Preload-adjusted maximal power: a novel index of left ventricular contractility in atrial fibrillation

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Background: Left ventricular contractility in atrial fibrillation is known to change in a beat to beat fashion, because there is no gold standard for contractility indices in atrial fibrillation, especially those measured non-invasively.

Objective: To determine whether the non-invasive index of contractility "preload-adjusted PWRmax" (maximal ventricular power divided by the square of end diastolic volume) can accurately measure left ventricular contractility in a beat to beat fashion in atrial fibrillation.

Methods: Atrial fibrillation was induced experimentally using 60 Hz stimulation of the atrium and maintained in 12 sheep; four received diltiazem, four digoxin, and four no drugs (control). Aortic flow, left ventricular volume, and left ventricular pressure were monitored simultaneously. Preload-adjusted PWRmax, the slope of the end systolic pressure–volume relation (Emax), and the maximum rate of change of left ventricular pressure (dP/dtmax) were calculated in a beat to beat fashion.

Results: Preload-adjusted PWRmax correlated linearly with load independent Emax (p < 0.0001) and curvilinearly with load dependent dP/dtmax (p < 0.0001), which suggested the load independence of preload-adjusted PWRmax. After five minutes of digoxin administration, preload-adjusted PWRmax, dP/dtmax, and Emax fell significantly (p < 0.0001) to 62%, 64%, and 61% of baseline, respectively. Changes were not significant after five minutes of diltiazem (103%, 98%, and 102%) or in controls (97%, 96%, and 95%).

Conclusions: Preload-adjusted PWRmax correlates linearly with Emax and is a useful measure of contractility even in atrial fibrillation. Non-invasive application of this method, in combination with echocardiography and tonometry, may yield important information for optimising the treatment of patients with atrial fibrillation.

Preload-adjusted maximal ventricular power (PWRmax) has recently been introduced as another load insensitive index of left ventricular contractility. Left ventricular power is the product of instantaneous pressure and flow. PWRmax is the peak value during the cardiac cycle, and it reflects the left ventricular contractile state. The preload sensitivity of PWRmax can be reduced by dividing PWRmax by the square of left ventricular end diastolic volume (VEd).

Although promising, preload-adjusted PWRmax has not yet been applied in the setting of atrial fibrillation. We believe that it has great potential in patients with atrial fibrillation because preload-adjusted PWRmax can be calculated non-invasively by combining echocardiography with tonometry. If this index is validated in atrial fibrillation, the application of the method will give us useful and important information about patients with this arrhythmia, including how to optimise rate control treatment and how best to manage tachycardic atrial fibrillation urgently in the operating room or intensive care unit.

Our purpose in this study was to evaluate the use of preload-adjusted PWRmax in atrial fibrillation in an animal model. Because there is no gold standard against which to measure contractility indices in atrial fibrillation, we tried to

Abbreviations: ABD, automated border detection; AIC, Akaike information criteria; BIC, Bayesian information criteria; Emax, slope of ESPVR; ESPVR, end systolic pressure-volume relation; dP/dtmax, maximum rate of change in left ventricular pressure; PWRmax, maximal ventricular power; VEd, end diastolic volume.
accomplish this validation by comparing preload-adjusted PWRmax with dP/dtmax and Emax.

METHODOLOGY

Animal preparation and surgical procedures

This study was approved by the Institutional Animal Care and Use Committee (IACUC) of The Cleveland Clinic Foundation. The Cleveland Clinic Foundation’s IACUC is accredited by the Association for the Assessment and Accreditation of Laboratory Animal Care. The animals received humane care in compliance with the Guide for the care and use of laboratory animals, prepared by the National Academy of Sciences and published by the National Institutes of Health, and institutional guidelines.

