30%) undergoing ECV showed AF progression. 3 of 14 (21%) patients undergoing both ECV and PVI showed AF progression.

Supplementary Table S1. RACE V inclusion and exclusion criteria

Inclusion criteria	Age > 18 years
	Total history <10 years of paroxysmal, self-terminating AF
	At least one documented episode of AF and 2 symptomatic episodes or two documented episodes, documented as: <ul style="list-style-type: none"> - AF on ECG, Holter-recording, loop recorder, event recorder or MyDiagnostick; or - Subclinical AF (SCAF) detected in a Medtronic pacemaker (atrial rate > 190 bpm lasting > 6 minutes)
	Able and willing to sign informed consent for the registry
	Able and willing to undergo implantation of ILR (in patients without a CIED)
	CHA2DS2-VASc score ≤5
	No other indication for oral anticoagulation (e.g. mechanical valve prosthesis)
Exclusion criteria	Non-self-terminating, persistent AF;
	Only AF due to a trigger (i.e. postoperative, due to infection)
	Congenital heart disease
	Refusing to temporarily stop (N)OAC for coagulation phenotyping (in patients already on (N)OAC before inclusion in this study), with the exception for patients with a history of ischemic stroke/ transient ischemic attack;
	Prior pulmonary vein isolation (PVI) or on waiting list for PVI or expected to be placed on waiting list within one year, since it is expected that those patients will not show much AF recurrences.
	Expected to start with, or currently using amiodarone, since it is expected that those patients will not show AF recurrences.
	Pregnancy
	ICD, CRT or pacemaker that is not a Medtronic pacemaker due to differences in AHRE algorithm or incompatibility with the type of home-monitoring
	Life expectancy of less than 2.5 years
	Ventricular pacing >50% in patients with a Medtronic pacemaker

Supplementary Table S2. List of 92 biomarkers, Olink Cardiovascular III panel (v.6113)

Abbreviations	Biomarkers	Uniprot ID	OlinkID
ALCAM	CD166 antigen	Q13740	OID00572
AP-N	Aminopeptidase N	P15144	OID00611
AXL	Tyrosine-protein kinase receptor UFO	P30530	OID00612
AZU1	Azurocidin	P20160	OID00597
BLM hydrolase	Bleomycin hydrolase	Q13867	OID00581
CASP-3	Caspase-3	P42574	OID00630
CCL15	C-C motif chemokine 15	Q16663	OID00629
CCL16	C-C motif chemokine 16	O15467	OID00654
CCL24	C-C motif chemokine 24	O00175	OID00592

CD163	Scavenger receptor cysteine-rich type 1 protein M130	Q86VB7	OID00577
CD93	Complement component C1q receptor	Q9NPY3	OID00639
CDH5	Cadherin-5	P33151	OID00587
CHI3L1	Chitinase-3-like protein 1	P36222	OID00633
CHIT1	Chitotriosidase-1	Q13231	OID00605
CNTN1	Contactin-1	Q12860	OID00586
COL1A1	Collagen alpha-1(I) chain	P02452	OID00641
CPA1	Carboxypeptidase A1	P15085	OID00624
CPB1	Carboxypeptidase B	P15086	OID00632
CSTB	Cystatin-B	P04080	OID00575
CTSD	Cathepsin D	P07339	OID00622
CTSZ	Cathepsin Z	Q9UBR2	OID00643
CXCL16	C-X-C motif chemokine 16	Q9H2A7	OID00601
DLK-1	Protein delta homolog 1	P80370	OID00598
EGFR	Epidermal growth factor receptor	P00533	OID00637
Ep-CAM	Epithelial cell adhesion molecule	P16422	OID00610
EPHB4	Ephrin type-B receptor 4	P54760	OID00569
FABP4	Fatty acid-binding protein, adipocyte	P15090	OID00589
FAS	Tumor necrosis factor receptor superfamily member 6	P25445	OID00615
Gal-3	Galectin-3	P17931	OID00578
Gal-4	Galectin-4	P56470	OID00626
GDF-15	Growth/differentiation factor 15	Q99988	OID00595
GP6	Platelet glycoprotein VI	Q9HCN6	OID05026
GRN	Granulins	P28799	OID00579
ICAM-2	Intercellular adhesion molecule 2	P13598	OID00646
IGFBP-1	Insulin-like growth factor-binding protein 1	P08833	OID00604
IGFBP-2	Insulin-like growth factor-binding protein 2	P18065	OID00650
IGFBP-7	Insulin-like growth factor-binding protein 7	Q16270	OID00638
IL-17RA	Interleukin-17 receptor A	Q96F46	OID00566
IL-18BP	Interleukin-18-binding protein	O95998	OID00640
IL-1RT1	Interleukin-1 receptor type 1	P14778	OID00613
IL-1RT2	Interleukin-1 receptor type 2	P27930	OID00627
IL-2RA	Interleukin-2 receptor subunit alpha	P01589	OID00570
IL-6RA	Interleukin-6 receptor subunit alpha	P08887	OID00602
ITGB2	Integrin beta-2	P05107	OID00565
JAM-A	Junctional adhesion molecule A	Q9Y624	OID00625
KLK6	Kallikrein-6	Q92876	OID00647

