Intracellular electrolytes in cardiac and skeletal muscle in fatal digitalis intoxication

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Samples from cardiac and skeletal muscle were obtained immediately after death in a case of fatal digitalis intoxication. Intra- and extracellular electrolytes were determined by atomic absorption spectrophotometry. No changes were found in the extracellular electrolytes, and only modest changes were noted in skeletal muscle. In cardiac muscle, however, profound electrolyte disturbances were recorded with an inward shift of sodium and an outward shift of potassium. This signifies an extensive and early inhibition of the membrane ATPase in cardiac muscle and stresses the importance of immediate and vigorous treatment of the intracellular electrolyte disturbances with a view to suppressing serious and potentially lethal arrhythmias.

Intoxication by digitalis glycosides usually results from therapeutic overdosage (Lown and Levine, 1954; Mason and Braunwald, 1968; Surawicz and Mortelmans, 1969) but an increasing number of cases caused by the ingestion of pills with suicidal intent or by accident have been reported in the past decade (McNamara et al., 1964; Buchanan, 1967; Smith and Willerson, 1971; Bismuth et al., 1973; Steentoft, 1973). These cases present difficult therapeutic problems but they also afford opportunities to record the effects of gross digitalis overdosage. The cause of early malignant arrhythmias in patients with serious digitalis intoxication is generally considered to be disturbance of the electrolyte balance across the cell membrane. There may be very little evidence of these electrolyte disturbances in serum determinations. This is illustrated by a fatal case of digitalis intoxication in which we were able to study the intracellular electrolytes.

Case history

The patient, a woman of 64, was being treated with digoxin 0-25 mg per day because of heart failure. With the intention of committing suicide she ingested 80 to 100 tablets, corresponding to 20 to 25 mg digoxin, and was admitted to the emergency ward a few hours later. She was then fully conscious but complained of breathlessness and had a variable degree of atrioventricular block. Despite the risk of vagal stimulation, she was given a gastric lavage with 4-5 litres of carbon suspension; only one tablet was found in the return fluid. The patient was then transferred to the CCU, where a temporary pacemaker was immediately inserted. During this procedure she developed ventricular fibrillation, which resisted all therapy including external DC shock, artificial respiration, external heart massage, and correction of the acidosis. There were no myocardial contractions despite satisfactory pacing electrode positioning as judged radiologically. The patient died four hours after admission. Blood samples for determination of electrolytes and digoxin had been taken on admission to the hospital.

Methods

Immediately after death, samples were taken from skeletal muscle and the right and left myocardium. A piece of the vastus lateralis muscle was obtained, using the instrument for biopsy described by Bergström (1962). All visible connective tissue and fat were removed and the specimens (40 to 80 mg) were weighed repeatedly on a Cahn 4700 electromagnetic balance. The material was then dried at 100° C to constant weight and the water content was calculated. Fat content was obtained after extraction with petroleum ether. The specimens were analysed for sodium, potassium, and magnesium by atomic absorption spectrophotometry after extraction with

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1 N nitric acid in quartz tubes. Chloride was calculated indirectly after precipitation with silver nitrate.

The intra- and extracellular electrolyte concentrations were determined on the basis of the assumption that chloride is freely diffusible across the cell membrane. The Nernst equation (Conway, 1957) was used to get the actual distribution, assuming a resting membrane potential of 87-2 mV (Bolte et al., 1963). With the aid of the total water and chloride content in the sample and the extracellular concentration of chloride corrected for a Donnan factor and a factor for plasma water, it is possible to estimate the intracellular electrolyte concentrations (Graham et al., 1967; Bergström and Bittar, 1969). Digoxin was determined by the radioimmunoassay method (Smith et al., 1969).

Results

Serum analyses showed sodium 144, potassium 4-9, and chloride 105 mmol/l, urea 3-34 mmol/l, and digoxin 30 ng/ml (therapeutic range 1-5-2-0).

Samples from skeletal muscle showed an ordinary intracellular potassium concentration, 166 mmol/kg intracellular water, well in accord with the values reported by Bergström (1962) in a healthy population (143 to 191 mmol/kg intracellular water). The intracellular sodium concentration was raised, 38-0 mmol/kg intracellular water (normal maximum value 10-9 mmol/kg intracellular water); the chloride concentration, 13-5 mmol/100 g fat-free dry solids (FFDS) was near the upper limit of normal, 14-0 mmol/100 g FFDS. Intracellular water was within normal limits, 256 g/100 g FFDS (normal 231-315 g/100 g FFDS). Magnesium was decreased, 3-70 mmol/100 g FFDS (normal 3-90-4-70 mmol/100 g FFDS).

Analyses of the samples from the right and left ventricular myocardium revealed a probable intracellular potassium deficit with values of 88-7 and 106 mmol/kg intracellular water, respectively. The value for intracellular sodium concentration was about 3 times higher than the corresponding value in the skeletal muscle, 122 and 167 mmol/kg intracellular water, respectively. The chloride concentration in both the left and the right ventricular myocardium was low compared with the values given in Handbook of Biological Data (Spector, 1956). Intracellular water was normal. Low values for magnesium were noted in both the left (3-36 mmol/100 g FFDS) and the right (3-11 mmol/100 g FFDS) ventricular myocardium. All the electrolyte values found are shown in Table 1 and normal values are presented for comparison in Table 2.