Twelve Suffolk blackface sheep (average body weight 60.3 kg) were anaesthetised with an intramuscular injection of ketamine (20 mg/kg) and isoflurane inhalation (2.0–4.0%). The anaesthesia was maintained with 0.5–1.5% isoflurane through a ventilator (Narkomed II, North American Drager, Telford, Pennsylvania, USA). Electrocardiograph leads were attached to the extremities to measure the RR interval. After a right thoracotomy was performed, umbilical tapes were passed around the superior and inferior venae cavae. A Transonic flow probe (model No A16, Transonic Systems, Ithaca, New York, USA) was placed around the ascending aorta. A conductance catheter with two Millar pressure sensors (model SPC-562, Millar Instruments, Houston, Texas, USA) was inserted into the left ventricle to acquire pressure-volume loops and aortic pressure. All haemodynamic data were recorded digitally at a sample rate of 200 Hz using the “PowerLab” data acquisition system (AD Instruments, Mountain View, California, USA).

Study protocol

Steady state haemodynamics (aortic flow, left ventricular volume, left ventricular pressure, and aortic pressure) were measured during normal sinus rhythm. The umbilical tapes around the superior and inferior venae cavae were temporarily occluded (bicaval occlusion) to obtain pressure–volume loops under various preloads. Atrial fibrillation was induced and maintained by stimulating the right atrium at 60 Hz. The baseline measurement after induction of atrial fibrillation was taken over one minute.

Two drugs, diltiazem and digoxin, were used to change contractility because they are commonly used in the urgent management of tachycardic atrial fibrillation.

In the diltiazem group (n = 4), 0.30 mg/kg of diltiazem was given intravenously for two minutes (time 0). A continuous infusion at a rate of 10 mg/h was then started. The haemodynamics were recorded for 30 seconds at 0, 5, 10, 15, 30, and 60 minutes after the initial 0.30 mg/kg load.

In the digoxin group (n = 4), 0.25 mg of digoxin was given intravenously for two minutes (time 0), with an additional 0.25 mg dose at 30 minutes. The haemodynamics were recorded for 30 seconds at 0, 5, 10, 15, 30, and 60 minutes after the initial 0.25 mg/kg load.

To evaluate contractility in the control group (n = 4), the same surgical protocol was followed, and the haemodynamic measurements were taken at the same intervals without any drug administration. All haemodynamic measurements were acquired while the mechanical ventilator was turned off.

Left ventricular contractility analysis

Data analysis was performed using a custom made visual basic program on Excel software (Excel 97 SR-1, Microsoft Corporation, California, USA). All variables described below were calculated in a beat to beat fashion. Left ventricular
volumes—as measured by a conductance catheter—were calibrated to a left ventricular angiograph using a two point calibration based on matching left ventricular end systolic volume and VED in a steady state condition; these two variables were calculated by the area–length method. The dP/dt max was the first derivative of the left ventricular pressure waveform.

We measured left ventricular contractility with E max, as described by Yamaguchi and colleagues. During atrial fibrillation, E max was identified as the slope of the line connecting the predetermined V0 of ESPVR and the upper left corner of each pressure–volume loop in a beat to beat fashion. The V0 of ESPVR was determined on the basis of the bicaval occlusion data in normal sinus rhythm using an iterative linear regression method.

Left ventricular power was calculated instantaneously by multiplying left ventricular pressure by aortic flow. PWR max was determined from the left ventricular power in a beat to beat fashion. Preload-adjusted PWR max was then calculated by dividing PWR max by the square of VED in each beat. We simultaneously measured left ventricular volume using a conductance catheter, which enabled us to calculate the preload adjustment for its non-invasive application, preload-adjusted PWR max was also calculated from aortic pressure.