LDL receptor	Low-density lipoprotein receptor	P01130	OID00564
LTBR	Lymphotoxin-beta receptor	P36941	OID00583
MB	Myoglobin	P02144	OID00616
MCP-1	Monocyte chemotactic protein 1	P13500	OID00576
MEPE	Matrix extracellular phosphoglycoprotein	Q9NQ76	OID00132
MMP-2	Matrix metalloproteinase-2	P08253	OID00614
MMP-3	Matrix metalloproteinase-3	P08254	OID00644
MMP-9	Matrix metalloproteinase-9	P14780	OID00568
MPO	Myeloperoxidase	P05164	OID00600
Notch 3	Neurogenic locus notch homolog protein 3	Q9UM47	OID00584
NT-proBNP	N-terminal prohormone brain natriuretic peptide	NA	OID00131
OPG	Osteoprotegerin	O00300	OID00571
OPN	Osteopontin	P10451	OID00621
PAI	Plasminogen activator inhibitor 1	P05121	OID00591
PCSK9	Proprotein convertase subtilisin/kexin type 9	Q8NBP7	OID00619
PDGF subunit A	Platelet-derived growth factor subunit A	P04085	OID00648
PECAM-1	Platelet endothelial cell adhesion molecule	P16284	OID00652
PGLYRP1	Peptidoglycan recognition protein 1	O75594	OID00623
PI3	Elafin	P19957	OID00609
PLC	Perlecan	P98160	OID00582
PON3	Paraoxonase	Q15166	OID00642
PRTN3	Myeloblastin	P24158	OID00618
PSP-D	Pulmonary surfactant-associated protein D	P35247	OID00608
RARRES2	Retinoic acid receptor responder protein 2	Q99969	OID00645
RETN	Resistin	Q9HD89	OID00603
SCGB3A2	Secretoglobulin family 3A member 2	Q96PL1	OID00636
SELE	E-selectin	P16581	OID00596
SELP	P-selectin	P16109	OID00574
SHPS-1	Tyrosine-protein phosphatase non-receptor type substrate 1	P78324	OID00628
SPON1	Spondin-1	Q9HCB6	OID00599
ST2	ST2 protein	Q01638	OID00634
TFF3	Trefoil factor 3	Q07654	OID00573
TFPI	Tissue factor pathway inhibitor	P10646	OID00590
TIMP4	Metalloproteinase inhibitor 4	Q99727	OID00585
TLT-2	Trem-like transcript 2 protein	Q5T2D2	OID00588
TNF-R1	Tumor necrosis factor receptor 1	P19438	OID00649

TNF-R2	Tumor necrosis factor receptor 2	P20333	OID00567
TNFRSF10C	Tumor necrosis factor receptor superfamily member 10C	O14798	OID00594
TNFRSF14	Tumor necrosis factor receptor superfamily member 14	Q92956	OID00563
TNFSF13B	Tumor necrosis factor ligand superfamily member 13B	Q9Y275	OID00617
t-PA	Tissue-type plasminogen activator	P00750	OID00635
TR	Transferrin receptor protein 1	P02786	OID00593
TR-AP	Tartrate-resistant acid phosphatase type 5	P13686	OID00606
uPA	Urokinase-type plasminogen activator	P00749	OID00631
U-PAR	Urokinase plasminogen activator surface receptor	Q03405	OID00620
vWF	Von Willebrand factor	P04275	OID00651

Supplementary Table S3. Inclusion distribution per participating centre

Centre	Number of inclusion
University Medical Centre Groningen	100
Maastricht University Medical Centre	106
Ommelander Hospital Groningen	31
Martini Hospital	111
Rijnstate Hospital	40
University of Amsterdam	22
Isala Hospital	4
Laurentius Hospital	2
VU Medical Centre Amsterdam	2

