Correlation between high frequency intravascular ultrasound and histomorphology in human coronary arteries

F Prati, E Arbustini, A Labellarte, B Dal Bello, L Sommariva, M T Mallus, A Pagano, A Boccanelli

Abstract
Objective—To test the efficacy of high frequency intravascular ultrasound (IVUS) transducers in identifying lipid/necrotic pools in atherosclerotic plaques.

Methods—40 MHz transducers were used for in vitro IVUS assessment of 12 arterial segments (10 coronary and two carotid arteries, dissected from five different necropsy cases). IVUS acquisition was performed at 0.5 mm/s after ligation of the branching points to generate a closed system. Lipid/necrotic areas were defined by IVUS as large echolucent intraplaque areas surrounded by tissue with higher echodensity. To obtain histopathological sections corresponding to IVUS cross sections, vessels were divided into consecutive 3 mm long segments using the most distal recorded IVUS image as the starting reference. Samples were then fixed with 10% buffered formalin, processed for histopathological study, serially cut, and stained using the Movat pentacrome method.

Results—122 sections were analysed. Lipid pools were observed by histology in 30 sections (25%). IVUS revealed the presence of lipid pools in 19 of these sections (16%; sensitivity 65%, specificity 95%).

Conclusions—in vitro assessment of lipid/necrotic pools with high frequency transducers was achieved with good accuracy. This opens new perspectives for future IVUS characterisation of atherosclerotic plaques.

(Heart 2001;85:567–570)

Keywords: intracoronary ultrasound; atherosclerosis; plaque morphology

Coronary intravascular ultrasound (IVUS) provides quantitative information on lumen and vessel dimensions and plaque severity, as well as qualitative information on plaque composition in terms of hard and soft components and calcification. Previous IVUS studies on plaque composition, mainly performed in the early 1990s with 20–30 MHz transducers, showed that the technique defines calcification with high sensitivity and specificity, but is less accurate in assessing soft tissue components.1-9

Thus, although 20 and 30 MHz transducers achieved appropriate definition of plaque morphology, the imaging of details such as the lipid pool and the fibrous cap remained poorly defined. No data are available on the characterisation of plaque morphology with high frequency transducers, which should allow more accurate definition of the soft components of the plaques.

In this study we correlated corresponding IVUS and histopathological findings in human arterial specimens obtained at necropsy from patients with atherosclerosis, to determine how accurately 40 MHz IVUS can identify lipid/necrotic pools.

Methods

We performed in vitro IVUS assessments, using continuous pull back, in arterial segments dissected from necropsy hearts. Arterial samples were serially sectioned in relation to IVUS markers. We then correlated the quantitative and qualitative evaluations of lipid/necrotic pools obtained from histopathological slides with those obtained from IVUS cross sections.

SAMPLE SERIES

The pathological series comprised 12 full length arteries, 10 coronary arteries (one left main, four left anterior descending, three circumflex, and two right) and two carotid arteries dissected from five different necropsy cases (all men, age range 52–72 years). All five patients died from acute myocardial infarction.

IVUS INVESTIGATIONS

The in vitro study used an imaging catheter with a 40 MHz mechanical transducer (Sci- med, Boston Scientific Corp, Maple Grove, Minnesota, USA). The catheter transducer (diameter 0.9 mm) provides high resolution cross sectional images.

Arterial segments were dissected from the epicardial fat, and all side branches and distal ends of the artery were ligated to generate a closed system. A 7 French valved sheath was fixed in the proximal end of the vessel segment by an external ring suture. The arterial segments were then suspended horizontally in a container by small metal hooks and placed in a beaker of water. Saline (0.9%) was infused through the lateral arm of the sheath at a constant pressure (60–80 mm Hg) using a syringomanometer.1 Internal markers (calcific deposits) and an external marker (a surgical needle) were used to confirm that ultrasound images and histological sections were aligned.
The IVUS catheter was then inserted into the introducer and advanced within the arterial samples until the external marker was clearly imaged. The IVUS catheter was kept coaxial with respect to the arterial segments and was pulled back at a constant speed of 0.5 mm/s.\(^{10,11}\) The first seconds of the automated pull back were discarded from analysis to avoid non-uniform movement of the catheter in the initial phase of the continuous pull back.

**QUALITATIVE AND QUANTITATIVE IVUS ASSESSMENT OF LIPID/NECROTIC POOLS**

Characteristics of the plaques were defined as follows:
- **Lipid/necrotic areas**—large echolucent areas within the plaque, circumscribed by tissue with higher echodensity.
- **Fibrous tissue**—plaque components having a density similar to or greater than that of the adventitia.
- **Calcific deposits**—highly echogenic segments having a density greater than that of the adventitia and causing acoustic shadowing.

Measurements of lipid/necrotic areas and of the thickness of the fibrous cap were obtained. The latter was defined as the minimum thickness of the tissue layer separating the lipid necrosis pool from the lumen.