**Statistical analysis**

Preload-adjusted PWR max was compared with E max and dP/dt max in both a linear random effects model and a non-linear random effects model (preload-adjusted PWR max compared with the square root of E max and dP/dt max) in a beat to beat fashion at baseline. The 12 sheep were considered the random effect. Additionally, preload-adjusted PWR max was compared with dP/dt max using the following more appropriate non-linear model (asymptotic fit):

\[
\text{Preload-adjusted PWR max} = \phi_1 \left(1 - \exp\left[-\exp\left(\phi_2 \times (dP/dt_{\max} - \phi_3)\right)\right]\right)
\]

According to this model, preload-adjusted PWR max increases from zero at dP/dt max = \phi 3 to \phi 1 asymptotically. The parameter \phi 2 controls how rapidly preload-adjusted PWR max approaches its asymptotic value.

Because these two models (square root and asymptotic fit) cannot be compared using standard p values, the Akaike information criteria (AIC) and Bayesian information criteria (BIC) were used to compare these two fits. Smaller values of AIC and BIC indicate better/more parsimonious fits.

Because abortive beats limit the usefulness of preload-adjusted PWR max, beats that had a preload-adjusted PWR max of less than 0.5 (W/cm² × 10⁴) were excluded from these analyses.

To evaluate the three contractility indices after drug administration, the values were averaged in each time point of data comparison. Changes in the three indices in the course of one hour were expressed as a percentage of the averages at baseline. Changes after drug administration in each index were compared using analysis of variance (ANOVA). Differences among the three indices were also compared at each data collection point using ANOVA.

The relation between preload-adjusted PWR max calculated from left ventricular pressure, and aortic pressure was evaluated in a beat to beat fashion, and the Pearson product moment correlation was used.

![Graph](image-url)

**Figure 2** Relations between preload-adjusted PWR max and dP/dt max at baseline. Two different random effects models (square root fit and asymptotic fit) were used for this analysis. Baseline, before drug administration; dP/dt max, maximum rate of change of left ventricular pressure; PWR max, maximal ventricular power.
The relations of the three contractility indices to RR1 and RR2 were also evaluated in a beat to beat fashion. The Pearson product moment correlation coefficient or a linear random effects model was used to evaluate the correlation relations between the preload-adjusted PWRmax and RR1 or RR2. A non-parametric (permutation) test was used to test the changes in this relation after drug administration.

The preload sensitivity of each index was also checked by its relation to VED. This analysis was done using bicaval occlusion data in normal sinus rhythm and baseline atrial fibrillation data before drug administration. A linear random effects model was used to provide the correlation coefficients in the relations between the three contractility indices and VED in a beat to beat fashion.

All data with a probability value of \( p < 0.05 \) were considered significant. The \( \alpha \) level was adjusted using Bonferroni's correction whenever multiple comparisons were done. We used SAS (version 8, SAS/STAT Software, SAS Institute, Cary, North Carolina, USA) and S-PLUS software (version 6.0, Mathsoft, Seattle, Washington, USA) to perform the analyses.

RESULTS

As a result of beat to beat analysis, preload-adjusted PWRmax was shown to correlate with both Emax and \( \frac{dP}{dt}_{\text{max}} \) at baseline in all 12 sheep (\( p < 0.0001 \)). However, the trend of these relations was clearly different for Emax and \( \frac{dP}{dt}_{\text{max}} \). Figure 1 shows the relations between preload-adjusted PWRmax and Emax in each sheep (each data point shows one beat). Linear relations (\( p < 0.0001 \)) without any curvilinear trends (square root fit, \( p = 0.71 \)) were observed in all the animals.

However, in order to obtain a better fit in the relations between preload-adjusted PWRmax and \( \frac{dP}{dt}_{\text{max}} \) (fig 2), it was necessary to employ the non-linear model (square root fit, \( p < 0.0001 \)), which was superior to the linear model (\( p = 0.07 \)). Furthermore, the asymptotic fit model resulted in a better fit than the square root fit (fig 2) using the AIC (3691.9 \( v \) 3815.5) and BIC (3725.1 \( v \) 3844.0). These results indicate that once preload-adjusted PWRmax reaches a particular point, it does not change as \( \frac{dP}{dt}_{\text{max}} \) changes. In addition, the relation deviated to the larger \( \frac{dP}{dt}_{\text{max}} \) side of the linear relation, suggesting that left ventricular contractility was overestimated by \( \frac{dP}{dt}_{\text{max}} \) in the higher contractility beats.