Supplementary Table S4. Baseline Characteristics of AF progression groups

Characteristic	No AF recurrence (group 1) (N=48)	AF recurrence without AF progression (group 2) (N=318)	AF progression without persistent AF (group 3) (N=16)	AF progression with persistent AF (group 4) (N=35)
Age (years)	63 (54-72)	65 (58-71)	63 (58-71)	65 (62-74)
Female sex	19 (40%)	145 (46%)	5 (31%)	10 (29%)
Total history AF (years)	1.6 (0.5-4.7)	2.6 (0.8-5.3)	2.5 (1.0-3.4)	3.6 (0.9-5.7)
Heart failure	11 (39%)	93 (50%)	6 (55%)	15 (63%)
HFrEF	2 (4%)	4 (1%)	1 (1 %)	3 (9%)
HFpEF	9 (19%)	89 (28%)	5 (31%)	11 (31%)
Hypertension	42 (88%)	250 (78%)	16 (100%)	30 (86%)
Diabetes mellitus	3 (6%)	26 (8%)	1 (6%)	4 (11%)
Coronary artery disease	4 (8%)	33 (10%)	2 (13%)	9 (26%)
Atherosclerosis*	25 (52%)	153 (48%)	9 (56%)	17 (49%)
Peripheral artery disease	0 (0%)	1 (0%)	0 (0%)	2 (6%)
Ischemic stroke	3 (6%)	15 (5%)	1 (6%)	0 (0%)

Chronic obstructive pulmonary disease	0 (0%)	19 (6%)	2 (13%)	2 (6%)
Number of Comorbidities**	2 (2-3)	2 (2-3)	3 (2-3)	3 (2-4)
CHA₂DS₂-VASc score***				
≤ 2	35 (73%)	230 (72%)	15 (94%)	30 (86%)
>2	13 (27%)	88 (28%)	1 (6%)	5 (14%)
EHRA class				
I	9 (19%)	23 (7 %)	4 (25%)	7 (20%)
IIa	16 (33%)	98 (31%)	4 (25%)	17 (49%)
IIb	16 (33%)	139 (44%)	6 (38%)	6 (17%)
III	7 (15%)	56 (18%)	2 (13%)	5 (14%)
IV	0 (0%)	2 (1%)	0 (0%)	0 (0%)
Physical examination				
Height (cm)	178 (172-183)	176 (168-184)	177 (172-187)	179 (170-183)
Weight (kg)	88 (77-98)	84 (74-96)	91 (72-104)	88 (75-100)
BMI (kg/m ²)	27(25- 30)	27 (24-30)	26 (25-30)	27 (24-32)
Obesity (BMI>30)	13 (27%)	79 (25%)	4 (25%)	11 (32%)
Waist circumference (cm)	99 (93-109)	100 (92-108)	103 (95-111)	106 (101-114)
Systolic blood pressure (mmHg)	133 (124-140)	134 (125-145)	130 (122-136)	130 (124-144)
Diastolic blood pressure (mmHg)	80 (75-85)	80 (74-85)	75 (72-83)	80 (72-85)
Laboratory results				
Creatinine (μmol/L)	77 (67-85)	80 (69-91)	89 (80-99)	87 (80-100)
eGFR (mL/min*1.73m ²)	85 (76-93)	81 (69-90)	71 (63-83)	76 (68-86)
Electrocardiogram				
PR-interval	166 (148-173)	164 (149-186)	170 (160-200)	180 (168-199)
QRS-interval	94 (88-101)	94 (86-104)	94 (90-103)	96 (90-110)
Medications				
β-blocker	25 (52%)	156 (49%)	11 (69%)	21 (60%)
Verapamil/Diltiazem	8 (17%)	58 (18%)	3 (19%)	4 (11%)
Digoxin	0 (0%)	4 (1 %)	0 (0 %)	2 (6%)
Class I antiarrhythmic drugs	13 (27 %)	76 (24%)	3 (19%)	2 (6%)
Class III antiarrhythmic drugs	1 (2%)	14 (4%)	1 (6%)	3 (9%)
ACE-inhibitor	12 (25%)	59 (19%)	3 (19%)	8 (23%)
Angiotensin Receptor Blocker	8 (17%)	58 (18%)	5 (31 %)	9 (26%)
Statin	10 (21%)	109 (34%)	10 (63%)	16 (46%)
Diuretic	7 (15%)	47 (15%)	3 (19%)	7 (20%)
Anticoagulant	29 (60%)	215 (68%)	13 (81%)	32 (91%)
Vitamin K antagonist	4 (8%)	41 (13%)	3 (19%)	7 (20%)
NOAC	25 (52%)	174 (55%)	10 (63%)	25 (71%)
Echocardiographic variables ^a				
Left atrial volume (mL)	58 (46-67)	58 (47-74)	60 (55-76)	68 (55-85)
Left atrial volume index (mL/m ²)	28 (22-34)	29 (24-36)	32 (25-35)	35 (26-39)
Left atrial reservoir function (%)	38 (31-47)	37 (29-43)	35 (27- 53)	31 (25-35)

