and that PPARα is an essential regulator of metabolism in hypoxia. PPARα−/− mice (n=18) and wild-type (wt) controls (n=20) were exposed to 3 weeks of normobaric hypoxia. Control treated mice were kept at 60 mm Hg PO2 and 83% ± 0.4% oxygen saturation, while normoxic mice were kept at 80 mm Hg PO2 and 100% oxygen saturation. Cardiac output was measured using pulsed Doppler echocardiography. A number of endpoints were assessed both under growing conditions (0.261 ± 0.03 vs 0.27 ± 0.05; p<0.05). While CSF-1 stimulation increased the spread area in wt cells above that of wt cells (0.261 ± 0.03 vs 0.28 ± 0.06; p<0.05), PPARα−/− mice (n=20) were housed in a room with 2% oxygen. In-vivo cardiac function was measured using multislice cardiac magnetic resonance imaging. Hearts were perfused in the Langendorff mode to measure palmitate oxidation and glycolysis using 3H-labelled substrates. Cardiac output was unchanged in hypoxic wt and normoxic PPARα−/− mice, but was reduced by 31% by hypoxia in PPARα−/− mice (p<0.02). Late-stage ventricular filling was 46% lower in hypoxic PPARα−/− mice (p<0.01). Hypoxia reduced palmitate oxidation by 27% in wt hearts, but did not affect PPARα−/− hearts. Hypoxia increased net lactate efflux 2.4-fold in wt hearts from wt animals (p<0.01), but lactate efflux from PPARα−/− hearts was unchanged with hypoxia. Hypoxia increased basal glycolytic flux 2.4-fold in wt hearts but did not alter the fluxes in PPARα−/− mice (p<0.01), which was already 3.7-fold greater than wt hearts. Thus PPARα−/− hearts lack the metabolic flexibility essential for adaptation to chronic hypoxia, and their inability to upregulate glycolysis probably impairs cardiac function.

**Conclusions**

Nox2−/− BMM display marked abnormalities in morphological and migratory behaviour that may contribute significantly to the ability of the monocyte to differentiate and migrate in vivo in response to pathological stimuli. This phenotype could underlie the protection against fibrosis observed in vivo in Nox2−/− mice.

### D06 NORMOBARIC HYPOXIA IMPAIRS CARDIAC ENERGETICS IN NORMAL HUMAN VOLUNTEERS

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**Background**

The in first few days of hypoxia exposure, left ventricular dysfunction is consistently observed in the human heart, yet the cellular mechanisms underlying the dysfunction are poorly understood.

**Objective**

Our hypothesis was that normobaric hypoxia impairs cardiac energetics, leading to cardiac dysfunction in healthy subjects.

**Methods**

Normal healthy volunteers underwent 20 h of normobaric hypoxia in purpose-built hypoxia chambers. The partial pressure of oxygen during end tidal expiration (PETo2) was kept between 50 and 60 mm Hg, while keeping peripheral oxygen saturation (SpO2) above 80%. Cardiac function was measured using magnetic resonance imaging and echocardiography. High-energy phosphate metabolism was measured as the ratio of phosphocreatine to ATP (PCr/ATP) by 31P magnetic resonance spectroscopy before and after 20 h of hypoxia. Healthy men (n=12, aged 24±2 years) were recruited from the University of Oxford.

**Results**

During hypoxia, PETO2 and SpO2 averaged 55±1 mm Hg and 83.6±0.4%, respectively. There was a 15% reduction in cardiac PCR/ATP, from 2.0±0.1 to 1.7±0.1 after hypoxia (p<0.01) and reduced diastolic function, measured as E/E′, from 6.1±0.4 to 7.5±0.7, p<0.01.

**Conclusion**

Short-term normobaric hypoxia led to rapid changes in cardiac metabolism and alterations in diastolic function in normal human hearts. Impaired high-energy phosphate metabolism may explain the cardiac dysfunction observed after hypoxic exposure, whether in health or disease.

### D07 ROLES OF P47PHOX S303/S304 PHOSPHORYLATION IN TNFα-INDUCED ENDOTHELIAL REACTIVE OXYGEN SPECIES PRODUCTION AND MITOGEN-ACTIVATED PROTEIN KINASE ACTIVATION

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Endothelial cells express constitutively a Nox2 oxidase, which by generating reactive oxygen species (ROS) plays an important role in TNFα signalling. The Nox2 has at least four regulatory subunits and p47phox is a major regulatory subunit of this enzyme. It has been reported that phosphorylation of double serines (S303/S304) in p47phox is a key step for Nox2 activation. In this study, we investigated the role of p47phox S303/S304 phosphorylation in TNFα-induced ROS production and mitogen-activated protein kinase (MAPK) activation in endothelial cells. Serines S303/304 (human p47phox cDNA) were replaced to alanines by site-directed mutagenesis and the wild-type and mutated p47phox were used to transfect a mouse microvascular endothelial cell line (SVEC4-10). Forty-eight hours after transfection, cells were stimulated with or