The responses of preload-adjusted PWRmax to the two drugs were similar to those of Emax and \( \frac{dP}{dt}_{\text{max}} \) (fig 3). Five minutes after diltiazem was given, the averages of preload-adjusted PWRmax, \( \frac{dP}{dt}_{\text{max}} \), and Emax fell significantly (\( p < 0.0001 \)) to 62%, 64%, and 61% of baseline, respectively. These decreases were maintained for one hour. These variables did not change much in the digoxin group five minutes after drug administration (103%, 98%, and 102%, respectively) or in the control group (97%, 96%, and 95%), and the values gradually decreased in the course of one hour, reaching significance only in the control group at 60 minutes (\( p < 0.001 \)).

Per cent changes among the three indices were not significantly different at any of the data collection time points.

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Per cent changes among the three indices were not significantly different at any of the data collection time points.
Although dP/dtmax tended to be higher than the other indices from 10 minutes after diltiazem administration, these changes were not significant.

The relation between preload-adjusted PWRmax and RR1 showed a positive correlation (p < 0.0001 by fitting the random effect model). Although this relation changed very little in the digoxin and control groups, it flattened out dramatically in the diltiazem group. The representative change is shown in fig 4. Similar relations with RR1 and similar changes in dP/dtmax and Emax were observed (data not shown). The relation between preload-adjusted PWRmax and RR2 was scattered at baseline (fig 5A). However, after excluding the beats that had an RR1 of less than 350 ms, the weak negative relation (average correlation coefficient = −0.407) appeared (fig 5B). In addition, this relation flattened out after diltiazem administration (fig 5A). Similar phenomena were observed in dP/dtmax and Emax (data not shown). These changes were significant (p < 0.006), suggesting that contractility and its dependence on RR intervals changed in the diltiazem group.

The preload insensitivity of preload-adjusted PWRmax and Emax was shown, in contrast to the preload sensitivity of dP/dtmax with normal sinus rhythm (data not shown). However, during atrial fibrillation, all indices—including the preload-adjusted PWRmax and Emax—were higher with a higher VED (fig 6). A linear random effects model provided the R values for dP/dtmax (0.90), Emax (0.90), and preload-adjusted PWRmax (0.81). This indicated that during atrial fibrillation contractility was high when VED was high.

**DISCUSSION**

Our main observation was that, during atrial fibrillation, preload-adjusted PWRmax correlated linearly with the load independent variable (Emax) and curvilinearly with the load dependent variable (dP/dtmax). These results support the use of preload-adjusted PWRmax as a load independent index of left ventricular contractility in atrial fibrillation.

The great advantage of preload-adjusted PWRmax compared with the other two indices is that the method can be used non-invasively. Measurement of dP/dtmax usually requires insertion of a Millar catheter into the left ventricle unless enough mitral regurgitation is observed by echocardiography. Measuring Emax is more complicated because it requires a conductance catheter system. It may even be impossible to measure because of the need to determine V0 in normal sinus rhythm. On the other hand, preload-adjusted PWRmax can be measured using the echocardiographic

**Figure 5** (A) Representative relation between preload-adjusted PWRmax and RR2 and the change after diltiazem administration. There was no trend in this relation at baseline, which changed significantly (p < 0.006) to flat after diltiazem administration. (B) The same relation at baseline after excluding beats that had an RR1 of less than 350 ms. There was a negative trend after this arrangement. Baseline, before diltiazem administration; PWRmax, maximal ventricular power; RR1, preceding RR interval; RR2, pre-preceding RR interval; 5 min after diltiazem, five minutes after the completion of the initial diltiazem load (0.30 mg/kg).

Although dP/dtmax tended to be higher than the other indices from 10 minutes after diltiazem administration, these changes were not significant.