Left atrial contractile function (%)	18 (13-23)	17 (13-21)	15 (10-23)	13 (11-15)
Left atrial conduction function (%)	19 (15-24)	19 (14-24)	23 (15-34)	18 (13-23)
Right atrial volume (mL)	39 (34-56)	48 (38-65)	55 (47-79)	55 (46-63)
Right atrial volume indexed (mL/m²)	21 (16- 29)	24 (20 - 31)	32 (23- 33)	28 (24 - 33)
Left ventricular ejection fraction (%)	51 ± 10	51 ± 8	52 ± 9	50 ± 8
Left ventricular mass (g)	150 (140-165)	148 (126-178)	152 (139-182)	161 (134-188)
Left ventricular mass index (g/m²)	75 (68-83)	76 (64-88)	86 (69-88)	78 (67-96)
Left ventricle strain	-14.5 ± 2.5	-14.0 ± 2.3	-14.2 ± 2.6	-14.3 ± 2.6
Computed Tomography ^b				
Calcium score (Agatston)	15 (0-75)	22 (0-227)	94 (15-3270)	152 (4-917)
Agatston >400				
Pericardial fat	171 (134-223)	166 (118- 232)	205 (160-224)	167 (137-235)
Epicardial fat	102 (74-132)	97 (70-128)	104 (90-126)	105 (72-137)
Vascular assessment ^c				
IMT max-CCA (mm)	0.90 (0.82-1.04)	0.92 (0.81-1.07)	1.03 (0.85-1.19)	0.97 (0.83-1.13)
IMT max-CCA >1mm	14 (33%)	95 (34%)	8 (53%)	11 (42%)
IMT max-all segments (mm)	1.04 (0.93-1.20)	0.98 (0.84-1.16)	1.00 (0.88-1.20)	1.02 (0.88-1.14)
IMT max-all segments >1mm	24 (57%)	130 (47%)	7 (47%)	13 (50%)
Pulse wave velocity (m/s)	8.60 (6.98-10.00)	8.49 (7.45-10.20)	8.46 (7.66-10.25)	9.20 (8.14-10.26)
Plaques	23 (64%)	102 (48%)	6 (60%)	9 (60%)
Plaques >3	2 (4%)	10 (3%)	2 (12%)	3 (9%)

Data are presented as mean±standard deviation, number of patients (%), or median (interquartile range). Abbreviations: ACE=angiotensin-converting enzyme; AF=atrial fibrillation; BMI=body mass index; CCA= common carotid artery; eGFR=estimated glomerular filtration rate; EHRA= European Heart Rhythm Association class for symptoms; HFpEF= heart failure with preserved ejection fraction; HFrEF= heart failure with reduced ejection fraction; IMT=intima media thickness; NOAC= novel oral anticoagulation; NT-proBNP=N-terminal pro-brain natriuretic peptide; *Atherosclerosis is presence of history of myocardial infarction, percutaneous coronary intervention, coronary artery bypass graft, ischemic cerebral infarction, peripheral vascular disease, Agatston score >400 or plaque; **The number of comorbidities was calculated by awarding points for hypertension, heart failure, age >65 years, diabetes mellitus; coronary artery disease, BMI>25kg/m², moderate or severe mitral valve regurgitation and kidney dysfunction (eGFR<60); ***The CHA₂DS₂-VASc score assesses thromboembolic risk. C=congestive heart failure/LV dysfunction, H=hypertension; A2=age ≥75 years; D=diabetes mellitus; S2=stroke/transient ischemic attack/systemic embolism; V=vascular disease; A=age 65-74 years; Sc=sex category (female sex).). ^aLeft atrial and ventricle strain measurements could not be performed in 75 patients. Measurements of right atrial strain could not be done in 123 patients. ^bAgatston score was not available for 10 patients, epicardial and pericardial fat could not be analysed for 21 patients. ^cIMT CCA was not available for 55 patients, IMT all segments for 56 patients and pulse wave velocity could not be measured in 78 patients and amount of plaques could not be measured in 145 patients.