Discussion

It is generally considered that Na-K-ATPase is the receptor for the toxic manifestations of cardiac glycosides. The toxic effects result from inhibition of the enzyme, which leads to an inhibition of the active transport of sodium out of the cell (Repke, 1965; Mason and Braunwald, 1968). This, in turn, causes an intracellular accumulation of sodium and, secondarily, an intracellular loss of potassium and increased inflow of chloride into the cell with an

<table>
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<tr>
<th>Table 1</th>
<th>Muscle electrolytes in a case of fatal digitalis intoxication</th>
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<tbody>
<tr>
<td></td>
<td>mmol/kg intracellular water</td>
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<tr>
<td></td>
<td>Na</td>
</tr>
<tr>
<td>Skeletal muscle</td>
<td>38-0</td>
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<tr>
<td>Left myocardium</td>
<td>167</td>
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<td>Right myocardium</td>
<td>122</td>
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<tr>
<th>Table 2</th>
<th>Reported mean values of muscle electrolytes (in mmol/kg wet weight) in healthy subjects and in patients with cardiac decompensation treated with digitalis (Widdowson and Dickerson, 1964)</th>
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<tbody>
<tr>
<td></td>
<td>No digitalis</td>
</tr>
<tr>
<td></td>
<td>Na</td>
</tr>
<tr>
<td>Skeletal muscle</td>
<td>36-3</td>
</tr>
<tr>
<td>Left myocardium</td>
<td>44-7</td>
</tr>
<tr>
<td>Right myocardium</td>
<td>47-8</td>
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Intracellular electrolytes in fatal digitalis intoxication

Intracellular expansion of cellular volume by the attraction of water (Page, 1964). The findings in the present study are in agreement with this theory.

In the calculation of the intracellular electrolyte concentration the normal value of 87.2 mV for the resting membrane potential (RMP) was used. However, in the patient concerned it is likely that there will have been some changes in the resting membrane potential due to the effect of the digitalis (Rosen et al., 1973), and the repeated DC shocks (Lown et al., 1965; Kleiger and Lown, 1966). Even extreme changes of resting membrane potential, however, amounting to ±25 mV, which is far in excess of what is thought to be compatible with life (Graham et al., 1967), will alter the calculated concentration of the intracellular electrolytes by no more than ±3 per cent, apart from the intracellular sodium concentration of skeletal muscle, which will change by ±13 or -2.5 per cent according to the direction of the change of the RMP. In the physiological range of 87.2 mV ± 10 mV, however, the changes will in all cases be well under 4 per cent. Thus the amount of the total chloride content calculated to be intracellular is relatively insensitive to changes in resting membrane potential, and this does not contribute a significant error in the calculation of extracellular water and hence of the intracellular electrolytes.

DC shocks will affect the cellular membranes and transiently the resting membrane potential, resulting in a loss of intracellular potassium. The change in resting membrane potential per se does not alter the calculated intracellular electrolyte concentrations significantly, but the supposed continuous leakage of potassium from the cells after the DC shocks (Kleiger and Lown, 1966) may have contributed to the extreme electrolyte disturbances recorded. DC shocks enhance the potassium losses induced by digitalis (Lown et al., 1965). In the present case, however, the electrolyte changes depending on digitalis were obviously severe enough to induce ventricular fibrillation, which was treated by DC shocks, probably resulting in a further enhancement of the already existing intracellular potassium deficit.

In the present case the electrolyte disturbances were much more pronounced in cardiac than in skeletal muscle, which may be explained by a higher affinity of digitalis for heart muscle than skeletal muscle or a greater resistance to digitalis in skeletal muscle. Doherty et al. (1967) studied the distribution and concentration of tritium-labelled digoxin in 4 cases and found the highest concentration in the myocardium, kidney, and liver; the level in skeletal muscle was far lower, about 1/8 of the concentration in the myocardium per gram wet weight.

In the present case the intracellular electrolyte pattern in skeletal muscle may be interpreted as the start of a sodium retention in the cell which has not yet resulted in the anticipated secondary increase in the loss of potassium. In the myocardium, however, the changes are already pronounced. This may be the result of the discrepancy in digoxin concentration between skeletal muscle and myocardium reported by Doherty et al. (1967). In the present case the intake of digitalis preceded death by only 6 to 7 hours, which indicates that pronounced intra-
cellular myocardial electrolyte disturbances may develop rapidly. Treatment should, therefore, take the electrolyte disturbances into account with a view to suppressing serious arrhythmias, which are often the immediate cause of death in severe digitalis intoxication.

In view of the inhibition of membrane ATPase by digitalis (Bonting et al., 1962), there seem to be good theoretical reasons for using magnesium in digitalis intoxication, because magnesium stimulates membrane ATPase.

In animal experiments several authors have shown an increased tolerance of digitalis when magnesium was administered at the same time (Zwillinger, 1935; Szekely, 1946; Szekely and Wynne, 1951). In man, however, the effect of magnesium in suppressing digitalis toxicity has so far been demonstrated in only a few cases with moderate overdosage (Lim and Jacob, 1972).

References


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