The relation between preload-adjusted PWRmax, calculated from left ventricular pressure and aortic pressure was closely linear in all the animals in a beat to beat fashion. The Pearson product moment correlation coefficient was 0.985 ± 0.011.

**Figure 6** Relations between VED and dP/dtmax, preload-adjusted PWRmax, and Emax during atrial fibrillation before drug administration in all 12 sheep. Emax, the slope of end systolic pressure–volume relation; dP/dtmax, maximum rate of change of left ventricular pressure; PWRmax, maximal ventricular power; VED, end diastolic left ventricular volume.
Study limitations
Because the average heart rate fell significantly after diltiazem administration, the effects of heart rate cannot be neglected in this study. However, as seen in fig 4, the beats of shorter RR intervals five minutes after diltiazem administration showed lower preload-adjusted PWRmax values than those of corresponding intervals at baseline, which supports our contention that preload-adjusted PWRmax can be used to estimate appropriate contractility in spite of the rate changes. Kass and Beyar suggested that multiplying preload-adjusted PWRmax by the length of the cardiac cycle (in seconds) can help normalise for pure rate effects. Although we attempted to perform this correction in our data in a beat to beat fashion, the relation between preload-adjusted PWRmax and dP/dtmax or Emax became scattered after this correction (data not shown). The strong regulation of contractility by the RR1 and RR2 intervals may have made this beat to beat correction impracticable in the setting of atrial fibrillation. Further investigation is needed to determine whether this correction is necessary in atrial fibrillation.

Because we did not use autonomic blockade in our study, the effects of the sympathetic and parasympathetic nervous system on measurements of preload-adjusted PWRmax or Emax cannot be eliminated. However, we did not induce autonomic blockade because we wanted the study conditions to correspond closely with those in the clinical setting.

Although preload-adjusted PWRmax was closely related to both Emax and dP/dtmax, it revealed values close to zero when abortive beats occurred. This phenomenon worsened these relations. With abortive beats, preload-adjusted PWRmax falls because aortic flow is close to zero. However, cardiac contractility still exists, as expressed in dP/dtmax or Emax. Mandarino and colleagues voiced similar concerns in the steady state condition, stating that severe hypovolaemia or notably raised arterial pressure may make preload-adjusted PWRmax less reliable. Although we excluded the beats that had a preload-adjusted PWRmax of less than 0.5 W/ml-10\(^{-7}\) in the analysis in beat to beat fashion, and included all the beats for analysis in averages, the optimum handling of the abortive beats remains to be elucidated.

Finally, complete non-invasive application of preload-adjusted PWRmax using ABD and tonometry needs to be validated in the next stage study.

Conclusions
Preload-adjusted PWRmax correlated linearly with Emax and seems to be a useful measure of contractility in atrial fibrillation. In addition, we believe that this index is a potentially useful non-invasive measure for patients with atrial fibrillation. The application of the method in this setting may yield important information about how to optimise treatment for patients with both acute and chronic atrial fibrillation.

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REFERENCES


IMAGES IN CARDIOLOGY

Computed tomography of the aortic valve

A 72 year old lady underwent multislice computed tomography of the thorax in order to assess the ascending aorta before aortic valve replacement for calcific aortic stenosis. In order to improve image quality the study was prospectively triggered to mid-to-late diastole (heart rate 72 beats/min, 273 ms scan window timed to complete at 85% of the cardiac cycle). A gantry rotation time of 0.5 s, and acquisition of 4 × 2.5 mm slices gave excellent image resolution of the ascending aorta. By removal of surrounding non-calculated structures, using volume rendering techniques, striking “incidental” three dimensional images of the heavily calcified aortic valve were reconstructed. There is increasing interest in quantification of aortic valve calcification. Computed tomography is unique among imaging modalities in its ability to quantify calcification accurately, and traditional “scoring” techniques based on the assessment of coronary artery calcification have been modified to assess aortic valve calcification.

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