Supplementary Table S5. Olink biomarkers at baseline

Characteristic	AF progression (N=51)	No AF progression (N=366)	Total (N=417)	P-value
ALCAM	6.06 (5.82 - 6.20)	6.07 (5.84 - 6.22)	6.07 (5.84 - 6.22)	0.772
AP-N	5.46 (5.26 - 5.69)	5.53 (5.33 - 5.76)	5.53 (5.33 - 5.76)	0.107
AXL	9.44 (9.24 - 9.63)	9.39 (9.16 - 9.60)	9.39 (9.16 - 9.60)	0.168
AZU1	4.59 (4.33 - 5.16)	4.61 (4.28 - 5.03)	4.61 (4.28 - 5.03)	0.298
BLM hydrolase	6.26 (6.02 - 6.56)	6.21 (5.91 - 6.48)	6.21 (5.91 - 6.48)	0.181
CASP-3	8.32 (7.56 - 9.40)	8.52 (7.31 - 9.49)	8.45 (7.36 - 9.49)	0.883
CCL15	8.08 (7.90 - 8.38)	8.04 (7.78 - 8.36)	8.04 (7.78 - 8.36)	0.259
CCL16	7.45 (7.09 - 7.71)	7.44 (7.07 - 7.71)	7.44 (7.07 - 7.71)	0.929
CCL24	6.37 (5.46 - 6.88)	6.24 (5.64 - 6.87)	6.24 (5.63 - 6.87)	0.926
CD163	8.02 (7.65 - 8.39)	7.96 (7.60 - 8.25)	7.96 (7.61 - 8.27)	0.323
CD93	11.82 (11.57 - 12.00)	11.79 (11.56 - 12.00)	11.79 (11.56 - 12.00)	0.690
CDH5	4.34 (4.06 - 4.59)	4.38 (4.08 - 4.59)	4.38 (4.08 - 4.59)	0.849
CHI3L1	7.46 (6.87 - 7.95)	7.22 (6.68 - 7.89)	7.22 (6.68 - 7.89)	0.128
CHIT1	7.29 (6.69 - 8.12)	7.16 (6.46 - 7.75)	7.16 (6.46 - 7.75)	0.281
CNTN1	4.75 (4.40 - 4.89)	4.78 (4.47 - 5.04)	4.78 (4.45 - 5.03)	0.161
COL1A1	3.45 (3.12 - 3.71)	3.47 (3.20 - 3.71)	3.46 (3.20 - 3.71)	0.446
CPA1	6.67 (6.34 - 7.20)	6.62 (6.21 - 7.02)	6.62 (6.21 - 7.02)	0.379
CPB1	6.61 (6.26 - 7.07)	6.53 (6.08 - 6.88)	6.53 (6.08 - 6.88)	0.130
CSTB	5.54 (5.19 - 5.80)	5.38 (5.05 - 5.76)	5.38 (5.05 - 5.76)	0.176
CTSD	5.22 (5.06 - 5.67)	5.11 (5.06 - 5.40)	5.11(5.06- 5.40)	0.021
CTSZ	5.97 (5.68 - 6.20)	5.91 (5.68 - 6.10)	5.92 (5.68 - 6.11)	0.259
CXCL16	5.85 (5.71 - 6.04)	5.90 (5.66 - 6.12)	5.90 (5.67 - 6.12)	0.489
DLK-1	7.01 (6.71 - 7.34)	6.95 (6.58 - 7.35)	6.95 (6.59 - 7.35)	0.435
EGFR	3.52 (3.38 - 3.75)	3.62 (3.45 - 3.82)	3.62 (3.45 - 3.82)	0.060
Ep-CAM	6.40 (5.91 - 7.39)	6.66 (5.91 - 7.50)	6.66 (5.91 - 7.50)	0.382
EPHB4	5.18 (4.89 - 5.31)	5.12 (4.89 - 5.32)	5.12 (4.89 - 5.32)	0.501
FABP4	6.64 (6.07 - 7.23)	6.40 (5.94 - 6.96)	6.40 (5.94 - 6.96)	0.107
FAS	6.14 (5.95 - 6.45)	6.14 (5.89 - 6.35)	6.14 (5.89 - 6.35)	0.188
Gal-3	6.61 (6.20 - 6.82)	6.51 (6.27 - 6.75)	6.51 (6.27 - 6.75)	0.281
Gal-4	4.17 (3.80 - 4.44)	4.14 (3.79 - 4.42)	4.14 (3.79 - 4.42)	0.475
GDF-15	6.60 (6.21 - 6.99)	6.38 (6.00 - 6.75)	6.38 (6.00 - 6.75)	0.010
GP6	3.10 (2.60 - 3.62)	3.12 (2.55 - 3.71)	3.13 (2.55 - 3.71)	0.904
GRN	6.92 (6.76 - 7.16)	6.93 (6.69 - 7.14)	6.93 (6.70 - 7.14)	0.331
ICAM-2	5.75 (5.42 - 5.96)	5.70 (5.38 - 5.97)	5.70 (5.39 - 5.97)	0.532
IGFBP-1	4.80 (3.99 - 5.56)	4.71 (3.82 - 5.66)	4.71 (3.82 - 5.66)	0.610
IGFBP-2	8.45 (7.82 - 8.90)	8.32 (7.65 - 8.83)	8.32 (7.65 - 8.83)	0.182
IGFBP-7	8.30 (8.07 - 8.66)	8.25 (7.99 - 8.50)	8.25 (7.99 - 8.50)	0.093
IL-17RA	4.36 (4.09 - 4.63)	4.41 (4.03 - 4.70)	4.40 (4.05 - 4.69)	0.879
IL-18BP	6.73 (6.53 - 6.99)	6.68 (6.44 - 6.95)	6.69 (6.45 - 6.96)	0.197