**REPRODUCIBILITY OF OBSERVATIONS**

In each cross section, qualitative and quantitative analyses were performed by two independent observers (interobserver variability). Blinded analyses were repeated by the first observer after an interval of at least four weeks (intraobserver variability).

**HISTOPATHOLOGICAL STUDY**

Arterial vessels were cut into consecutive 3 mm long segments from distal to proximal ends using the most distal recorded IVUS image as the starting reference. A coding number was assigned to each segment. A corresponding number had been used to mark the IVUS images. Arterial samples were then fixed with 10\% buffered formalin, routinely processed for histopathological study, and embedded in paraffin. Serial sections from each block were cut at 0.5 mm intervals.

**STATISTICAL ANALYSIS**

All continuous variables are expressed as mean (SD). Continuous data were compared with a paired two tailed Student’s \(t\) test. The results of measurements of lipid pool area and fibrous cap thickness, repeated by the first observer (intraobserver variability) and calculated by two independent observers (interobserver variability), were analysed. In order to assess intra- and interobserver variability, the results were compared using a \(k\) test of concordance for categorical data and correlation analysis for continuous variables.\(^{12}\) A probability value \(p < 0.05\) was considered significant.
Table 1 Reproducibility of the measurements

<table>
<thead>
<tr>
<th></th>
<th>Analysis Ia</th>
<th>Analysis Ib</th>
<th>Analysis II</th>
</tr>
</thead>
<tbody>
<tr>
<td>Area of lipid/necrotic pools (mm²)</td>
<td>0.55 (0.35)</td>
<td>0.45 (0.36)</td>
<td>1.00 (0.48)</td>
</tr>
<tr>
<td>Thickness of fibrous cap (mm)</td>
<td>0.38 (0.14)</td>
<td>0.39 (0.19)</td>
<td>0.36 (0.13)</td>
</tr>
</tbody>
</table>

Intraobserver variability: first (Ia) v second (Ib) observation; interobserver variability: first (Ia) v second (II) observer. *p < 0.05.
CLINICAL IMPLICATIONS

The possibility of characterising atherosclerotic plaques more accurately is of paramount importance. The role of the lipid content of the plaque in the pathogenesis of acute coronary events has not been fully elucidated. In current hypotheses, a large lipid core is one of the markers that define vulnerable plaques—that is, plaques that are prone to rupture and further thrombosis. However, recent studies have shown that vessel thrombosis also occurs in “fibrous” plaques without a lipid core, and in atherosclerotic plaques with thick caps. In these lesions plaque erosion rather than rupture is the local substrate for the thrombosis that causes acute coronary syndromes. Thus better definition of plaque components and lipid necrosis pools may be keys to providing more reliable markers for future studies on the fate of vulnerable plaques. Possibly the in vivo characterisation of plaques with a large lipid core, and their distinction from plaques with prevalent fibrous/proliferative composition, may contribute to identifying potentially vulnerable plaques. Furthermore, proper characterisation of plaque composition in serial IVUS studies intervals could be of help in clinical studies on the effects of drug treatment on plaque size, composition, and possible regression.

TECHNICAL LIMITATIONS

Although our study shows that 40 MHz transducers can identify necrotic lipid pools accurately, there are still some shortcomings in the definition of plaque component. Based on our results, qualitative assessment of lipid pools can be misdiagnosed in the presence of highly echogenic tissue or where the lipid content is distributed diffusely within the atherosclerotic plaque. In the former, highly echogenic tissue—such as calcific or dense fibrous tissue—imparts the identification of lipid pools because acoustic shadowing produces an area of low echogenicity. In the latter, the irregular diffuse distribution hampers the assessment of lipid pools, which can be better visualised if they are well circumscribed in a definite region of the atherosclerotic plaque.

The possibility of visualising and measuring structural details such as lipid/necrotic areas and the thickness of the fibrous cap could provide valuable additional information for the in vivo identification of vulnerable lesions. With this in mind, we attempted to measure the lipid/necrotic pool area and the fibrous cap thickness, but we were unable to obtain reliable measurements in either case, the results being poorly reproducible, both within and between observers.

Further improvements in IVUS technology are required to overcome these limitations and to provide additional morphological information, such as precise measurements of core and fibrous cap thickness. Potential options include IVUS transducers with frequencies higher than 40 MHz, digital processing of IVUS imaging, and radiofrequency analysis of backscatter signals.

A possible limitation of our study is that our information was derived from an in vitro system and therefore plaque assessment might have been facilitated by the absence of backscatter from blood cells. However, previous in vivo studies on plaque characterisation—obtained with transducers at lower frequencies—showed that blood cells can be properly identified and separated from plaque components.

CONCLUSIONS

In vitro assessment of lipid/necrotic pools with high frequency transducers was achieved with good accuracy. This opens new perspectives on the future IVUS characterisation of atherosclerotic plaques.