IL-1RT1	6.90 (6.74 - 7.12)	6.92 (6.68 - 7.11)	6.92 (6.68 - 7.11)	0.697
IL-1RT2	5.60 (5.44 - 5.80)	5.63 (5.41 - 5.85)	5.63 (5.41 - 5.84)	0.448
IL2-RA	4.75 (4.51 - 5.14)	4.66 (4.332 - 4.93)	4.66 (4.33 - 4.92)	0.054
IL-6RA	12.82 (12.50 - 13.05)	12.85 (12.50 - 13.13)	12.84 (12.49 - 13.12)	0.634
ITGB2	6.42 (6.12 - 6.72)	6.35 (6.047 - 6.66)	6.35 (6.05 - 6.66)	0.250
JAM-A	6.82 (6.15 - 7.77)	6.95 (6.11 - 7.92)	6.95 (6.11 - 7.92)	0.738
KLK6	6.16 (5.89 - 6.40)	6.11 (5.84 - 6.33)	6.11 (5.86 - 6.34)	0.250
LDL receptor	5.42 (5.01 - 5.84)	5.52 (5.10 - 5.90)	5.52 (5.10 - 5.90)	0.385
LTBR	4.685 (4.38 - 4.98)	4.68 (4.40 - 4.91)	4.68 (4.40 - 4.92)	0.683
MB	7.36 (7.04 - 7.68)	7.23 (6.82 - 7.60)	7.23 (6.82 - 7.60)	0.060
MCP-1	4.42 (4.15 - 4.63)	4.44 (4.20 - 4.65)	4.44 (4.20 - 4.65)	0.455
MEPE	6.24 (5.86 - 6.55)	6.15 (5.88 - 6.45)	6.16 (5.87 - 6.46)	0.429
MMP-2	4.58 (4.34 - 4.80)	4.54 (4.27 - 4.79)	4.56 (4.29 - 4.79)	0.232
MMP-3	7.35 (6.82 - 7.66)	7.05 (6.58 - 7.50)	7.08 (6.59 - 7.54)	0.042
MMP-9	4.91 (4.45 - 5.30)	4.86 (4.27 - 5.48)	4.86 (4.27 - 5.48)	0.668
MPO	4.15 (3.93 - 4.56)	4.078 (3.83 - 4.37)	4.078 (3.83 - 4.37)	0.075
Notch 3	5.32 (4.88 - 5.65)	5.30 (4.95 - 5.62)	5.30 (4.95 - 5.62)	0.926
NT-proBNP	5.65 (4.75 - 6.41)	4.94 (4.02 - 5.69)	4.99 (4.09 - 5.78)	<0.001
OPG	4.570 (4.21 - 4.77)	4.50 (4.20 - 4.74)	4.50 (4.20 - 4.74)	0.488
OPN	7.00 (6.49 - 7.33)	6.82 (6.50 - 7.17)	6.82 (6.50 - 7.17)	0.211
PAI	5.70 (5.19 - 6.29)	5.78 (5.21 - 6.43)	5.77 (5.20 - 6.41)	0.714
PCSK9	3.11 (2.96 - 3.36)	3.07 (2.81 - 3.38)	3.07 (2.81 - 3.38)	0.214
PDGF subunit A	4.48 (4.00 - 4.89)	4.46 (3.87 - 5.17)	4.46 (3.87 - 5.13)	0.718
PECAM-1	5.76 (5.37 - 6.44)	5.86 (5.33 - 6.46)	5.86 (5.33 - 6.46)	0.978
PGLYRP1	8.43 (8.12 - 8.72)	8.20 (7.90 - 8.55)	8.20 (7.90 - 8.55)	<0.001
PI3	4.59(4.29 - 4.89)	4.39 (4.00 - 4.80)	4.39 (4.00 - 4.80)	0.011
PLC	7.35 (7.19 - 7.59)	7.30 (7.11 - 7.53)	7.30 (7.11 - 7.53)	0.080
PON3	5.69 (5.28 - 6.27)	6.01 (5.53 - 6.47)	6.01 (5.53 - 6.47)	0.004
PRTN3	5.62 (5.41 - 6.15)	5.50 (5.20 - 5.87)	5.50 (5.20 - 5.87)	0.019
PSP-D	3.48 (2.99 - 3.85)	3.29 (2.81 - 3.80)	3.29 (2.82 - 3.80)	0.142
RARRES2	12.19 (11.97 - 12.44)	12.26 (12.01 - 12.44)	12.26 (12.01 - 12.44)	0.477
RETN	7.03 (6.75 - 7.42)	6.876 (6.60 - 7.22)	6.88 (6.60 - 7.22)	0.013
SCGB3A2	3.19 (2.80 - 3.72)	3.15 (2.67 - 3.63)	3.15 (2.67 - 3.63)	0.756
SELE	13.30 (12.95 - 13.78)	13.23 (12.77 - 13.59)	13.24 (12.78 - 13.59)	0.324
SELP	11.11 (10.61 - 11.76)	11.10 (10.56 - 11.76)	11.10 (10.56 - 11.76)	0.799
SHPS-1	3.84 (3.63 - 4.06)	3.87 (3.66 - 4.12)	3.87 (3.65 - 4.12)	0.541
SPON1	1.21 (0.99 - 1.39)	1.15 (0.89 - 1.38)	1.146 (0.89 - 1.38)	0.152
ST2	4.97 (4.58 - 5.26)	4.94 (4.58 - 5.25)	4.94 (4.58 - 5.25)	0.888
TFF3	6.00 (5.76 - 6.25)	5.85 (5.589 - 6.08)	5.85 (5.58 - 6.08)	0.002
TFPI	10.16 (9.96 - 10.44)	10.33 (10.09 - 10.55)	10.29 (10.08 - 10.54)	0.012
TIMP4	4.25 (4.03 - 4.55)	4.17 (3.90 - 4.54)	4.20 (3.91 - 4.54)	0.248
TLT-2	5.40 (5.14 - 5.70)	5.44 (5.12 - 5.74)	5.44 (5.13 - 5.74)	0.921
TNF-R1	7.08 (6.78 - 7.32)	6.91 (6.67 - 7.16)	6.91 (6.67 - 7.16)	0.021

TNF-R2	5.87 (5.56 - 6.11)	5.68 (5.41 - 5.96)	5.68 (5.41 - 5.96)	0.005
TNFRSF10C	7.01 (6.65 - 7.38)	6.88 (6.50 - 7.24)	6.88 (6.50 - 7.24)	0.036
TNFRSF14	5.41 (5.07 - 5.60)	5.27 (5.012 - 5.52)	5.27 (5.01 - 5.52)	0.204
TNFSF13B	7.595 (7.35 - 7.88)	7.58 (7.34 - 7.83)	7.58 (7.34 - 7.83)	0.820
t-PA	7.77 (7.40 - 8.36)	7.84 (7.26 - 8.74)	7.84 (7.26 - 8.74)	0.790
TR	5.54 (5.02 - 5.80)	5.29 (4.88 - 5.74)	5.29 (4.88 - 5.74)	0.115
TR-AP	4.73 (4.46 - 4.97)	4.81 (4.55 - 5.03)	4.79 (4.54 - 5.02)	0.136
uPA	6.33 (6.07 - 6.59)	6.30 (6.09 - 6.57)	6.31 (6.08 - 6.57)	0.873
U-PAR	5.91 (5.66 - 6.16)	5.81 (5.55 - 6.05)	5.81 (5.55 - 6.05)	0.061
vWF	7.85 (7.05 - 9.10)	7.91 (7.05 - 9.09)	7.91 (7.05 - 9.09)	0.965

Data is presented in a log2 scale in median (interquartile range).

Supplementary Table S6. Coagulation markers at baseline

Coagulation markers	Total (N=417)
Factor XIIa:C1inh (pM)	862.64 (747.28- 989.81)
Factor XIIa:antithrombin (pM)	11.28 (11.28-36.06)
Plasma Kallikrein:C1inh (nM)	0.30 (0.3-1.45)
Factor XIa:C1inh (pM)	72.04 (72.04-197.22)
Factor XIa:AT (pM)	7.90 (7.90-7.90)
Factor XIa:a1AT (pM)	56.17 (56.17-92.04)
Factor Xa:AT (pM)	421.62 (348.16- 497.15)
Factor IXa:AT (pM)	170.20 (170.20- 170.20)
Thrombin:AT (ug/L)	2.04 (2.04-3.54)

Data is presented in median (interquartile range). AT= antithrombin; a1AT= alpha-1-antitrypsin; C1inh=C1-Esterase inhibitor; nM=nanomolar; pM=picomolar

Supplementary Table S7. Age and sex adjusted of clinical factors related to AF progression

	OR	95% CI	P-value
Female sex	0.48	0.25-0.91	0.024
CHA₂DS₂-VASc score >1	3.73	1.40-9.95	0.009
Heart failure with reduced ejection fraction	4.31	1.12-16.58	0.034
Plaques >3	5.54	1.59-19.36	0.008
Peripheral artery disease	11.98	1.05-136.78	0.046
Waist circumference (per SD)	1.46	1.06-2.02	0.022
PR interval (per SD)	1.45	1.10-1.92	0.009
Left atrial contractile function (per SD)	0.60	0.39-0.92	0.019
Left atrium end diastolic volume (per SD)	1.46	1.08-1.97	0.014
Left atrium end diastolic volume indexed for BSA (per SD)	1.44	1.07-1.95	0.017
Right atrium end systolic volume (per SD)	1.53	1.05-2.23	0.026
Right atrium end systolic volume indexed for BSA (per SD)	1.44	1.01-2.04	0.044

Logistic regression adjusted for age and sex. BSA= body surface area; CI=confidence interval; OR=odds ratio. CHA₂DS₂-VASc score assesses thromboembolic risk. C=congestive heart failure/LV dysfunction, H=hypertension; A₂=age ≥75 years; D=diabetes mellitus; S₂=stroke/transient ischemic attack/systemic embolism; V=vascular disease; A=age 65-74 years; Sc=sex category (female sex).

Supplementary Table S8. Age and sex adjusted analysis including biomarkers

	OR	95% CI	P-value
Female sex	0.48	0.25-0.91	0.024
CHA₂DS₂-VASc score >1	3.73	1.40-9.95	0.009
Heart failure with reduced ejection fraction	4.31	1.12-16.58	0.034
Plaques >3	5.54	1.59-19.36	0.008
Peripheral artery disease	11.98	1.05-136.78	0.046
Waist circumference (per SD)	1.46	1.06-2.02	0.022
PR interval (per SD)	1.45	1.10-1.92	0.009
Left atrial contractile function (per SD)	0.60	0.39-0.92	0.019
Left atrial end diastolic volume (per SD)	1.46	1.08-1.97	0.014
Left atrium end diastolic volume indexed for BSA (per SD)	1.44	1.07-1.95	0.017
Right atrium end systolic volume (per SD)	1.53	1.05-2.23	0.026
Right atrium end systolic volume indexed for BSA(per SD)	1.44	1.01-2.04	0.044
CTSD (per SD)	1.32	1.01-1.73	0.043
FABP4 (per SD)	1.47	1.06-2.05	0.021
NTproBNP (per SD)	2.05	1.43-2.94	<0.001
PCSK9 (per SD)	1.37	1.03-1.81	0.030
PGLYRP1(per SD)	1.44	1.09-1.91	0.011
PON3 (per SD)	0.72	0.54-0.96	0.027
PRTN3 (per SD)	1.29	1.00-1.66	0.046
RETN (per SD)	1.34	1.01-1.76	0.041
TFPI (per SD)	0.72	0.53-0.97	0.030
TNF-R2 (per SD)	1.49	1.11-2.01	0.009
TNFRSF10C (per SD)	1.43	1.03-1.98	0.033
Factor XIIa:antithrombin (> median)	0.39	0.17-0.91	0.030

Factor XIIa:C1-esterase inhibitor (> median)	0.38	0.20-0.75	0.005
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Logistic regression adjusted for age and sex, with imputation. BSA= body surface area; CI=confidence interval; CTSD=capthesin D; FABP4=fatty acid binding protein 4; NTproBNP=N-terminal pro-brain natriuretic peptide; OR=odds ratio; PCSK9= Proprotein convertase subtilisin/kexin type 9; PGLYRP1= peptidoglycan recognition protein 1; PON3= paraoxonase 3; PRTN3=myeloblastin; RETN=resistin; TFPI= tissue factor pathway inhibitor; TNF-R2=tumor necrosis factor receptor 2; TNFRSF10C=tumor necrosis factor receptor superfamily member 10C. CHA₂DS₂-VASc score assesses thromboembolic risk. C=congestive heart failure/LV dysfunction, H=hypertension; A₂=age ≥75 years; D=diabetes mellitus; S₂=stroke/transient ischemic attack/systemic embolism; V=vascular disease; A=age 65-74 years; Sc=sex category (female sex).